

RESEARCH PAPER

Detection of polymorphism of ATM gene in Leukemia

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The Ataxia telangiectasia mutated gene encodes the ATM protein, a key element in the DNA damage response (DDR) signalling pathway responsible for maintaining genomic integrity within the cell. In order to study the genetic changes of ATM gene in Leukemia, we collected the blood samples of healthy persons for control and leukaemia persons for disease samples. We isolated DNA and performed PCR to analyse by Gene sequencing. As a result we found less number of sequencing result in nucleotide BLAST due to which we are unable to detect the compared result of mutation. Larger and/or combined association studies are needed to conclude the result.

Key words : ATM gene, Gel electrophoresis, Polymerase chain reaction, DNA sequencing

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INTRODUCTION

The Ataxia-telangiectasia-mutated (ATM) gene in humans was identified as the basis of a rare autosomal disorder leading to cancer susceptibility and is now well known as an important signal transducer in response to DNA damage (Johnson *et al.*, 2007). ATM gene is large, spanning about 150 kbs of genomic DNA, and is located on chromosome 11q22-23. It is expressed in many tissues throughout the body and has a key role in the DNA repair response and in conducting cell cycle arrest and apoptosis (Shiloh, 2003). The loss of ATM function leads to genome instability and an increased risk for cancer, neurodegeneration, and impaired glucose tolerance (Kastan and Bartek, 2004). MicroRNAs play critical roles in development, differentiation and disease progression and are predicted to target more than 60 per cent of mRNAs in humans (Brennan *et al.*, 2012).

RESEARCH METHODOLOGY

Sample collection:

48 healthy blood samples were collected from CytoGene students, relatives and friends and 48 disease samples were collected from different hospitals by the rules and guidelines of W.H.O.

Isolation of DNA from blood sample :

DNA from blood sample were isolated by DNA isolation kit.

RESEARCH FINDINGS AND ANALYSIS

Various blood samples were collected from healthy and leukemia infected persons. These samples were placed in EDTA tubes as to prevent it from coagulating. These samples were then used to extract DNA.

DNA extraction (Gel electrophoresis) :

Fig. 1 and 2 showing isolated DNA that was run on



Fig. 1: Blood sample were collected in EDTA tubes

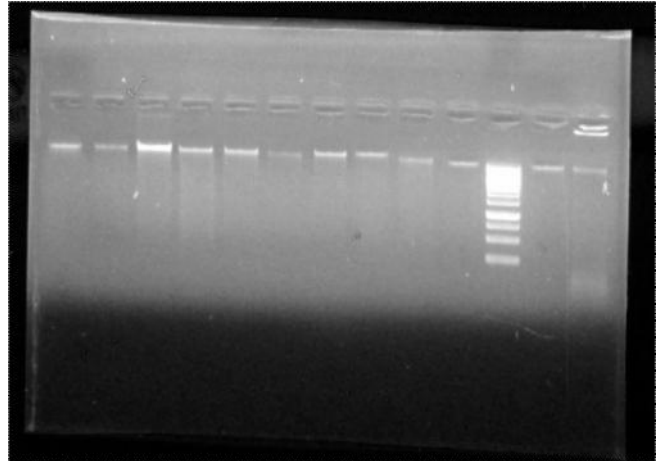


Fig. 4: Showing ladder in lane 1 and disease samples 1, 2, 3 and 4 in lane 2, 3, 4 and 5, respectively

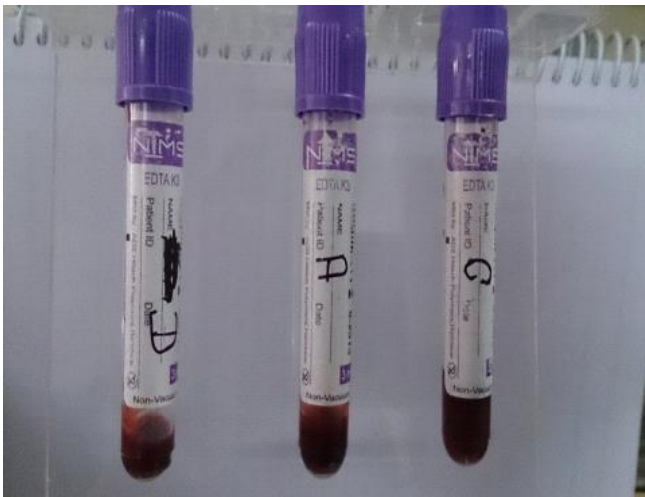


Fig. 2: Blood samples obtained from patients

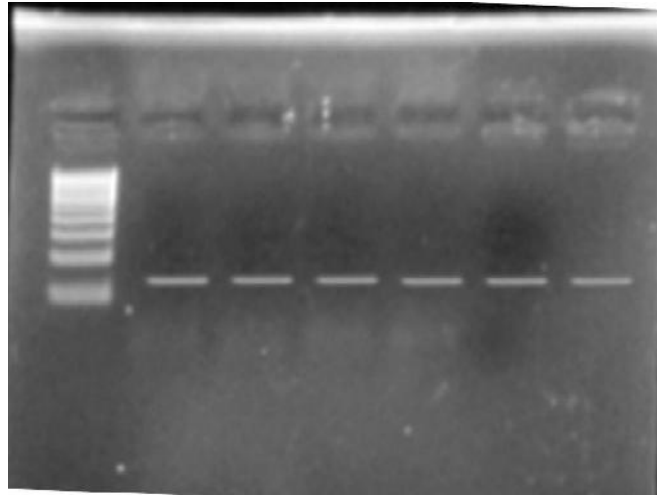


Fig. 5: Shows ladder in lane 1, and control samples in lane 2, 3, 4 and 5, respectively

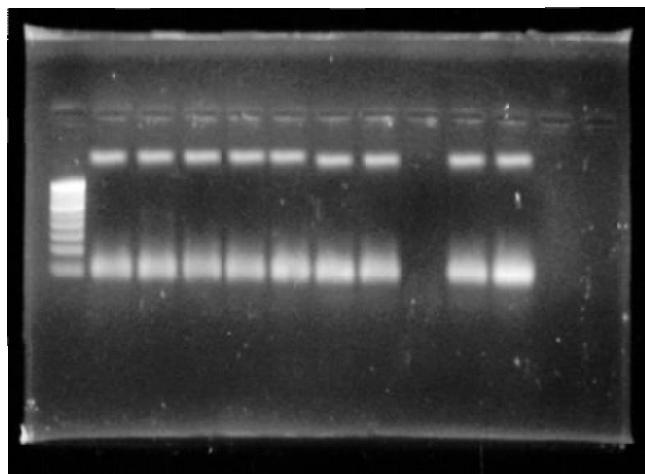


Fig. 3: Showing ladder in lane 1, and control samples in lane 2, 3, 4 and 5, respectively

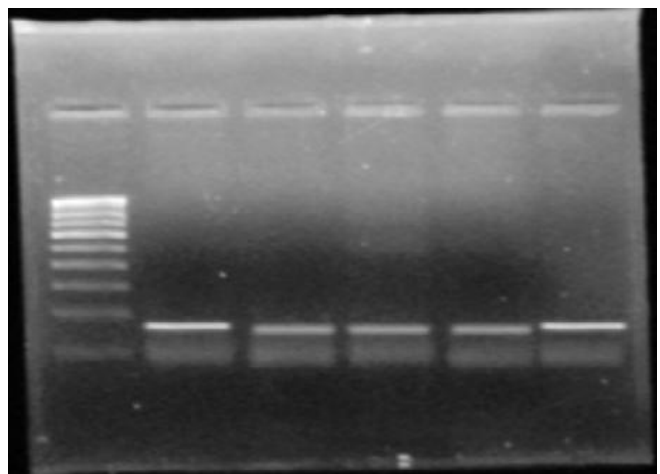


Fig. 6: Shows ladder in lane 1 and disease samples in lane 2, 3, 4 and 5, respectively

a 0.8 per cent agarose gel at 100 V for 15 mins. The gel was stained with ethidium bromide and then visualized in a gel documentation system.

PCR :

Fig. 5 and 6 showing amplified DNA that was run on a 1.2 per cent agarose gel at 75V for 20 mins. The gel was stained with ethidium bromide and then visualized in a gel documentation system.

Sequencing result (Appendix 1) :

This is the sequencing result of ATM gene obtained by SANGER and SANGER METHOD.

After getting this result we did pairwise sequence alignment by EMBL-EBI and got which place is mutated.

ATM GENE control sequence

```
ATGGATGCTCATACGCTCGAGAGCTCTCCG
AGAGATCCTCTCGAAGGTTTGTCTAGAAAT
AGCCTGGTTATCAGAAATTGTCCCAAGCAAT
AGATTACGACTGATGAAACAACATCGCACAT
GGGTTGAATAATTCTGAAAATGTAAGAAAG
AGAAATCTTCACATTTCAATGTTCTTCTTGT
TAGATAAAACAAAATTGTTTCATGAATTATTAG
TGACACATGAGATAGACACATCGTCTCAATTGG
TATGATAAATGTTATTTAAACGAAGAACCGAAT
TCTTCACCAGACATATAGCATCATCCTAAATTAG
CGCTATGCTTACAACGTTTAGGTGCTAATCGA
CTGTTTAAACAAGTTCAGTTATAGCAATAATGA
ATCAATAGACTTAAACAGTTATCGGTCAGTGTT
GCGATCAGCTGCTGCATGGCGTCTATACGCG
CCGTAAAGCCGATTGGATAATAAATCCCCTGA
TAGATTCCATTTAAATCGTTAATTTGTCTTCC
TACTGATTGGAAAATGTGTGGTTTAGAAACTT
CATTTTACTGGCTTTTACGAGCTGCTTACAATC
AAACTGGTTTAGAGTATCAGAAATAGTTGCT
TCTCAAAGTGAACCTTCTTATTGAAGAGCTGTA
TTCTGATACTGTACAATTCATCTCTTTAGCGC
GCTAGAGCCCCTTGGGCTGCTGATCTCTCCTA
CATGCGAATCAGGAGCAATGTTTACAATTGA
CATTAACCTTACTTTGTCATGCATTAATTTCTA
TGAATTTCTCATCTGCTACAATCCCCTAGCTCGT
ATCTTACTCATGATCAATTAACCTTCAATCTAC
ATTGGATTCCATTGAACCTTCTTACATTGTCA
ACTAGTTTTATTGATGTAGTATTATCAAAAAA
TATTTTTCTAATCATGTTCAATGTCAATTAAG
AAGTTTCTTAATTCATACTTACCTTTATTTTGC
GGAAACTGCAACAACAAGCACAAATTAATTT
GCAACCGTCGG
```

A TM GENE disease sequence :

```
ATGGATGCTCATACGCTCGAGAGCTCTCCG
AGAGATCCTCTCGAAGGTTTGTCTAGAAATTA
GCCTGGTTATCAGAAATTGTCCCAATGAATAGA
TTACGACTGATGAAACAATTCATCGCACATG
GGTTGAATAATTCTGAAAATGTAAGAAAG
GAAATCTTCACATTTCAATGTTCTTCTTGTGTA
GATAAAACAAAATTGTTTCATGAATTATTAGT
GACACATGAGATAGACAGATCGTCTCAATTG
CTATGATAAATGTTATTTAAACGAAGAACCGA
ATTCTTCACCAGACAATAGTGTACATCCTAAAT
TAGCGCTATGCTTACAACGTTTAGGTGCTA
ATCGACTGTTTAAACAAGTTCAGTTATAGCAATA
ATGAATCAATAGACTTAAACAGTTATCGGTCAGT
CGGATATAACATTAAGAGTTGCGATCAGC
TGCTGCATGGCGTCTATACGCGCCGTAAAGCCG
ATTGGATAATAAATCAAATGATATACCTCATTTA
AATCGTTAATTTGTCTTCTTACTGATTGGAAA
TGTGTGGTTTAGAAACTTCAATTTACTGGCTT
TTACGAGCTGCTTACAATCAAACCTGGTTTAGA
GTATCAGAAATAGTTGCTTCTCAAAGTGAAC
TCTTATTGAAGAGCTGTATTCTGATACTGTACA
ATTCATCTCTTTAGCGCGCTAGAGCCCCTTG
GGCTGCTGATCTCTCCTACATGCGAATCAGGAGC
AATGTTTACAATTGACATTAACCTTACTTTGT
CATGCATTAATTTCTATGAATTTCTCATCTGCTA
CAATCCCCTAGCTCGTATCTTACTCATGATCA
ATTAACCTTCAATCTACATTGGATTCCATTGA
ACCATTTCTTACATTGTCAACTAGTTTTATTGAT
GTAGTATTATCAAAAAATATTTTCTAATCA
TGTTCAATGTCAATTAAGAAGTTTCTTAATTC
ATACTTACCTTTATTTGCGGAAACTGCAACAA
CAAGCACAAATTAAT
```

After getting this result we did pairwise sequence alignment by EMBL-EBI and got which place is mutated.

```
#####
# Programme : needle
# Rundate : 10 June 2014 19:56:31
# Commandline : needle
# -auto
# -stdout
# -asequence emboss_needle-I20140708-195630-0864-53790836-pg.asequence
# -bsequence emboss_needle-I20140708-195630-0864-53790836-pg.bsequence
# -datafile EDNAFULL
# -gapopen 10.0
# -gapextend 0.5
# -endopen 10.0
# -endextend 0.5
```

```

# -aformat3 pair
# -snucleotide1
# -snucleotide2
# Align_format: pair
# Report_file: stdout
#####
#=====
#
# Aligned_sequences: 2
# 1: EMBOSS_001
# 2: EMBOSS_001
# Matrix: EDNAFULL
# Gap_penalty: 10.0
# Extend_penalty: 0.5
#
# Length: 1033
# Identity: 984/1033 (95.3%)
# Similarity: 984/1033 (95.3%)
# Gaps: 36/1033 ( 3.5%)
# Score: 4818.5
#
#
#=====
EMBOSS_001 1
ATGGATGCTCATACGCTCGAGAGCTCTCCGA
GAGATCCTCTCGAAGGTTT 50
|||||
EMBOSS_001 1
ATGGATGCTCATACGCTCGAGAGCTCTCCGA
GAGATCCTCTCGAAGGTTT 50

EMBOSS_001 51
GTTTCTAGAATTAGCCTGGTTATCAGAAAT
TGTCCCAAGCAATAGATTAC 100
|||||
EMBOSS_001 51
GTTTCTAGAATTAGCCTGGTTATCAGAAATTGT
CCCAATGAATAGATTAC 100

EMBOSS_001 101
GACTGATGAAACAA—
CATCGCACATGGGTTGAATAATTCTGAAAATGTA
148
|||||
EMBOSS_001 101
GACTGATGAAACAATTCATCGCACATGG
GTTGAATAATTCTGAAAATGTA 150

EMBOSS_001 149
CTGAAAGAGAAATCTTCACATTTCAATGT
TCTTCCTCTTGTAGATAAAAC 198
|||||
EMBOSS_001 151
CTGAAAGAGAAATCTTCACATTTCAATGT
TCTTCCTCTTGTAGATAAAAC 200

EMBOSS_001 199
AAAATTGTTTCATGAATTATTAGTGACACAT
GAGATAGACACATCGTCTCA 248
|||||
EMBOSS_001 201
AAAATTGTTTCATGAATTATTAGTGACACAT
GAGATAGACAGATCGTCTCA 250

EMBOSS_001 249
ATTGGTATGATAAATGTTATTTAAACGAAG
AACCGAATTCTTCACCAGAC 298
|||.|||||
EMBOSS_001 251
ATTGCTATGATAAATGTTATTTAAACGAAGAAC
CGAATTCTTCACCAGAC 300

EMBOSS_001 299 ATATAGCAT-
CATCCTAAATTAGCGCTATGCTTACAACGT TT
AGGTGCTA 347
| |||.|||||
EMBOSS_001 301 A-
ATAGTGATACCTAAATTAGCGCTATGCTT
ACAACGTTTAGGTGCTA 349

EMBOSS_001 348
ATCGACTGTTTAAACAAGTTCAGTTATAGC
AATAATGAATCAATAGACTTA 397
|||||
EMBOSS_001 350
ATCGACTGTTTAAACAAGTTCAGTTATAGCAA
TAATGAATCAATAGACTTA 399

EMBOSS_001 398
ACAGTTATCGGTCAGT-----
GTTGCGATCAGCTGC 428
|||||
EMBOSS_001 400
ACAGTTATCGGTCAGTCGGATATAACATTAATA
GAGTTGCGATCAGCTGC 449

EMBOSS_001 429

```

TGCATGGCGTCTATACGCGCCGTAAGCCG	478	ATTA AATTCTATGAATTTCT	778
ATTGGATAATAAATCCCCTG			
		EMBOSS_001 750	
EMBOSS_001 450		ACAATTGACATTA ACTTTACTTTGTCATGCAT	
TGCATGGCGTCTATACGCGCCGTAAGCCGATTGG		TAAATTCTATGAATTTCT	799
ATAATAAATCAAATG	499		
		EMBOSS_001 779	
EMBOSS_001 479		CATCTGCTACAATCCCCTAGCTCGTATCTTA	
ATAGATTCCATTTAAATCGTTTAATTTGTCTTCC		CTCATGATCAATTAACTT	828
TACTGATTGGAAAATG	528		
.		EMBOSS_001 800	
EMBOSS_001 500		CATCTGCTACAATCCCCTAGCTCGTATCTTA	
ATATACCTCATTTAAATCGTTTAATTTGTCTT		CTCATGATCAATTAACTT	849
CCTAC TGATTGGAAAATG	549		
		EMBOSS_001 829	
EMBOSS_001 529		TCAATCTACATTGGATTCCATTGAACCATTCTTA	
TGTGGTTTAGAACTTCATTTACTGGCTTT		CATTGTCAACTAGTT	878
TACGAGCTGCTTCACAATC	578		
		EMBOSS_001 850	
EMBOSS_001 550		TCAATCTACATTGGATTCCATTGAACCATT	
TGTGGTTTAGAACTTCATTTACTGGCTTT		CTTACATTGTCAACTAGTT	899
TACGAGCTGCTTCACAATC	599		
		EMBOSS_001 879	
EMBOSS_001 579		TTATTGATGTAGTATTATCAAAAAATATTT	
AAACTGGTTTAGAGTATCAGAAATAGTTGCT		TTCCTAATCATGTTCAATGT	928
TCTCAAAGTGA ACTTCTTA	628		
		EMBOSS_001 900	
EMBOSS_001 600		TTATTGATGTAGTATTATCAAAAAATATTT	
AAACTGGTTTAGAGTATCAGAAATAGTTG		TCCTAATCATGTTCAATGT	949
CTTCTCAAA GTGA ACTTCTTA	649		
		EMBOSS_001 929	
EMBOSS_001 629		CAATTAAGAAGTTTCTTAATTCATACTTAC	
TTGAAGAGCTGTATTCTGATACTGTACAAT		CTTTATTTTGC GGAACTGC	978
TCATCTCTTTAGCGCGCTAG	678		
		EMBOSS_001 950	
EMBOSS_001 650		CAATTAAGAAGTTTCTTAATTCATACTTACC	
TTGAAGAGCTGTATTCTGATACTGTACAAT		TTTATTTTGC GGAACTGC	999
TCATCTCTTTAGCGCGCTAG	699		
		EMBOSS_001 979	
EMBOSS_001 679		AACAACAAGCACAAA	
AGCCCCTTGGGCTGCTGATCTCTCCTACAT		TTAATTTGCAACCGTCGG	1011
GCGAATCAGGAGCAATGTTT	728		
		EMBOSS_001 1000	
EMBOSS_001 700		AACAACAAGCACAAAT	
AGCCCCTTGGGCTGCTGATCTCTCCTACAT		TAAT	1019
GCGAATCAGGAGCAATGTTT	749		
		EMBOSS_001 729	
EMBOSS_001 729		ACAATTGACATTA ACTTTACTTTGTCATGC	

The ATM protein is a serine/threonine kinase involved in DNA double-strand break repair (Lavin *et al.*, 2005 and Taylor and Byrd, 2005). Ataxiatelangiectasia is caused by the inheritance of biallelic deleterious mutations in the ATM gene and occurs in 1/40,000 to 1/

300,000 live births. Ataxia-telangiectasia is characterized by progressive cerebellar ataxia, oculomotor apraxia, telangiectasias of the conjunctiva and skin, immunodeficiency, sensitivity to ionizing radiation, and an increased rate of malignancies, particularly lymphoma and leukemia (Swift *et al.*, 1987).

Conclusion :

The Ataxia-telangiectasia-mutated (ATM) gene in humans was identified as the basis of a rare autosomal disorder leading to cancer susceptibility and is now well known as an important signal transducer in response to DNA damage. Ataxia telangiectasia mutated gene (ATM) is a newly identified member of PI3 kinase family that is mutated in human autosomal

recessive disease, ataxia telangiectasia (AT) . ATM gene is large, spanning about 150 kbs of genomic DNA, and is located on chromosome 11q22- 23. Mutations in the ATM gene are the cause of a rare autosomal recessive syndrome, ataxia-telangiectasia (AT). The finding that ATM heterozygotes have an increased breast cancer risk was supported by some studies but not confirmed by others. In our study, the entire coding sequence of the ATM gene was prescreened for mutations by the protein truncation test to detect the chain-terminating mutations that are highly predominant in patients with AT. This review covers the role of ATM in DDR signalling and describes the interaction of the ATM kinase with other proteins in order to fulfil its various functions.

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