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# **RESEARCH PAPER**

# Green house evaluation of IAA and GA<sub>3</sub> producing yeast isolates on growth tomato crop by root dip method

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**Abstract :** A green house experiment was conduct by use of IAA (MZL -8 and TCL -1) and GA<sub>3</sub> (CAL – 1 and ACL - 3) producing yeast isolates on growth of tomato crop by root dip method with 8 treatments and 3 replication at Department of Agricultural Microbiology, GKVK, Bengaluru. The yeastisolate TCL -1 recorded maximum plant height, number of leaves per plant, number of branches per plant, root length, fresh shoot biomass, dry shoot biomass, fresh root biomass, dry root biomass, IAA and GA<sub>3</sub> content, 44.73 cm, 80, 11.40, 14.70 cm, 24.00 g/plant, 11.75 g/plant, 7.98 g/plant, 3.91 g/plant, 1.205  $\mu$ g/g of leaf and 0.550  $\mu$ g/g of leaf respectively. The control (T<sub>1</sub>) recorded Lowest plant height, number of leaves per plant, number of branches per plant, root length, fresh shoot biomass, dry root biomass, IAA and GA<sub>3</sub> content, 22.20 cm, 55.70, 8.00,9.00 cm, 11.00 g/plant, 6.23 g/plant, 3.67 g/plant, 2.07 g/plant, 0.384  $\mu$ g/g of leaf and 0.200  $\mu$ g/g of leaf respectively at 50 DAT.

Key Words : Indole -3-acetic acid (IAA), Gibberellins (GA<sub>3</sub>), *Manilkara zapota* leaf isolate (MZL), *Coffea robusta* leaf isolate (CAL), *Theobroma cocoa* leaf isolate (TCL)

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# INTRODUCTION

Yeasts are group of fungi in which unicellular form are predominant in environment. Yeasts are generally of three types a) Ascomycetous b) Basidiomycetous and c) Deuteromycetous. Most of the yeasts are represented in Sub Division Ascomycotina and Basidiomycotina of the kingdom Mycota. Dimorphic yeasts are also present which become filamentous under certain environmental conditions. They play their role in the dynamics of biological and chemical turnover in soil, plants, animals and water. They are generally present in the natural resources leaf surface, fruit surface soil, animal surfaces and in the intestinal tracts of warm-blooded animals (Rose and Harrison, 1987).

Plant hormones regulate the cellular and physiological processes such as cell division, cell enlargement, bud dormancy, flowering, fruit ripening, seed dormancy, seed germination and leaf abscission. Auxin, one of the plant hormones stimulates differentiation of phloem and xylem, root initiation on stem cutting and also the development of branch roots. Auxin mediates the tropism (bending), response to gravity and light. Indole -3-acetic acid (IAA) is the common natural auxin that

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shows all auxin do actions and extensively affects plants physiology.

The indole-3-acetic acid (IAA) stimulates rapid and long-term responses in plants and has been identified in plant-associated bacteria, molds and yeasts. The role of microbial IAA in plant-microbe interactions has recently received increased attention. Therefore IAA-producing microbes have been suggested as sources of biofertilizer. The IAA is a signaling molecule in certain microorganisms and modifies gene expression. Therefore, IAA might act as a reciprocal signaling molecule in microbeplant interactions (Pei *et al.*, 2014).

Gibberellins are an important group of isoprenoid phytohormones that occur in small amounts in higher plants. They are involved in the development and regulation of different growth processes throughout the life-cycle of plants. Numerous reports have shown gibberellins (GA<sub>3</sub>) play a role in the growth, development and sexual differentiation processes of different plant species.

# MATERIAL AND METHODS

Experiments were conducted by collected yeast isolates from the Department of Agricultural Microbiology, University of Agricultural Sciences, GKVK, Bengaluru to study the effect of yeast isolates on tomato crop under green house condition.

# Green house evaluation of efficient yeast isolates for plant growth promotion in tomato crop:

Pot culture experiment :

The Abhinava hybrid seedlings were transplanted to sterilized soil filled in bags and mentioned for experiment under green house condition.

# Method of application of yeast isolates:

Root dip:

The yeast isolates were grown in YM broth with constant shaking at 150 rpm for 48 hr at room temperature. Tomato seedling roots were dipped (10 minutes) in different yeast cultures grown in broth and transplanted to the bags.

## **Treatments:**

 $T_1$ - Control,  $T_2$ - MZL – 8,  $T_3$ - TCL- 1,  $T_4$ - CAL – 1,  $T_5$ - ACL – 3,  $T_6$ - IAA (1 ppm),  $GA_3\,$  (1 ppm) and  $T_8$ - YM Broth.

#### **Observations recorded:**

The Plant height (cm), Number of branchesand Number of leaves were recorded at 25 and 50 days after transplanting.

## Total biomass content:

The fresh and dry weight of shoot and root (g) were recorded at50 days after transplanting and expressed in grams per plant.

## **Plant analysis:**

*Estimation of IAA and GA in tomato leaf sample Leaf sample extraction :* 

The extraction and purification was made following the method of Kettner and Doerffling (1995).

## Estimation of indole-3-acetic acid (IAA):

Ethyl acetate (15ml) fraction was taken and 4 ml of Salper's reagent was added and incubated in darkness for 1 hr at 28°C (Gordon and Weber., 1951). The intensity of the pink colour developed was read in spectrophotometer at 535 nm. By referring to a standard graph prepared with chemical grade indole-3-acetic acid, the quantity of IAA in the sample was determined.

## Estimation of gibberellic acid (GA):

Ethyl acetate (15ml) fraction was taken and 2 ml of zinc acetate solution was added (Paleg, 1965). After 2 min, 2 ml of Potassium ferrocyanide solution was added and the mixture was centrifuged at 10,000 rpm for 10 min. Five ml of supernatant was added to 5 ml of 30 % hydrochloric acid (HCl) and the mixture was incubated at 20 °C for 75 min. The blank was prepared with 5% HCl. The absorbance was measured at 254 nm in a spectrophotometer. By referring to a standard graph prepared with chemical grade of  $GA_3$ , the quantity of  $GA_3$  in the sample was determined.

# **RESULTS AND DISCUSSION**

The data pertaining to plant height, number of leaves and number of branches at 25 and 50 DAT is given in the Table 1.At 25 DAT significantly higher plant height, number of leaves and number of branches were recorded by the treatment  $T_3$  (TCL – 1) 24.50 cm, 53.30 and 8.90 respectively and lowest was recorded by treatment  $T_1$ (control) 16.40 cm, 40.30 and 6.70, respectively. At 50 significantly higher plant height, number of leaves and number of branches were recorded by the treatment  $T_3$  (TCL - 1) 44.73 cm, 80.00 and 11.40, respectively and lowest was recorded by treatment T<sub>1</sub> (control) 22.20 cm, 55.70, and 8.00, respectively.

Such enhancement effect of yeast isolates Root dip might be attributed to the favorable influence on metabolism and biological activity and their stimulatory effect on photosynthetic pigments, leaf area, cell turgor and enzyme activity which helped for vegetative growth of plants (Mady, 2009, Abou and Mady, 2012).

These results are were also conformation with findings of Mona *et al.* (2013) They have reported that application of yeast extract increased the The Plant height (cm), Number of branches and Number of leaves in

soybean plants (Mekki, and Amal, 2005). Similer results were obtained by Abdel *et al.* (2012) in Snap bean crop.

The data pertaining to root length, fresh shoot, root and dry shoot, root biomass at 50 DAT is given in the Table 2.The treatment  $T_3$  (TCL-1) was recorded maximum root length, fresh shoot biomass, dry shoot biomass, fresh root biomass and dry root biomass, 14.70 cm, 24.00 g/plant, 11.75 g/plant, 7.98 g/plant and 3.91 g/ plant respectively, the lowest were recorded by control treatment ( $T_1$ ) 9.00 cm, 11.00 g/plant, 6.23 g/plant, 3.67 g/plant and 2.07 g/plant, respectively at 50 DAT.

The significant increase in root length, fresh shoot, root and dry shoot, root biomass at 50 DAT was observed



Tuantumenta	Plant height (Cm)		Number of leaves at 25 DAT		Number of branches at 25 DAT	
meatiments	25 DAT	50 DAT	25 DAT	25 DA T	25 DAT	25 DAT
T <sub>1</sub> - Control	16.40	22.20	40.30	55.70	6.70	8.00
T <sub>2</sub> - MZL - 8	23.50	43.20	52.30	74.70	8.70	10.70
T <sub>3</sub> - TCL- 1	24.50	44.73	53.30	80.00	8.90	11.40
T <sub>4</sub> - CAL – 1	22.30	40.50	47.30	68.00	7.90	9.70
T <sub>5</sub> - ACL - 3	21.30	38.27	46.00	66.00	7.70	9.40
T <sub>6</sub> - IAA (1 pp m)	22.10	41.25	49.00	71.70	8.20	10.20
T <sub>7</sub> - GA <sub>3</sub> (1 ppm)	22.50	42.28	50.30	72.30	8.40	10.30
T <sub>8</sub> - YM Broth	19.50	24.20	41.00	62.70	6.80	9.00
Sem $\pm$	1.15	2.25	3.78	3.25	1.10	1.32
CD @ 5%	4.76	6.10	8.95	8.62	2.86	3.21

Note: MZL - 8 = Manilkara zapota leaf isolate - 8, TCL - 1= Theobroma cocoa leaf isolate - 1

 $CAL - 1 = Coffea \ robusta \ leaf \ isolate - 1, \ ACL - 3 = Averrhoa \ carambola \ leaf \ isolate - 3$ 

IAA = Indole acetic acid, GA<sub>3</sub>= Gibberellic acid, YM Broth = Yeast Extract Malt Extract Broth

Table 2 : Effect of IAA and GA <sub>3</sub> producing yeast isolates on tomato crop root length, fresh shoot biomass, dry shoot biomass, fresh root biomass	
and dry root biomass under pot condition at 50 DAT by root dip method	

v	1	F 1 1 (1)	D 1 (	E 1 (1)	D (1)	
Treatments	Root length (cm)	(g/plant)	biomass (g/plant)	(g/plant)	(g/plant)	
T <sub>1</sub> - Control	9.00	11.00	6.23	3.67	2.07	
T <sub>2</sub> - MZL - 8	12.30	23.98	11.51	7.75	3.62	
T <sub>3</sub> - TCL- 1	14.70	24.00	11.75	7.98	3.91	
T <sub>4</sub> - CAL – 1	10.30	23.00	11.00	7.62	3.67	
T <sub>5</sub> - ACL - 3	12.00	22.75	10.25	7.58	3.42	
T <sub>6</sub> - IAA (1 ppm)	12.70	16.75	10.50	5.58	3.50	
T <sub>7</sub> - GA <sub>3</sub> (1 ppm)	11.70	23.25	10.98	7.48	3.31	
T <sub>8</sub> - YM Broth	9.70	11.50	6.50	3.83	2.17	
Sem $\pm$	0.86	2.21	2.24	0.94	0.56	
CD @ 5%	2.56	6.10	6.23	2.56	1.64	

Note: MZL - 8 = Manilkara zapota leaf isolate - 8, TCL - 1= Theobroma cocoa leaf isolate - 1

 $CAL - 1 = Coffea \ robusta \ leaf \ isolate - 1, \ ACL - 3 = Averrhoa \ carambola \ leaf \ isolate - 3$ 

 $IAA = Indole \ acetic \ acid \ , GA_3 = \ Gibberellic \ acid \ , YM \ Broth = Yeast \ Extract \ Malt \ Extract \ Broth$ 

due to inoculation of yeast isolates to tomato crop at 50 DAT. These results are in conformation with the findings of Ramadan *et al.* (2013). They reported that application of yeast as a biofertilizer increased the root length fresh shoot, root and dry shoot, root biomass at 50 DATin Sugar beet.

The data pertaining to IAA and GA<sub>3</sub> content at 50 DAT is given in the Table 3. The significantly maximum IAA content in tomato leaf sample was recorded in plants roots dipped with the yeast isolate TCL – 1 (1.205  $\mu$ g/g of leaf) followed by plants roots dipped with MZL-8 (1.123  $\mu$ g/g of leaf). The minimum IAA content in tomato leaf sample was recorded in control plants (0.384  $\mu$ g/g of leaf).

Table 3 : Effect of IAA and GA <sub>3</sub> producing yeast isolates on tomato							
crop leaf IAA and GA3 content in leaf under pot condition at 50 DAT by root din method							
Treatments	IAA content (µg/g of leaf)	GA <sub>3</sub> content (µg/g of leaf)					
T <sub>1</sub> - Control	0.384	0.200					
$T_2$ - MZL – 8	1.123°	0.500					
T <sub>3</sub> - TCL- 1	1.205	0.550					
$T_4$ - $CAL - 1$	0.984	0.320					
T <sub>5</sub> - ACL - 3	0.783	0.450					
T <sub>6</sub> - IAA (1 ppm)	0.834	0.420					
T <sub>7</sub> - GA <sub>3</sub> (1 ppm)	0.543	0.480					
T <sub>8</sub> - YM Broth	0.432	0.250					
T <sub>1</sub> - Control	0.384	0.200					
Sem $\pm$	0.242	0.452					
CD @ 5%	0.620	1.602					

Note: MZL - 8 = Manilkara zapota leaf isolate - 8, TCL -

1= Theobroma cocoa leaf isolate – 1

CAL - 1 = Coffea robusta leaf isolate - 1, ACL - 2 = 4 userbase cargenbala leaf isolate - 2

 $3 = Averrhoa \ carambola \ leaf \ isolate - 3$ IAA = Indole acetic acid, GA<sub>3</sub>= Gibberellic acid, YM

Broth = Yeast Extract Malt Extract Broth

Significantly higher GA<sub>3</sub> content in tomato leaf was noticed in plants inoculated with the yeast isolates compared to control plants. The plants roots dipped with the yeast isolate TCL – 1 was showed significantly higher GA<sub>3</sub> content (0.550  $\mu$ g/g of leaf) in tomato leaf which was on par with plants roots dipped with MZL-8. Significantly lower GA<sub>3</sub> content (0.200  $\mu$ g/g of leaf) in tomato leaf sample was recorded in control plants.

This may be due to the production of IAA and GA<sub>3</sub> by inoculated yeast isolates which may promote various

physiological activities in plant which are considered to be indispensable for proper growth and development of plants which is in close agreement with the findings of Salwa (2013) who reported that application of yeast increased IAA and  $GA_3$  content in the shoot of field bean.

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