

## RESEARCH ARTICLE

# Effects of mercury and its antagonistic effect with magnesium and sucrose on growth and NR activity in wheat *Triticum aestivum*(L.)

■ NITIKA

### SUMMARY

The present investigation was designed to evaluate the effects of different doses of mercury (0.02, 0.04, 0.06, 0.08, 0.10 mM) and combined effect of different doses of mercury with magnesium and sucrose (0.02, 0.04, 0.06, 0.08, 0.10 mM) on growth and NR activity in two varieties of *Triticum aestivum* (L.) A concentration dependent decrease in growth parameters like seedling height, fresh weight of seedling chlorophyll content and carbohydrate content was observed under the influence of different doses of mercury in both the varieties. The lowest dose of mercury (0.02 mM) increased the NR activity and protein content while subsequent doses (0.02, 0.04, 0.06, 0.08, 0.10 mM) showed inhibitory effects. The lower doses of the combined treatment of mercury with magnesium and mercury with sucrose (0.02 and 0.04 mM) increased the growth parameters, NR activity and protein content of *Triticum aestivum* while subsequent concentrations (0.06, 0.08 and 0.10 mM) showed inhibitory effect. Thus, the recovery of inhibitory effects of mercury on growth parameters, NR activity and protein content was noticed in combination treatments of mercury with magnesium and mercury with sucrose treatments.

**Key Words :** Antagonism, Magnesium, Mercury, Phytotoxicity, Sucrose

**How to cite this article :** Nitika (2016). Effects of mercury and its antagonistic effect with magnesium and sucrose on growth and NR activity in wheat *Triticum aestivum* (L.). *Internat. J. Plant Sci.*, **11** (1): 109-114.

**Article chronicle :** Received : 17.07.2015; Revised : 08.12.2015; Accepted : 18.12.2015

Most toxic ions are ubiquitous in the environment and many of them are essential for the growth of plant. However some of them (*viz.*, Zn, Cu, Hg, Mn, Fe, Mo and Ni) proved to be acute toxic if present in higher concentration (Saxena and Saxena, 2002). Agarwal and Bhattacharya (1989) postulated that mercury ( $Hg^2$ ) binds with sulphhydryl group (-SH) of protein and alter the molecular structure

of protein. Parmar *et al.* (2002) studied the effect of  $Hg^{2+}$  and  $Cr^{2+}$  in phaseolus seedling and found that both the heavy metals inhibited plant growth and peroxides activity. The inhibition of plant growth and development at higher concentration of mercury was also observed by Sutter *et al.* (2002). Heavy metals in combination with sucrose also increase the NR activity in bean leaf segments (Puranik and Srivastava, 1983).

Thus, the present investigation has been undertaken to study the individual and combined effect of mercury with magnesium and sucrose on growth, NR activity and

— AUTHOR FOR CORRESPONDENCE —

NITIKA, Department of Botany, G. M. V. Rampur Maniharan, SAHARANPUR (U.P.) INDIA

protein content in *Triticum aestivum* (L.).

## **MATERIAL AND METHODS**

The seeds of two cultivars of *Triticum aestivum* (L.), PBW – 343 and UP- 338 were presoaked in distilled water and were sown in three separate plots of field. In first, the irrigation of different doses of mercury (0.02, 0.04, 0.06, 0.08, 0.10 mM) and in second and third plot, the combined treatment of different doses of mercury with magnesium and mercury with sucrose ( in ratio 1:1, 0.02, 0.04, 0.06, 0.08, 0.10 mM) was given at the interval of 15 days, respectively. The effects of mercury alone and in combination with magnesium and sucrose was studied on seedling height, fresh weight of seedling, chlorophyll content, carbohydrate content, NR activity and protein content. The chlorophyll content was determined by the method of Arnon (1949), carbohydrate content was determined by the method of Morris (1948), protein content by Lowry *et al.* (1951) and NR activity was determined by the method of Srivastava (1974).

## **RESULTS AND DISCUSSION**

The results obtained from the present investigation as well as relevant discussion have been summarized under following heads :

### **Seedling height and fresh weight of seedling :**

A concentration dependent decrease was noticed in seedling height and fresh weight of seedling under the influence on different doses of mercury (0.02, 0.04, 0.06, 0.08, 0.10 mM) in both the varieties of *Triticum aestivum*. The maximum reduction in seedling height and fresh weight of seedling was obtained with the higher dose of mercury (0.10 mM) in both the varieties. In the combined treatment of mercury with magnesium, the lower doses (0.02 and 0.04) showed promotary effect on seedling height and fresh weight of seedling but the more pronounced increased was noticed with 0.02 mM concentration of mercury with magnesium. The subsequent concentration of the mercury with magnesium (0.06, 0.08, 0.10 mM) showed inhibitory effect on seedling height and fresh weight of seedling. (Table 1).

Similarly in case of the combination treatment of mercury with sucrose, the lower dosed (0.02 and 0.04 mM) showed promotary effect on seedling height and fresh weight of seedling. An increase in seedling height and fresh weight of seedling was more with 0.02 mM

concentration of mercury with sucrose treatment as compared to the 0.04 mM concentration of mercury with sucrose treatment. The increase in seedling height and fresh weight of seedling was more in mercury with magnesium treatment as compared to mercury with sucrose treatment (Table 1).

### **Chlorophyll and carbohydrate content :**

Concentration dependent decrease was also noticed in chlorophyll and carbohydrate content where the maximum reduction in nitrogen content was observed with the higher dose of mercury (0.10 mM) in both the varieties. The combined treatments of mercury with magnesium and mercury with sucrose showed increase in chlorophyll and carbohydrate contents at lower concentrations (0.02 and 0.04 mM) but the more pronounced increase was noticed with 0.02 mM concentration as compared to 0.04 mM concentration of treatments. The subsequent concentrations of mercury with magnesium and mercury with sucrose treatments (0.06, 0.08, 0.10 mM) showed inhibitory effect on chlorophyll and carbohydrate contents as compared to control. The reduction was more in variety PBW- 343 as compared to variety UP- 338. (Table 1).

### **NR activity and protein content :**

The NR activity and protein contents were increased with the lowest concentration of mercury (0.02 mM) while the subsequent concentrations of mercury (0.04, 0.06, 0.08, 0.10 mM) inhibited the NR activity and protein content. In combined treatment of mercury with magnesium and mercury with sucrose the similar pattern of increase and decrease of NR activity and protein content was observed as noticed in case of chlorophyll and carbohydrate contents (Table 2).

Heavy metals have been known to exert toxic influence at different level on the tissues of higher organisms. At an elevated level in soil heavy metals generally becomes toxic and can ultimately cause the death of plants. In the present investigation the concentration dependent decrease was noticed in seedling height and fresh weight of seedling under the influenced different doses of mercury (0.02, 0.04, 0.06, 0.08, 0.10 mM) and the maximum decrease was noticed under the influence of higher dose of mercury (0.10 mM). Heavy metal toxicity at higher concentration is well known (Mhatre and Chaphekarn, 1982; Lata, 1983; Sarkar and Aery, 1990; Salgare and Singh, 2001 and Sutter *et al.*, 2002). The higher concentrations of heavy

metals have also been reported to retard cell division and differentiation and reduce their elongation and affect ..... *et al.*, 1998 and Tomar *et al.*, 2000). Further the seedling height and fresh weight of seedling were increased with the lower concentration (0.02 and 0.04 mM) of combined

treatments of mercury with magnesium and mercury with sucrose but at the higher concentration (0.06, 0.08, 0.10 mM), they were decreased. Thus, mercury with magnesium and sucrose at lower concentration showed antagonistic effects.

The similar trend of reduction of chlorophyll and

**Table 1 : Effects of mercury and its combined treatments with magnesium and sucrose on growth parameters of *Triticum aestivum* (L.)**

Concentration of treatment (mM)	Seedling height (c.m.)	Fresh weight of seedling (g)	Chlorophyll content (mg g <sup>-1</sup> fr.wt.)	Carbohydrate content (mg g <sup>-1</sup> fr.wt.)
<b>Variety UP-338</b>				
Control	19.45±0.35	20.35±0.22	1.45±0.45	80.99±0.85
Hg <sup>2+</sup> 0.02	19.25±0.46	19.75±0.48	1.35*±0.92	80.42±0.69
Hg <sup>2+</sup> 0.04	17.05*±0.55	19.05*±0.82	1.22*±1.25	77.99±0.95
Hg <sup>2+</sup> 0.06	16.30*±1.25	18.06*±1.25	1.18*±2.45	67.69*±1.28
Hg <sup>2+</sup> 0.08	14.75*±2.15	16.52*±3.16	0.95*±0.91	61.55*±3.19
Hg <sup>2+</sup> 0.10	13.92*±4.25	12.10*±1.10	0.80*±1.13	54.28*±2.18
Hg <sup>2+</sup> 0.02 + Mg <sup>2+</sup> 0.02	22.95*±0.95	24.85*±0.86	1.67*±0.85	98.45*±0.92
Hg <sup>2+</sup> 0.04 + Mg <sup>2+</sup> 0.04	21.22*±2.11	22.65*±0.52	1.52*±0.81	94.16*±1.45
Hg <sup>2+</sup> 0.06 + Mg <sup>2+</sup> 0.06	17.95*±3.23	19.86*±1.25	1.23*±1.05	79.12*±2.39
Hg <sup>2+</sup> 0.08 + Mg <sup>2+</sup> 0.08	16.90*±1.15	17.30*±1.29	1.01*±2.93	74.06*±1.45
Hg <sup>2+</sup> 0.10 + Mg <sup>2+</sup> 0.10	16.12*±1.13	16.00*±1.15	0.99*±3.10	68.28*±2.15
Hg <sup>2+</sup> 0.02 + sucrose 0.02	22.95*±1.14	22.75*±0.95	1.67*±0.95	97.15*±1.24
Hg <sup>2+</sup> 0.04 + sucrose 0.04	20.80*±1.82	21.55*±1.25	1.58*±1.46	95.09*±1.45
Hg <sup>2+</sup> 0.06 + sucrose 0.06	18.03*±3.16	19.80*±1.35	1.41*±2.32	77.25*±1.82
Hg <sup>2+</sup> 0.08 + sucrose 0.08	16.55*±1.25	18.00*±2.15	1.23*±1.85	73.36*±2.89
Hg <sup>2+</sup> 0.10 + sucrose 0.10	15.7.2*±1.43	16.65*±1.32	0.61*±1.33	68.16*±1.87
<b>Variety PBW – 343</b>				
Control	19.10±0.85	20.00±1.45	1.77±0.95	79.30±0.85
Hg <sup>2+</sup> 0.02	19.45±0.41	19.95±1.32	1.54*±1.29	79.19±1.27
Hg <sup>2+</sup> 0.04	18.25*±0.92	18.45*±1.62	1.24*±3.13	66.27*±1.82
Hg <sup>2+</sup> 0.06	17.06*±1.25	16.50*±1.89	1.07*±3.42	59.45*±1.11
Hg <sup>2+</sup> 0.08	16.42*±1.43	13.40*±3.10	0.99*±1.05	55.11*±2.17
Hg <sup>2+</sup> 0.10	14.16*±1.29	12.91*±1.33	0.85*±2.95	49.42*±2.22
Hg <sup>2+</sup> 0.02 +Mg <sup>2+</sup> 0.02	22.22*±1.12	22.95*±1.31	1.73*±1.35	99.26*±1.12
Hg <sup>2+</sup> 0.04 +Mg <sup>2+</sup> 0.04	20.40*±1.49	21.20*±2.11	1.45*±1.67	92.33*±3.16
Hg <sup>2+</sup> 0.06 +Mg <sup>2+</sup> 0.06	18.45*±1.36	18.52*±2.49	1.29*±1.98	76.32*±1.66
Hg <sup>2+</sup> 0.08 +Mg <sup>2+</sup> 0.08	16.92*±2.67	18.00*±1.17	0.94*±2.15	73.43*±2.11
Hg <sup>2+</sup> 0.10 +Mg <sup>2+</sup> 0.10	14.20*±1.85	16.06*±1.31	0.89*±1.65	64.52*±1.68
Hg <sup>2+</sup> 0.02 +sucrose 0.02	22.00*±2.45	22.05*±0.89	1.58*±1.28	93.06*±1.15
Hg <sup>2+</sup> 0.04 +sucrose 0.04	21.05*±1.35	20.25*±2.63	1.43*±1.42	93.13*±2.88
Hg <sup>2+</sup> 0.06 +sucrose 0.06	18.12*±1.47	19.02*±1.68	1.31*±2.85	74.26*±2.12
Hg <sup>2+</sup> 0.08 +sucrose 0.08	15.25*±1.62	18.26*±1.11	0.94*±1.37	69.41*±1.85
Hg <sup>2+</sup> 0.10 +sucrose 0.10	13.82*±1.85	15.40*±3.21	0.82*±1.45	63.77*±1.43

\* indicate significance of value at P=0.05

carbohydrate content was also noticed under the influence of different doses of mercury (0.02, 0.04, 0.06, 0.08, 0.10 mM). Similarly, the reduction in chlorophyll and carbohydrate contents by mercury was also noticed by Vyas and Puranik (1993), Sharma *et al.* (1999); Mousume *et al.* (2000); Baszinski *et al.* (1980) and Prasad and Prasad (1987) where they concluded that there are two enzymes *i.e.* aminolaevulinic acid (ALA) dehydrates and protochlorophyllide reductases which are

involved in chlorophyll biosynthesis in higher plants and are inhibited by  $Hg^{2+}$  and  $Pb^{2+}$  because these metals bind with functional sulphhydryl (-SH) group of the enzymes. The similar trend of increase and decrease was also noticed in case of chlorophyll and carbohydrate contents as obtained in seedling height and fresh weight of seedling under the influence of combined treatment of mercury with magnesium and sucrose. Arvind (2004) found that heavy metal protects *Ceratophyllum*

**Table 2 : Effects of mercury and its combined treatments with magnesium and sucrose on growth parameters of *Triticum aestivum* (L.)**

Concentration of treatment (mM)	NR activity ( $M\ NO_2^{-1}hr^{-1}fr.wt.$ )	Protein content ( $mgg^{-1} fr.wt.$ )
<b>Variety- foundation</b>		
Control	16.14*±1.33	52.96±1.55
Hg <sup>2+</sup> 0.02	14.15*±1.24	54.16±1.32
Hg <sup>2+</sup> 0.04	25.25*±1.65	52.99±1.66
Hg <sup>2+</sup> 0.06	26.36*±1.82	44.63*±1.00
Hg <sup>2+</sup> 0.08	19.64*±2.46	41.56*±1.95
Hg <sup>2+</sup> 0.10	17.26*±1.80	36.15*±2.11
Hg <sup>2+</sup> 0.02 + Mg <sup>2+</sup> 0.02	13.45*±2.11	64.23*±1.05
Hg <sup>2+</sup> 0.04 + Mg <sup>2+</sup> 0.04	28.23*±1.13	61.08*±1.82
Hg <sup>2+</sup> 0.06 + Mg <sup>2+</sup> 0.06	24.19*±2.98	49.49*±1.25
Hg <sup>2+</sup> 0.08 + Mg <sup>2+</sup> 0.08	17.35*±1.34	45.39*±2.02
Hg <sup>2+</sup> 0.10 + Mg <sup>2+</sup> 0.10	16.57*±1.44	38.32*±0.92
Hg <sup>2+</sup> 0.02 + sucrose 0.02	14.30*±1.00	64.59*±1.06
Hg <sup>2+</sup> 0.04 + sucrose 0.04	27.13*±1.13	60.00*±2.86
Hg <sup>2+</sup> 0.06 + sucrose 0.06	22.14*±2.98	47.09*±1.11
Hg <sup>2+</sup> 0.08 + sucrose 0.08	16.85*±1.3	43.42*±1.05
Hg <sup>2+</sup> 0.10 + sucrose 0.10	13.15*±1.00	38.43*±3.01
<b>Variety- BH-902</b>		
Control	19.10±0.93	51.65±1.32
Hg <sup>2+</sup> 0.02	20.95±1.82	53.95±1.83
Hg <sup>2+</sup> 0.04	19.00*±1.02	52.35±1.23
Hg <sup>2+</sup> 0.06	17.53*±2.00	43.76*±1.02
Hg <sup>2+</sup> 0.08	15.51*±1.33	41.54*±0.92
Hg <sup>2+</sup> 0.10	12.45*±1.11	38.26*±1.26
Hg <sup>2+</sup> 0.02 +Mg <sup>2+</sup> 0.02	29.69*±2.05	64.68*±1.13
Hg <sup>2+</sup> 0.04 +Mg <sup>2+</sup> 0.04	25.65*±1.69	60.06*±1.18
Hg <sup>2+</sup> 0.06 +Mg <sup>2+</sup> 0.06	18.80*±1.01	45.34*±2.15
Hg <sup>2+</sup> 0.08 +Mg <sup>2+</sup> 0.08	16.55*±2.22	42.43*±1.16
Hg <sup>2+</sup> 0.10 +Mg <sup>2+</sup> 0.10	13.65*±1.83	37.06*±2.22
Hg <sup>2+</sup> 0.02 +sucrose 0.02	26.42*±3.42	62.53*±1.88
Hg <sup>2+</sup> 0.04 +sucrose 0.04	21.97*±1.25	60.43*±1.11
Hg <sup>2+</sup> 0.06 +sucrose 0.06	18.54*±2.00	45.35*±1.32
Hg <sup>2+</sup> 0.08 +sucrose 0.08	13.76*±1.19	41.73*±2.22
Hg <sup>2+</sup> 0.10 +sucrose 0.10	12.95*±1.63	38.78*±3.15

\* indicate significance of value at P=0.05

*demersum* L. and Pedler *et al.* (2004) found that combined treatment of zinc with magnesium showed promotary effect in wheat and radish.

The enzyme nitrate reductase is substrate inducible enzyme which is found in cytosole and the reduction of nitrate by the enzyme nitrate reductase, is rate limiting step in over all nitrate assimilation and its activity often controls the rate of protein synthesis in plants (Srivastava, 1980). In the present investigation the NR activity and protein content were increased with the lower concentration of heavy metal (0.02 mM) whereas the subsequent concentrations of (0.04, 0.06, 0.08 and 0.10 mM) of mercury showed inhibitory effect. Similar findings were also obtained by Veerappa and Samy (1999) where they found that lower concentration of mercury promote the NR activity whereas subsequent higher concentrations showed inhibitory effect in *Zea mays*. According to Singh *et al.* (1998), the inhibition of NR activity under the influences of heavy metals may be multifacial, eg. due to reduced supply of NADPH, disorganization of chloroplast, less NO<sub>3</sub> supply to the site of synthesis caused by water stress and direct effect of heavy metals on protein synthesis because it has a strong affinity for functional –SH group of the enzyme. Further the lower dose of combined treatments of mercury with magnesium and mercury with sucrose (0.02 and 0.04mM) were found to be promotary for NR and protein content whereas subsequent higher doses (0.06, 0.08 and 0.10 mM) showed inhibitory effect. Puranik and Srivastava (1983) and Vyas and Puranik (1993) observed that sucrose enhance the stability of NR and mobilization of endogenous nitrate pool. Bose and Mishra (1999) noticed that the magnesium salts increase the NR activity and protein content in *Brassica Juncea*. Kiss (1989) and Bose and Mishra (1992) concluded that application of magnesium to soil increases the nitrogen status of plants and ultimately the protein content.

Thus, the present investigation showed that all the concentrations of mercury exert toxic effect on all the parameters studied in *Triticum aestivum* (L.). NR activity and protein content which increased at lowest concentration (0.02 mM) but decreased at subsequent higher concentrations (0.04, 0.06, 0.08 and 0.10 mM) of mercury. The lower concentrations of combined treatments of mercury with magnesium and sucrose (0.02 and 0.04 mM) showed promotary effects on all the parameters studied while higher concentrations (0.06, 0.08 and 0.10 mM) showed inhibitory effect. Further,

the increase in all the parameters was more clear in the combined treatment of mercury with magnesium as compared to mercury with sucrose. The variety PBW – 343 was found to be more susceptible to metal toxicity as compared to variety UP-338.

## REFERENCES

- Agarwal, A. and Bhattacharaya, S. (1989). Binding property of rat and limulus C-reactive protein (CRP) to mercury. *Experientia*, **45** : 567-570.
- Arnon, D.I. (1949). Copper enzymes in isolated chloroplasts polyphenol oxidase in *Beta vulgaris*, *Plant physiol.*, **24** : 1-15.
- Arvind, P. (2004). Zinc protects chloroplast and associated photochemical functions in cadmium exposed *Ceratophyllum demersum* (L.) a fresh water macrophytes. *Plant Sci.*, **166** (4) : 1321-1327.
- Baszynski, T., Wajda, L., Krol, M., Wolinska, D., Krupa, Z., Tukendorf, A. (1980). Photosynthetic activities of cadmium-treated tomato plants. *Physiol. Plant.*, **48**: 365-370.
- Bose, B. and Mishra, T. (1992). Response of wheat seeds to pre-sowing seed treatments with Mg(NO<sub>3</sub>)<sub>2</sub> *Ann. Agric. Res.*, **13** : 132-136.
- Bose, B. and Mishra, T. (1999). Influence of pre-sowing soaking treatment in *Brassica juncea* seeds with Mg salts on growth, nitrate reductase activity, total protein content and yield responses. *Physiol. Mol. Biol. Plant.*, **5** : 83-88.
- Kastori, R., Plesnicar, M., Sakae, Z., Pnakovic, D. and Arsenijevic Lisksimovie, I. (1998). Effect of excess lead on sunflower growth and photosynthesis. *J. Plant Nutr.*, **21** : 75-85.
- Kiss, A.S. (1989). Effect of magnesium ions on anion uptake of plants. *Acta. Agron. Acad. Sci. Hung.*, **38** : 23-30.
- Lata, K. (1983). Effect of heavy metal (zinc) on seedling growth of some legumes. *J. Indian Bot. Soc.*, **62** :82.
- Lowery, O.H. Roseborough, N.J., Farr, A.L. and Radall, R.J. (1951). Protein measurement with Folin-Phenol reagent. *J. Biol, Chem.*, **193** : 265-279.
- Morris, Luzon, Daniel (1948). Quantitative determination of carbohydrates with Dreywood's Anthrone reagent. *Science*, **107** : 254-255.
- Parmar, N.G., Bithalani, S.D. and Chanda, S.V. (2002). Alternation in growth and peroxidase activity by heavy metals in phaseolus seedlings. *Acta Physiol. Plantarum*, **24** (1) : 89-95.

- Pedlar, J.F., Kihraide, T.B. and Parker, D.R. (2004). Zinc rhizo toxicity in wheat and radish is alleviated by micro molar level of magnesium and postassium in solution culture. *Plant & Soil.*, **259** : 1919 – 1929.
- Prasad, D.D.K. and Prasad, A.R.K. (1987). Effect of lead mercury on chlorophyll synthesis in mungbean seedling. *Phytochem.*, **26** : 881-883.
- Puranik, R.M. and Srivastava, H.S. (1983). Increase in nitrate reductase activity in the presence of sucrose in bean leaf segments. *Phytochem.*, **22** : 2383-2387.
- Salgare, S.A. and Singh, Sanju (2001). Effect of zinc sulphate on the rate of pollen germination of successive flowers of white flowered cultivar of *Catharanthus roseus*. *Bionotes.*, **3** (3) : 239-241.
- Sarkar, Sundra and Aery, N.C. (1990). Effect of zinc on growth of soybean. *Indian J. Plant Physiol.*, **33** (3) : 239-241.
- Saxena, Anuj and Saxena, D.K. (2002). Nitrate reductase activity and chlorophyll content in Sphagnum as affected by kinetin glutathione and metals. *Indian J. Plant Physiol.*, **77**(1) : 83-85.
- Sharma, Reeta, Sindhu, R.S. and Sharma, R. (1999). Bioaccumulation of lead, cadmium and mercury by *Mussaenda luteota*. *Indian J. Forest.*, **22**(3):226-227.
- Singh, R.P., Sabas, S., Choudhary, A. and Maheshwari, R. (1998). Effect of lead on nitrate reductase activity and alleviation of lead toxicity by inorganic salts and 6-benzylaminopurine. *Biologia Planterum.*, **40** (3) : 339-404.
- Srivastava, H.S. (1974). *In vivo* activity of nitrate reductase in maize seedlings. *Indian J. Biochem & Biophys.*, **11** : 230-232.
- Srivastava, H.S. (1980). Regulation of nitrate reductase activity in higher plants. *Phytochemistry*, **19** : 725-733.
- Sunder, S., Pareek, B.L. and Sharma, S.K. (2003). Effect of phosphorus and zinc on dry matter, uptake of nutrients and quality of clusterbean (*Cyamopsis tetragonoloba* L.) Tabu, *Ann. Agric. Res. New Series.*, **24** (1) : 19-196.
- Sutter, Kristin, Jungklaus and Krauss – Gerd-Joachin (2002). Effect of heavy metals on the nitrogen metabolism of the aquatic moss *Fontinalis antipyretica*. L. exhedw : A 15 N trace study. *Environ. Sci. & Pollut. Res. Internat.*, **9**(6) : 417-421.
- Tomar, Manju, Kaur, Indradeep, Neelu and Bhatnagar, A.K. (2000). Effect of enhanced lead in soil on growth and development on *Vigna radiate* (L.) Witezek. *Indian J. Plant Physiol.*, **5** (1) : 13-18.
- Veerappa, N. Sami and Samy, Arockia, D.I. (1999). Zinc ion toxicity and its alleviation by manganese and copper ions in maize seedlings (*Zea mays*) with special reference to nitrate reductase activity. *Physiol. Mol. Biol. Plants*, **5** : 63-66.
- Vyas, Jot and Puranik, Rekha (1993). Inhibition of NRA by mercury in bean leaf segment. *Indian J. Plant Physiol.*, **36** (1) : 55-60.

11<sup>th</sup>  
Year  
★★★★★ of Excellence ★★★★★