



RESEARCH PAPER

RAPD based molecular diversity analysis of different varieties of pomegranate (*Punica granatum* L.)

AMOL S. SHINDE*, SAGAR R. MAHAJAN AND SAGAR H. KAKDE
Lokmangal Agricultural Biotechnology College, Wadala, SOLAPUR (M.S.) INDIA
(Email: amolshinde0110@gmail.com)

Abstract : Genetic diversity of eight pomegranate varieties was carried out using five RAPD primers. The DNA was extracted from young leaves using CTAB method. The PCR for RAPD was performed with two primers from OPA series, and four primers from OPB series. The RAPD analysis with five arbitrary oligonucleotide primers amplified a total of 28 DNA bands out of which 25 were found to be polymorphic. The average polymorphism recorded by the RAPD loci was 89.28 per cent. The number of DNA fragment varied from four to seven. The mean number of polymorphic bands per primer among eight pomegranate varieties was 5.6 and per cent polymorphism ranged from 75 to 100. The size of PCR amplified DNA fragment ranged from 88.49 to 1430.11bp and PIC value varied from 0.70 to 0.83. The dendrogram constructed using pooled RAPD loci data clearly showed two varieties (Ganesh and Mrudula) were highly similar and different from other genotypes. The genetic similarities ranged from 0.32 to 0.72 and mean similarity co-efficient was 0.61.

Key Words : RAPD, *Punica granatum* L., PCR, Primer, Molecular marker

View Point Article : Shinde, Amol S., Mahajan, Sagar R. and Kakde, Sagar H. (2015). RAPD based molecular diversity analysis of different varieties of pomegranate (*Punica granatum* L.). *Internat. J. agric. Sci.*, **11** (1): 141-145.

Article History : Received : 08.10.2014; Revised : 24.11.2014; Accepted : 10.12.2014

INTRODUCTION

Pomegranate (*Punica granatum* L.) belongs to Punicaceae family and is an important fruit tree of tropical and subtropical regions of the world which is valued highly for its delicious edible fruits. The cultivated varieties of pomegranate, *Punica granatum* is to contain $2n=2x=16, 18$ chromosomes. It belongs to the subclass Rosidae and believed to be native to the region between Iran to northern India (Stover and Mercure, 2007). In India, pomegranate grows wild in Western Himalayan regions that include states like Himachal Pradesh, Jammu and Kashmir and Uttarakhand (Misra *et al.*, 1983; Pandey *et al.* (2008). Pomegranate may be classified according to the acidity of its fruit into sour, sour-sweet or sweet.

The total area under cultivation of pomegranate in India

is 113.240 thousand ha and production is around 744.950 MT tonnes in 2012-13. Maharashtra is the leading producer of pomegranate having 55.76 per cent production followed by Karnataka, Andhra Pradesh, Gujarat and Tamil Nadu. The total area under cultivation of pomegranate in Maharashtra is 78 thousand ha and production is around 408 thousand MT in 2012-13. Ganesh, Bhagwa, Ruby, Arakta and Mridula are the different varieties of pomegranates produced in Maharashtra. Currently, it is an important fruit species for India, Iran, USA and Mediterranean countries like Greece, Spain, Tunisia. Pomegranate fruit juice makes an excellent drink which contains potassium, phosphorus and calcium as well as micronutrients like iron, manganese, zinc and copper. The antioxidant, immune-boosting, and anti-carcinogenic properties of the pomegranate offers its multiple potential

* Author for correspondence

medical applications (Kaplan *et al.*, 2001). These have been demonstrated to be beneficial in combating high blood pressure and other serious diseases such as diabetes and various cancers (Shishodia *et al.*, 2006). Although the chemical composition of the fruit is affected from cultivar, growing region, climate, maturity, cultural practice and storage (Melgarejo *et al.*, 2000). In addition, the tree is also cultivated for its pharmaceutical and ornamental usages (Levin, 1994).

DNA markers are independent from environmental interactions and show high level of polymorphism therefore, they are considered as useful tools for determining genetic relationships and diversity. Polymorphisms are detected from differences in the length of the amplified fragments by polyacrylamide gel electrophoresis (PAGE) (Matthes *et al.*, 1998) or by capillary electrophoresis. They also have been used to investigate relationships of closely related taxa (Miller and Tanksley, 1990; Lanner *et al.*, 1997), as fingerprinting tools (Fang *et al.*, 1997), for diversity studies (Debreuil *et al.*, 1996). Although a wide range of morphological and physiological characters show variability's in the pomegranate, molecular studies of the pomegranate have been restricted to examinations of RAPD (Durgac *et al.*, 2008; Zamani *et al.*, 2007 and Sarkhosh *et al.*, 2009), to investigate the population dynamics of economically important cultivars.

MATERIAL AND METHODS

The present investigation entitled RAPD based molecular diversity analysis of different varieties of pomegranate (*Punica granatum* L.) was carried out at Lokmangal Agricultural Biotechnology College, Wadala, Solapur, Maharashtra.

Plant material :

Experimental material comprised of eight cultivated varieties of pomegranate namely Ganesh, Bhagwa, Arakta, Supper Bhagwa, Mrudulla, Rubby, G-137 and Tissue culture Bhagwa were collected from different pomegranate cultivating area of Maharashtra.

DNA isolation :

Genomic DNA was isolated from juvenile fresh leaves of 8 different varieties of pomegranate collected from different pomegranate cultivation area of Maharashtra following CTAB (Cetyltrimethyl Ammonium Bromide) extraction method given by Gawel and Jarret (1991) with some modifications.

RAPD analysis :

Five of the available decamer random oligonucleotide primers were used to ascertain polymorphism among eight different varieties of pomegranate. The polymerase chain reaction (PCR) as adopted by Mathews *et al.* (2007) with minor modifications was carried out in 25 µl of reaction mix (1X Taq buffer, 17µl sterile DDH₂O, 25mM MgCl₂, 2.5mM dNTP, 20pmol

primer, 1U Taq DNA polymerase, 50 ng DNA). Amplification reactions were carried out for 40 cycles. Each cycle comprised of 1 min at 94°C, 1:30 min at 37°C and 2 min at 72°C. Amplified product were separated on 1.6 per cent agarose gel, stained with ethidium bromide and photographed under UV light.

Data analysis :

Data were scored for computer analysis on the basis of the presence or absence of the PCR products. If a product was present in a genotype, it was designated as '1' and if absent; it was designated as '0'. The data generated by RAPD and SSR loci were analyzed with the software NTSYSpc version 2.02 (Rohlf, 1994). The PIC values were calculated with formula $PIC=1-\sum p_i^2$ (where pi is the frequency of the ith allele) given by Smith *et al.* (1997).

RESULTS AND DISCUSSION

The findings of the present study as well as relevant discussion have been presented under following heads :

Random amplified polymorphic DNA (RAPD) :

The genomic DNA extracted from each genotype was subjected to polymerase chain reaction using fourteen random decamer primers from OPA, OPB and OPP series. Molecular characterization of eight different pomegranate varieties carried out using RAPD primer. In RAPD assay total five primers were selected for pooled analysis from fourteen primers. PCR amplification of DNA, using five primers from OPA and OPB series for RAPD analysis, produced 28 DNA fragments. All the selected 5 primers amplified DNA fragments across 8 genotypes. The total number of amplified fragments was varying from four (OPB-18) to seven (OPA-07), with size ranged from 88.49 to 1430.11bp (Table 1). Same result was found by Hasnaoui *et al.* (2010) in their study on molecular polymorphisms in Tunisian pomegranate. Out of 28 DNA fragments 25 were polymorphic giving 89.28 per cent average polymorphism for total OPA and OPB series. The average polymorphic band per primer was 5.6 and per cent polymorphism ranged from 75 (OPB-18) to 100 *i.e.* OPA-07 and OPB-15, respectively (Plate 1). Near about same results were revealed by Jambhale *et al.* (2007) in their study on molecular characterization of pomegranate cultivars with RAPD markers. The PIC value varied from 0.70 (OPB-18) to 0.83(OPA-07) (Fig. 1).

Dendrogram (Fig. 1) constructed with the data generated by all five OPA and OPB primers and their amplicons grouped the all pomegranate varieties in to one cluster *i.e.* cluster A. The cluster A grouped all seven varieties of pomegranate *i.e.* T.C. Bhagwa, S. Bhagwa, Arakta, Mrudulla, Ganesh, Rubby and Bhagava except one variety *i.e.* G-137. Cluster A further divided into sub cluster A1 and sub-cluster A2. Sub-cluster A1 grouped total five different varieties *i.e.* T.C. Bhagwa, Mrudulla, Ganesh, Arakta and Rubby while sub-cluster A2

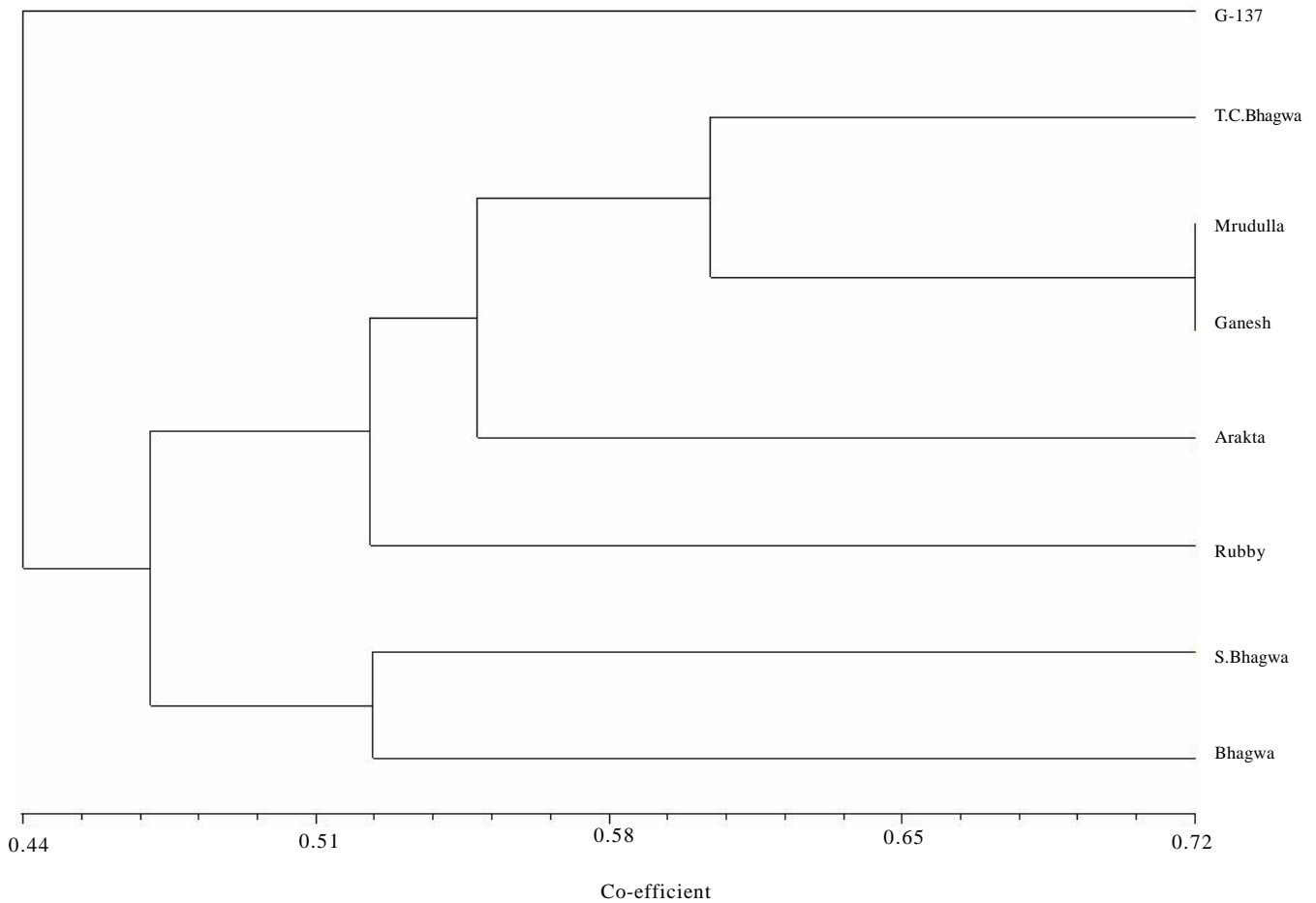


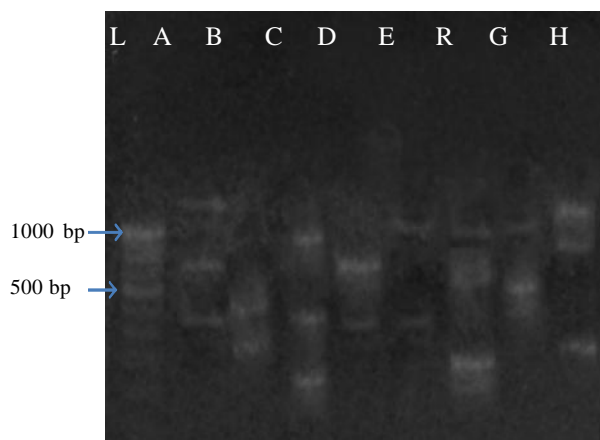
Fig. 1 : Dendrogram showing clustering of eight pomegranate varieties obtained from RAPD marker

Table 1 : Result of RAPD primers

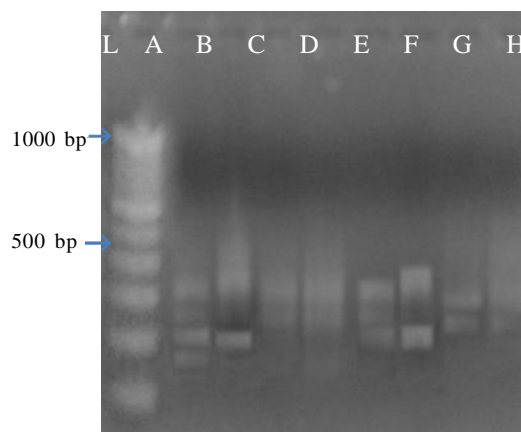
Sr. No.	Primer code	Mole wt. (bp)		Total no. of band	No. of polymorphic band	Per cent polymorphism	PIC value
		High	Low				
1.	OPB-15	1430.11	174.81	6	6	100	0.79
2.	OPB-18	304.84	134.9	4	3	75	0.70
3.	OPA-19	1013.16	308.19	5	4	80	0.74
4.	OPB-07	451.72	88.49	6	5	83.33	0.77
5.	OPA-07	458.95	90.42	7	7	100	0.83
Total				28	25	Average	89.28

Table 2 : Jaccard's similarity co-efficient for eight pomegranate varieties based on RAPD data analysis

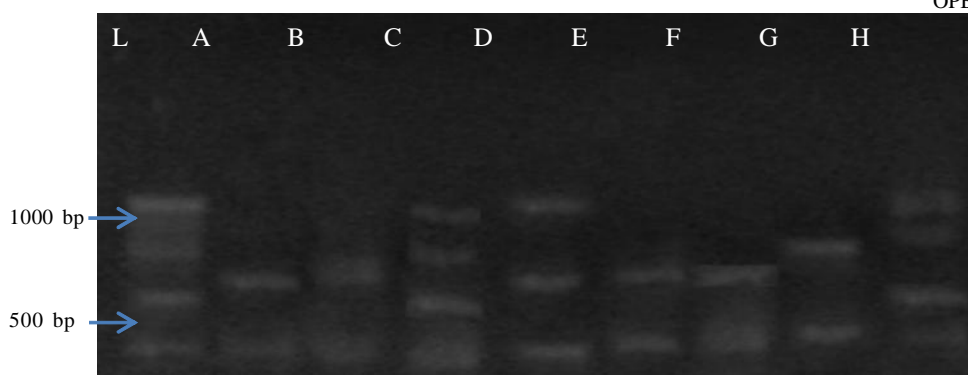
Varieties	G-137	T.C. Bhagwa	S. Bhagwa	Arakta	Mrudulla	Ganesh	Rubby	Bhagwa
G-137	1.0000							
T.C. Bhagwa	0.5500	1.0000						
S. Bhagwa	0.3200	0.3913	1.0000					
Arakta	0.4500	0.4736	0.5000	1.0000				
Mrudulla	0.4761	0.5789	0.6842	0.6470	1.0000			
Ganesh	0.4545	0.6315	0.5000	0.5263	0.7222	1.0000		
Rubby	0.4166	0.5000	0.5217	0.4761	0.5714	0.5454	1.0000	
Bhagwa	0.4090	0.3636	0.5238	0.4000	0.4285	0.4761	0.4347	1.0000



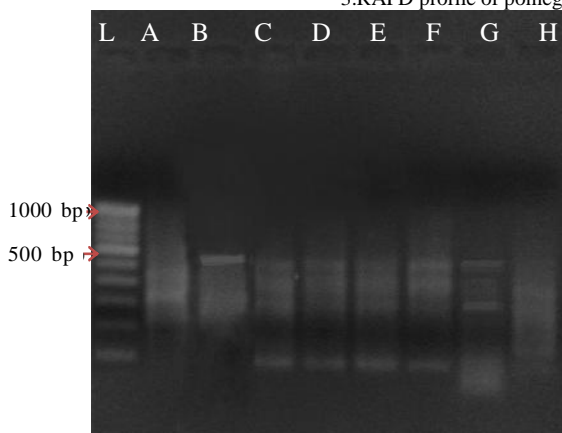
1. RAPD profile of pomegranate (*Punica grantum* L.) with primer OPB-15



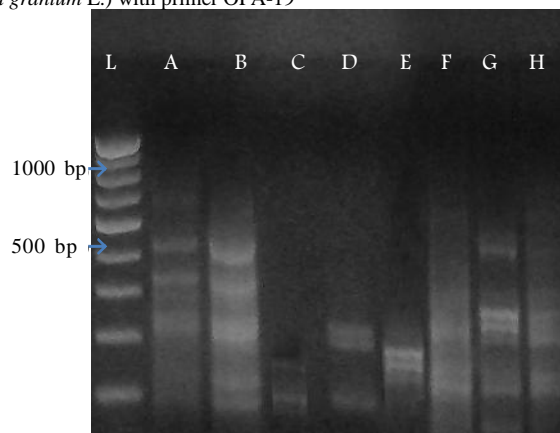
2. RAPD profile of pomegranate (*Punica grantum* L.) with primer OPB-18



3. RAPD profile of pomegranate (*Punica grantum* L.) with primer OPA-19



4. RAPD profile of pomegranate (*Punica grantum* L.) with primer OPB-07



5. RAPD profile of pomegranate (*Punica grantum* L.) with primer OPA-07

L – Ladder 100 bp, A -G-137, B -T.C.Bhagwa, C -S. Bhagwa, D –Arakta, E – Mrudula, F-Ganesh, G-Rubby, H-Bhagwa.

Plate 1 : RAPD pattern of different pomegranate varieties produced by OPB-15, 18, 07, OPA- 19, 07.

grouped remaining two varieties *i.e.* S. Bhagwa and Bhagwa. Jaccard's pair-wise similarity co-efficient values for eight genotypes were calculated and are presented in Table 2. The genetic similarities ranged from 0.32 to 0.72. The present finding

is near about similar with result obtained by Sheidai *et al.* (2008) studied on pomegranate cultivars. The average genetic similarity among these 8 genotypes was 0.61 (Table 2). The highest similarity index value of 0.72 was found between

Ganesh and Mrudula (Table 2) and lowest similarity index value of 0.32 was found between S. Bhagwa and G-137 (Table 2).

Conclusion :

The result indicates the OPB-15 and OPA-07 primers were more informative as compared to other primers for pomegranate genotyping. The dendrogram constructed using molecular data generated by five RAPD primers showed higher similarity in between Mrudula and Ganesh while lowest similarity found in S Bhagava and G-137. RAPD markers were found to be highly polymorphic and can be utilized for genetic diversity analysis of pomegranate genotypes.

REFERENCES

- Dubreuil, P., Dufour, P., Krejci, E., Causse, M., De Vienne, D., Gallais, A. and Charcosset, A. (1996).** Organization of RFLP diversity among inbred lines of maize representing the most significant heterotic groups. *Crop Sci.*, **36** : 790-799.
- Durgac, C., Ozgen, M., Simsek, O., Kacar, Y.A., Kýyga, Y., Celebi, S., Gunduz, K. and Serce, S. (2008).** Molecular and pomological diversity among pomegranate (*Punica granatum* L.) cultivars in Eastern Mediterranean region of Turkey. *African J. Biotechnol.*, **7** : 1294-1301.
- Fang, D.R., Krueger, R. and Roose, M.L. (1998).** Phylogenetic relationships among selected citrus germplasm accessions revealed by inter-simple sequence repeat (ISSR) markers. *J. American Soc. for Hort. Sci.*, **123** (4) : 612-617.
- Gawel, N.J. and Jarret, R.L. (1991).** A modified CTAB DNA extraction procedure for musa and ipomoea. *Plant Molecular Biol. Reporter*, **9** (3) : 262-266.
- Hasnaoui, N., Mars, M., Chibani, J. and Trifi, M. (2010).** Molecular polymorphisms in tunisian pomegranate (*Punica granatum* L.) as revealed by RAPD fingerprints. **2** : 107-114.
- Jambhale, V.M., Patil, S.C., Pawar, S.V., Jadhav, A.S., Patil, H.E. and Dahake, K.D. (2007).** Molecular characterization of pomegranate cultivars with RAPD markers. *Asian J. Hort.*, **2**(1) : 176-179.
- Kaplan, M., Hayek, T., Raz, A., Coleman, R., Dornfeld, L., Vaya, J. and Aviram, M. (2001).** Pomegranate juice supplementation to atherosclerotic mice reduces macrophage lipid peroxidation, cellular cholesterol accumulation and development of atherosclerosis. *J. Nutr.*, **131**(8) : 2082-2089.
- Lanner, H.C., Gustafsson, M., Falt, A.S. and Bryngelsson, T. (1996).** Diversity in natural populations of wild *Brassica oleracea* as estimated by isozyme and RAPD analysis. *Genet Resour Crop Evol.*, **43** (1) : 13-23.
- Levin, G.M. (1994).** Pomegranate (*Punica granatum*) plant genetic resources in Turkmenistan. *Plant Genetic Resource Newsletters*, **97**: 31-36.
- Mathews, M.D., Srinivasachary, Sujatha, R., JeVrey, L.B., Mike, D., Gale and Katrien, M.D. (2007).** The genetic map of finger millet *Eleusine coracana*. *Theor. Appl. Genet.*, **114** (2) : 321-332.
- Matthes, M.C., Daly, A. and Edwards, K.J. (1998).** Amplified fragment length polymorphism (AFLP). Molecular tools for screening biodiversity. *Chapman & Hall, Cambridge*, **1**(99) : 183-19.
- Melgarejo, P., Salazar, D.M., Artes, F. (2000).** Organic acids and sugar composition of harvested pomegranate fruits. *Eur. Food Res. Technol.*, **211** (3) : 185-190.
- Miller, J.C. and Tanksley, S.D. (1990).** Effects of restriction enzymes, probe source and probe length on detecting restriction fragment length polymorphism in tomato. *Theor. Appl. Genet.*, **80** (3) : 385-389.
- Misra, R.S., Srivastava, R.P., Kuksal, R.P. (1983).** Evaluation of some pomegranate cultivars for valley areas of Garhwal hills. *Prog. Hort.*, **15** : 24-26.
- Pandey, A., Tomer, A.K., Bhandari, D.C. and Pareek, S.K. (2008).** Towards collection of wild relatives of crop plants in India. *Genet. Resour. Crop Evol.*, **55** (2) : 187-202.
- Rohlf, F.J. (1994).** NTSYS-PC. Numerical taxonomy and multivariate analysis system version 2.02. Stat University of New York, Stonybrook, NEW YORK, U.S.A.
- Sarkhosh, A., Zamani, Z., Fatahi, R. and Ebadi, A. (2006).** RAPD markers reveal polymorphism among some Iranian pomegranate (*Punica granatum* L.) genotypes. *Scientia Horticulturae*, **111** (1) : 24-29.
- Shishodia, S., Adams, L., Bhatt, I.D. and Aggarwal, B.B. (2006).** Anticancer potential of pomegranate. In: Seeram NP, Schulman RN, Heber D (Eds.) *Pomegranates: ancient roots to modern medicine*. CRC Press Taylor and Francis Group, Boca Raton, 107-116pp.
- Smith, J.S.C., Chin, E.C.L., Shu, H., Smith, O.S., Wall, S.J., Senior, M.L., Mitchel, S.E., Kresorich, S. and Tiegle J. (1997).** An evaluation of the utility of SSR loci as molecular markers in maize (*Zea mays* L.): comparisons with data from RFLPs and pedigree. *Theor. & Appl. Genet.*, **95** (1-2) : 163-173.
- Stover, E. and Mercure, E.W. (2007).** The pomegranate: A new look at the fruit of paradise. *Hort. Sci.*, **42** (5) : 1088-1092.
- Zamani, Z., Sarkhosh, A., Fatahi, R. and Ebadi, A. (2007).** Genetic relationships among pomegranate genotypes studied by fruit characteristics and RAPD markers. *J. Hort. Sci. & Biotechnol.*, **82**: 11-18.

11th
Year
★★★★★ of Excellence ★★★★★