Determination of hosts of sunflower necrosis virus



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SUMMARY —

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Correspondence to : N.S. PANKAJA A.I.C.R.P. on Rice Agricultural Research Station, GANGAVATI (KARNATAKA) INDIA Various crop plants tested by mechanical sap and thrips inoculation, the virus infected *Helianthus* annuus (cv. KBSH-44), Citrullus lanatus (cv. Arka Manik), Cucurbita moschata (cv. Arka Suryamukhi), Arachis hypogaea (cv. JL-24), Lablab purpureus (cv. HA-3), Macrotyloma uniflorum (cv. PHG-9), Vigna unguiculata (cv. C-152), Nicotiana tabacum (cv. Xanthi) and Glycine max (cv. KB-79). The highest mean per cent transmission was recorded on *Helianthus annuus* (cv. KBSH-44) both by sap inoculation and thrips transmission (53.33 and 26.67, respectively). Among thirty six weed plants tested, twenty four weeds viz., Lagascea mollis, Alternanthera sessilis, Commelina benghalensis, Crotalaria spectabilis, Euphorbia hirta, Cassia obtusifolius, Ocimum sanctum, Sida rhombifolia, Oxalis corniculata, Physalis minima, Galinsoga parviflora, Euphorbia geniculata, Solanum nigrum, Phyllanthus niruri, Malvestrum coromandelianum, Ageratum conyzoides, Achyranthus aspera, Abutilon indicum, Ocimum canum, Crotalaria striata, Bidens pilosa, Stachytarpeta indica, Acanthospermum hispidum and Xanthium strumarium were able to show symptoms through sap and thrips inoculation. Highest mean per cent transmission however was observed in case of Achyranthes aspera (48.00) through sap and in Galinsoga parviflora (25.00) through thrips.

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serious virus disease on sunflower with Anecrotic symptoms causing severe yield loss was reported to occur around Bangalore (Anonymous, 1997; Singh et al., 1997). Because of its fast spreading nature, this necrosis virus was considered as one of the deadly virus diseases on this crop in India (Nagaraju et al., 1998). Necrosis virus can cause infection at any stage of the plant growth causing necrosis of a part of the leaf lamina and making the leaf to twist, followed by varied type of necrosis and mosaic symptoms (Nagaraju and Hanumantha Rao, 1999). Ajith Prasad and Nagaraju (2005) investigated and determined the disease transmission through sap and Thrips palmi (Karny). Host-range studies carried out by Ramaiah et al. (2001) revealed that an isometric virus causing sunflower necrosis disease could infect members of plants belonging to families Amaranthaceae, Chenopodiaceae and Fabaceae. Present study was conducted in order to know the various plants and weed

species to which the virus can infect.

MATERIALS AND METHODS -

An attempt was made to study the transmission of sunflower necrosis virus to thirty nine different crop plants and thirty seven different weeds found in and around sunflower fields at Zonal Agricultural Research Station, GKVK, UAS, Bangalore belonging to different families *viz.*, Asteraceae, Brassicaceae, Cucurbitaceae, Caricaceae, Euphorbiaceae, Fabaceae, Malvaceae, Pedaliaceae, Solanaceae, Convolvulaceae, Amranthaceae, Labiateae, Commelinaceae, Portulacaceae and Oxalidaceae, by mechanical sap and thrips inoculations.

Transmission of SNV by mechanical sap inoculation:

A set of thirty nine crop plants and thirty seven weed plants (three replications each) were sap inoculated at glasshouse conditions. Young tender leaves showing typical symptoms of sunflower necrosis disease were collected from naturally infected sunflower plants from the fields in and around the campus. These leaves were used to prepare standard inoculum using the 0.05M phosphate buffer. The mechanically inoculated plants were labeled and kept for symptom expression in glasshouse and observed upto 40 days.

Transmission of SNV through Thrips palmi:

About 20-30 nymphs were released on young sunflower leaves showing clear disease symptoms. The nymphs were allowed for acquisition access period (AAP) of three days under normal room temperature. Nymphs fed on healthy leaves served as check. After three days, 20-25 nymphs were transferred on to the test crop and weed plants belonging to different families raised in insect proof wooden cages. Three replications of the same were maintained. The plants were kept undisturbed allowing the nymphs to feed. These test plants were kept under observation upto 50 days for symptom expression.

RESULTS AND DISCUSSION -----

Sunflower necrosis virus infected the crop plants viz., Helianthus annuus (cv. KBSH-44), Citrullus lanatus (cv. Arka Manik), Cucurbita moschata (cv. Arka Suryamukhi), Arachis hypogaea (cv. JL-24), Lablab purpureus (cv. HA-3), Macrotyloma uniflorum (cv. PHG-9), Vigna unguiculata (cv. C-152), Nicotiana tabacum (cv. Xanthi) and Glycine max (cv. KB-79) on mechanical sap and thrips inoculation. Similar results were obtained by Halakeri (1999) who reported Vigna unguiculata (cv. C-152), Anil Kumar (1999) reported Cucumis sativus (cv. GREEN LONG) as the hosts of SNV (Table 1).

Mean per cent transmission of SNV through sap ranged from 15.00 to 53.33. Incase of *Helianthus annuus* (KBSH-44) highest mean per cent transmission of 53.33 was observed.

Mean per cent transmission of SNV through *Thrips* palmi Karny ranged from 6.67 to 26.67. Highest mean per cent transmission of 26.67 was also observed incase of *Helianthus annuus* (cv. KBSH-44), *Arachis* hypogaea (cv. JL-24) and *Citrullus lanatus* (cv. Arka Manik). However, *Glycine max* (cv. KB-79) did not produce any symptoms due to thrips transmission.

The symptoms produced were chlorotic patches on leaves, followed by necrotic lesions, downward twisting of leaves, vein thickening and stunted growth incase of *Helianthus annuus* (cv. KBSH-44). In Arachis hypogaea (cv. JL-24), the virus produced symptoms as

d upto 40puckering reduction in the leaf size was observed. In
Citrullus lanatus (cv. ARKA MANIK) mild mosaic to
chlorotic patches, necrosis of leaves and growing bud
were recorded. On crops viz., Lablab purpureus (cv.
n young
oms. The
(cv. KB-79) mosaic symptom was recorded. Whereas
incase of Cucurbita moschata (cv. ARKA SURYAMUKHI)
chlorotic patches with necrotic lesions on half of the
leaves was recorded. However, incase of Nicotiana
tabacum (cv. XANTHI) mosaic mottling were observed.
Similar observations on symptoms were made by Anil
Kumar (1999) and Halakeri (1999).allowing
ept under
ton.Among thirty six weed plants tested twenty four
weeds viz., Lagascea mollis, Alternanthera sessilis,
Commelina benghalensis, Crotalaria spectabilis,
Euphorbia hirta, Cassia obtusifolius, Ocimum
sanctum, Sida rhombifolia, Oxalis corniculata,
Physalis minima, Galinsoga parviflora, Euphorbia
geniculata, Solanum nigrum, Phyllanthus niruri,

Commelina benghalensis, Crotalaria spectabilis, Euphorbia hirta, Cassia obtusifolius, Ocimum sanctum, Sida rhombifolia, Oxalis corniculata, Physalis minima, Galinsoga parviflora, Euphorbia geniculata, Solanum nigrum, Phyllanthus niruri, Malvestrum coromandelianum, Ageratum conyzoides, Achyranthus aspera, Abutilon indicum, Ocimum canum, Crotalaria striata, Bidens pilosa, Stachytarpeta indica, Acanthospermum hispidum and Xanthium strumarium were able to show symptoms on SNV inoculation through sap and through thrips. The symptoms were observed at 20-25 days after inoculation. These findings are similar to the results obtained by Lokesh (2005), who reported Galinsoga parviflora, Euphorbia geniculata, Phyllanthus niruri, Malvestrum coromandelianum, Achyranthus aspera, Abutilon indicum, Ocimum canum, Crotalaria striata, Bidens pilosa, Acanthospermum hispidum, Guizotia abyssinica and Ageratum conyzoides as weed hosts for SNV.

mild mosaic with chlorotic patches, vein thickening,

necrosis on the tip of the leaf and also at the growing

bud. Further, reduction in leaf size and stunted growth

was observed. Incase of Macrotyloma uniflorum (cv.

PHG-9), mild mosaic and necrosis, vein thickening, severe

On Alternanthera sessilis, Commelina benghalensis, Sida rhombifolia Oxalis corniculata, Euphorbia hirta, Lagascea mollis, Solanum nigrum, Phyllanthus niruri, Ocimum canum, Xanthium strumarium and Bidens pilosa mosaic and mosaic mottling was observed.

Chlorotic patches were observed on *Cassia* obtusifolius, Ocimum sanctum, Physalis minima, Galinsoga parviflora, Euphorbia geniculata, Ageratum conyzoides, Abutilon indicum and Crotalaria striata. Where as chlorosis followed by vein clearing and by

DETERMINATION OF HOSTS OF SUNFLOWER NECROSIS VIRUS

Tabl	e 1: Transmission of SNV to crop	echanical sap inoculation		М		
Sr. No.	Scientific name	Common	variety Family		transmission	Symptoms
		name			(%)	
1.	Helianthus annuus	Sunflower	KBSH-44	Asteraceae	53.33	CP, NL, DTL, VT, SG
2.	Carthamus tinctorius	Safflower	C-18	Asteraceae		
3.	Guizotia abyssinica	Niger	S-26	Asteraceae		
4.	Tagetus erecta	Marigold	Chinthamani	Asteraceae		
5.	Zinnia elegans	Zinnia	ZE-231	Asteraceae		
6.	Brassica nigra	Mustard	Kanakpura local	Brassicaceae		
7.	Brassica oleracea var.	Knol khol	KO-O4-890	Brassicaceae		
	gongylodes					
8.	Brassica oleracea var capitata	Cabbage	B0-01-001	Brassicaceae		
9.	Raphanus sativus	Radish	Mino Long	Brassicaceae		
10.	Cucumis sativus	Cucumber	PS –24	Cucurbitaceae		
11.	Citrullus lanatus	Watermelon	Arka Manik	Cucurbitaceae	48.33	M, CP, NGT
12.	Cucumis melo	Musk melon	Deepti	Cucurbitaceae		
13.	Cucurbita moschata	Pumpkin	Arka	Cucurbitaceae	25.00	CP, NL
			suryamukhi			
14.	Lagenaria siceraria	Bottle gourd	PSPR	Cucurbitaceae		
15.	Luffa acutangula	Ridge gourd	Arka sujata	Cucurbitaceae		
16.	Momordica charantia	Bitter gourd	Arka Harit	Cucurbitaceae		
17.	Trichosanthes cucumerina	Snake gourd	C0-2	Cucurbitaceae		
18.	Ricinus communis	Castor	DCS-9	Euphorbiaceae		
19.	Arachis hypogaea	Groundnut	JL-24	Fabaceae	50.00	CP, NL, NGT
20.	Glycine max	Soybean	KB-79	Fabaceae	21.67	Μ
21.	Cajanus cajan	Pigeonpea	TTB-7	Fabaceae		
22.	Pisum sativum	Pea	KPMR-1	Fabaceae		
23.	Vigna radiate	Green gram	Pusa Baisaki	Fabaceae		
24.	Vigna mungo	Black gram	T-9	Fabaceae		
25.	Lablab purpureus	Field bean	HA-3	Fabaceae	26.67	Μ
26.	Macrotyloma uniflorum	Horse gram	PHG-9	Fabaceae	48.33	MM, MN, VT, RL, SP
27.	Phaseolus vulgaris	Common	Topcrop	Fabaceae		
		bean				
28.	Crotalaria juncea	Sunhemp	Local	Fabaceae		
29.	Cicer arietinum	Chickpea	A-1	Fabaceae		
30.	Vigna unguiculata	Cowpea	C-152	Fabaceae	26.67	Μ
31.	Cyamopsis tetragonoloba	Cluster bean	Pusa Navadhar	Fabaceae		
32.	Abelmoschus esculentus	Bhendi	Arke Komar	Malvaceae		
33.	Sesamum indicum	Sesamum	KL-31	Pedaliaceae		
34.	Capsicum annuum	Capsicum	Pusa Jwala	Solanaceae		
35.	Praecitrullus fistulosus	Round melon	Arka Tinda	Solanaceae		
36.	Lycopersicon esculentum	Tomato	Pusa Ruby	Solanaceae		
37.	Nicotiana tabacum	Tobacco	Xanthi	Solanaceae	45.00	MML
38.	Gossypium hirsutum	Cotton	Varalakshmi	Malvaceae		
39.	Solanum melongena	Brinjal	BB-12	Solanaceae		

CP = Chlorotic patches, DTL = Downward twisting of leaf, M = Mosaic, MM = Mild mosaic, MML = Mosaic mottling MN = Mild necrosis, NGT = Necrosis on growing bud, NL = Necrotic lesion, RL = Reduction in leaf size, SP= Severe puckering SG = Stunted growth, VT = Vein thickening

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Table 2: Transmission of SNV to crop plants through The space of			<i>rips palmi</i> Variety Family		Maan	Sumptom	
No.	Scientific name	Common Variety Family Mean transmissi		transmission	Symptoms		
		name			(%)		
1.	Helianthus annuus	Sunflower	KBSH-44	Asteraceae	26.67	CP, NL, DTL, VT, SG	
2.	Carthamus tinctorius	Safflower	C-18	Asteraceae			
3.	Guizotia abyssinica	Niger	S-26	Asteraceae			
4.	Tagetus erecta	Marigold	Chinthamani	Asteraceae			
5.	Zinnia elegans	Zinnia	ZE-231	Asteraceae			
6.	Brassica nigra	Mustard	Kanakpura local	Brassicaceae			
7.	<i>Brassica oleracea</i> var.	Knol khol	KO-O4-890	Brassicaceae			
	gongylodes						
8.	Brassica oleracea var Capitata	Cabbage	B0-01-001	Brassicaceae			
9.	Raphanus sativus	Radish	Mino Long	Brassicaceae			
10.	Cucumis sativus	Cucumber	PS –24	Cucurbitaceae	15.00	M, CP	
11.	Citrullus lanatus	Watermelon	Arka Manik	Cucurbitaceae	26.67	M, CP, NGT	
12.	Cucumis melo	Muskmelon	Deepti	Cucurbitaceae			
13.	Cucurbita moschata	Pumpkin	Arka	Cucurbitaceae	13.33	CP, NL	
			suryamukhi				
14.	Lagenaria siceraria	Bottle gourd	PSPR	Cucurbitaceae			
15.	Luffa acutangula	Ridge gourd	Arka sujata	Cucurbitaceae	12.00	M, CP	
16.	Momordica charantia	Bitter gourd	Arka Harit	Cucurbitaceae			
17.	Trichosanthes cumerina	Snake gourd	C0-2	Caricaceae			
18.	Ricinus communis	Castor	DCS-9	Euphorbiaceae			
19.	Arachis hypogaea	Groundnut	JL-24	Fabaceae	26.67	CP, NL, NGT	
20.	Glycine max	Soybean	KB-79	Fabaceae			
21.	Cajanus cajan	Pigeonpea	TTB-7	Fabaceae			
22.	Pisum sativum	Pea	KPMR-1	Fabaceae			
23.	Vigna radiata	Green gram	Pusa Baisaki	Fabaceae			
24.	Vigna mungo	Black gram	T-9	Fabaceae			
25.	Lablab purpureus	Field bean	HA-3	Fabaceae	06.67	М	
26.	Macrotyloma uniflorum	Horse gram	PHG-9	Fabaceae	23.33	MM, MN, VT, SP, RL	
27.	Phaseolus vulgaris	Common	Topcrop	Fabaceae			
		bean					
28.	Crotalaria juncea	Sunhemp	Local	Fabaceae			
29.	Cicer arietinum	Chickpea	A-1	Fabaceae			
30.	Vigna unguiculata	Cowpea	C-152	Fabaceae	16.67	М	
31.	Cyamopsis tetragonoloba	Cluster bean	Pusa Navadhar	Fabaceae			
32.	Abelmoschus esculentus	Bhendi	Arke Komar	Malvaceae			
33.	Sesamum indicum	Sesamum	KL-31	Pedaliaceae			
34.	Capsicum annuum	Capsicum	Pusa Jwala	Solanaceae			
35.	Praecitrullus fistulosus	Round melon	Arka Tinda	Solanaceae			
36.	Lycopersicon esculentum	Tomato	Pusa Ruby	Solanaceae			
37.	Nicotiana tabacum	Tobacco	Turkish	Solanaceae	10.00	MML	
38.	Gossypium hirsutum	Cotton	Varalakshmi	Malvaceae			
39.	Solanum melongena	Brinjal	BB-12	Solanaceae			

CP = Chlorotic patches, DTL = Downward twisting of leaf, M = Mosaic, MM = Mild mosaic, MML = Mosaic mottling MN = Mild necrosis, NGT = Necrosis on growing bud, NL = Necrotic lesion, RL = Reduction in leaf size, SP= Severe puckering SG = Stunted growth, VT = Vein thickening

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Table 3: Transmission of SNV to weed species through sap inoculation						
Sr. No	Scientific name	ies Common name	_ Family	Mean transmission	Symptoms	
1.	Lagascea mollis	Sickle leaf	Asteraceae	21.00	MM	
2.	Parthenium hysterophorus	Congress grass	Asteraceae			
3.	Sonchus oleraceous	Annual sowthistle	Asteraceae			
4.	Alternanthera sessilis	Sessile joyweed	Amaranthaceae	20.00	М	
5.	Amaranthus spinosus	Spiny amaranth	Amaranthaceae			
6.	Commelina benghalensis	Benghal dayflower	Commelinaceae	16.00	М	
7.	Ipomoea pandurata	Bigroot morning glory	Convulvulaceae			
8.	Crotalaria spectabilis	Showy crotalaria	Euphorbiaceae	20.00	М	
9.	Euphorbia hirta	Asthama weed	Euphorbiaceae	18.00	M, MML	
10.	Cassia obtusifolius	Sickle pod	Fabaceae	20.00	CL	
11.	Ocimum sanctum	Holi basil	Labiatae	20.00	CL	
12	Sida rhombifolia	Arrowleaf sida	Malvaceae	16.00	M	
13.	Oxalis corniculata	Creeping wood sorrel	Oxalidaceae	15.00	M	
14.	Portulaca oleraceae	Common purslane	Portulaceae			
15.	Physalis minima	Wild capegooseberry	Solanaceae	20.00	CL	
16.	Galinsoga parviflora	Ouick weed	Asteraceae	42.00	CL	
17.	Eupatorium odoratum	Siam weed	Asteraceae			
18.	Euphorbia geniculata	Mexican fireplant	Euphorbiaceae	45.00	CL	
19.	Solanum nigrum	Black night shade	Solanaceae	25.00	MM	
20.	Phyllanthus niruri	Seed under the leaf	Euphorbiaceae	30.00	ММ	
21.	Malvestrum coromandelianum	False mellow	Malvaceae	45.00	CL. VC	
22.	Ageratum convzoides	Goat weed	Asteraceae	45.00	CL	
23.	Polygonium plebjum	Spring weed	Polygonaceae			
24.	Datura stramonium	Jimson weed	Solanaceae			
25.	Tridax procumbense	Tridax daisy	Asteraceae			
26.	Achyranthes aspera	Prickly achyranthes	Amaranthaceae	48.00	M, CL, RL	
27.	Abutilon indicum	Monkey bush	Malvaceae	25.00	CL	
28.	Ocimum canum	Hairy basil	Labiatae	20.00	MM	
29.	Argemone mexicana	Mexican poppy	Papaveraceae			
30.	Crotalaria striata	Smooth crotalaria	Fabaceae	30.00	CL	
31.	Bidens vilosa	Spanish needle	Asteraceae	15.00	ММ	
32.	Borreria stricta	Broad button weed	Rubiaceae			
33.	Stachytarpeta indica	Nettleleaf vervain	Verbenaceae	18.00	CL. NL	
34.	Leucas aspera	Dead white nettle	Labiatae			
35.	Acanthospermum hispidum	Hispid starrburr	Asteraceae	20.00	MNL	
36.	Synedrella nodiflora	Nodeweed	Asteraceae			
37.	Xanthium strumarium	Cocklebur	Asteraceae	30.00	М	

CL – Chlorosis of leaf, M – Mosaic, MM – Mild mosaic, MML – Mosaic mottling, MNL – Marginal necrosis of leaves NL – Necrosis of leaf, RL – Reduction in leaf size, VC – Vein clearing

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Table 4 : Transmission of SNV to weed species through Thrips palmi						
Sr. No	Scientific name	Weed species me Common name		Mean transmission	Symptoms	
1.	Lagascea mollis	Sickle leaf	Asteraceae	18.00	MM	
2.	Parthenium hysterophorus	Congress grass	Asteraceae			
3.	Sonchus oleraceous	Annual sowthistle	Asteraceae			
4.	Alternanthera sessilis	Sessile joyweed	Amaranthaceae	15.00	М	
5.	Amaranthus spinosus	Spiny amaranth	Amaranthaceae			
6.	Commelina benghalensis	Benghal dayflower	Commelinaceae	12.00	М	
7.	Ipomoea pandurata	Bigroot morning glory	Convulvulaceae			
8.	Crotalaria spectabilis	Showy crotalaria	Fabaceae	10.00	М	
9.	Euphorbia hirta	Asthama weed	Euphorbiaceae	13.00	M, MML	
10.	Cassia obtusifolius	Sickle pod	Fabaceae	15.00	CL	
11.	Ocimum sanctum	Holi basil	Labiatae	18.00	CL	
12.	Sida rhombifolia	Arrowleaf sida	Malvaceae	10.00	М	
13.	Oxalis corniculata	Creeping wood sorrel	Oxalidaceae	10.00	М	
14.	Portulaca oleraceae	Common purslane	Portulaceae			
15.	Physalis minima	Wild capegooseberry	Solanaceae	14.00	CL	
16.	Galinsoga parviflora	Quick weed	Asteraceae	25.00	CL	
17.	Eupatorium odoratum	Siam weed	Asteraceae			
18.	Euphorbia geniculata	Mexican fireplant	Euphorbiaceae	23.00	CL	
19.	Solanum nigrum	Black night shade	Solanaceae	14.00	MM	
20.	Phyllanthus niruri	Seed under the leaf	Euphorbiaceae	11.00	MM	
21.	Malvestrum coromandelianum	False mellow	Malvaceae	18.00	CL, VC	
22.	Ageratum conyzoides	Goat weed	Asteraceae	15.00	CL	
23.	Polygonium plebjum	Spring weed	Polygonaceae			
24.	Datura stramonium	Jimson weed	Solanaceae			
25.	Tridax procumbense	Tridax daisy	Asteraceae			
26.	Achyranthes aspera	Prickly achyranthes	Amaranthaceae	15.00	M, CL, RL	
27.	Abutilon indicum	Monkey bush	Malvaceae	15.00	CL	
28.	Ocimum canum	Hairy basil	Labiatae	12.00	MM	
29.	Argemone mexicana	Mexican poppy	Papaveraceae			
30.	Crotalaria striata	Smooth crotalaria	Fabaceae	10.00	CL	
31.	Bidens pilosa	Spanish needle	Asteraceae	15.00	MM	
32.	Borreria stricta	Broad button weed	Rubiaceae			
33.	Stachytarpeta indica	Nettleleaf vervain	Verbenaceae	10.00	CL, NL	
34.	Leucas aspera	Dead white nettle	Labiatae			
35.	Acanthospermum hispidum	Hispid starrburr	Asteraceae	10.00	MNL	
36.	Synedrella nodiflora	Nodeweed	Asteraceae			
37.	Xanthium strumarium	Cocklebur	Asteraceae	15.00	М	

CL – Chlorosis of leaf, M – Mosaic, MM – Mild mosaic, MML – Mosaic mottling, MNL – Marginal necrosis of leaves NL – Necrosis of leaf, RL – Reduction in leaf size, VC – Vein clearing

necrotic lesions was noted on *Malvestrum* coromandelianum and Stachytarpeta indica, respectively (Table 3 and 4).

The results are similar to the report of Lavanya *et al.* (2005), who reported that SNV has wide host range comprising of 15 plant species belonging to Fabaceae, three plant species belonging to Malvaceae, six plant species belonging to Cucurbitaceae, three plant species belonging to Solanaceae and one each belonging to Cruciferaceae and Moringaceae.

Thus, by the above study it can be concluded that these nine plants and twenty four weeds carry virus inoculum and harbour thrips during offseason thereby acting as collateral hosts for both virus and vector. Further, it can be said that these plants also help in perpetuation of thrips, survival and spread of the virus..

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