# Effect of bavistin foliar spray on VAM colonization and growth parameters in six groundnut cultivars

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The impact of application of plant protection chemical bavistin as foliar spray on AM colonization and plant growth parameters like root length, shoot length, shoot fresh weight, shoot dry weight, nodule number and nodule weight of six groundnut cultivars *viz.*, 87779 ICGV, CS 52, FDRS 4, 414332, Gangapuri and J 11 was studied. For the study, the cultivars were raised in field plots. The phosphorus and nitrogen contents of plant material was also estimated. The bavistin spray significantly decreased the per cent infection and number of spores in roots of six cultivars at 50days, 70days and 90days samplings. Except the shoot dry weight at 70days and 90days samplings, all the other parameters studied including 'P' and 'N' contents were negatively affected by the bavistin foliar spray.

Key words : Bavistin, Groundnut cultivars, AM colonization.

#### INTRODUCTION

The tripartite associations, in legumes, formed of plants, L nodules and endomycorrhizas have potential for increasing the crop yields. Among the biological processes involved in the rhizoplane, the unique role of symbiotic bacteria and the AM fungi which ensure fixation and mobilization and availability of nitrogen and phosphorus of plants have been well recognized. It is also a wellestablished fact that the AM always prefer certain host exhibiting maximum symbiotic response and increased the growth and yield of crop mainly through improved uptake of nutrients (Allen, 1991) and especially through uptake of phosphorus in soils of low fertility (Smith and Read, 1997). Most agricultural soils possess an indigenous VAM spore strains, the role of which in crop productivity has been examined in sufficient details (Tilak, 1993). Changing agricultural practices have resulted in increased application of pesticides and fungicides for the control of diseases. There is an increasing awareness on the deleterious effects of these chemicals on the beneficial mycorrhizal fungi (Bailey and Safir, 1978). But the reports on the effect of fungicides or insecticides on VAM fungi when applied as foliar sprays are few, though the practice of spraying agricultural chemicals on crops is so common.

## MATERIALS AND METHODS

The investigation was carried out by raising the six cultivars of groundnut in field plots in split-plot design. A field plot of size 16 m x 7 m in the University botanical and experimental garden was selected for the study. The

plot was divided into three rows of three blocks each with block size of 5 m x 2 m. In each block, seeds of six groundnut cultivars *viz.*, 87779 ICGV, CS 52, FDRS 4, 414332, Gangapuri and J 11 were sown in three rows of each. Bavistin at a concentration of 1 g/lit and water were sprayed on to foliage as fungicide treatment and control, respectively. The foliar sprays were carried out thrice, at twenty days interval, starting from 40<sup>th</sup> day after sowing of seeds.

The plant samplings were collected at 50th, 70th and 90th days after sowing. For each cultivar, three plants were sampled from each replicate of the treatment and control. The root samples of treatment and control were collected, processed and stained in 0.1% trypan blue prepared in lactophenol (Philips and Hayman, 1970). The per cent root colonization was calculated by the method of Giovanetti and Mosse (1980). At each sampling time, shoot fresh weight, shoot dry weight, nodule number, nodule weight, root length, shoot length were recorded. The phosphorus content of shoots was estimated by the Vanado-molybdate yellow method (Jackson, 1973). The per cent nitrogen in the plant material was determined by micro-kjeldahl 'N' method (AOAC, 1978). The data of all parameters at each sampling was subjected to statistical analysis to find out the F-value due to treatments as well as cultivars.

## **RESULTS AND DISCUSSION**

The bavistin foliar spray significantly decreased the per cent infection and number of vesicles or spores in all the

six cultivars and at all sampling stages (Table 1 and 2). Greater reduction in infection over control was observed in the cultivar CS 52 at all the three stages due to bavistin spray. The degree of reduction in per cent infection and number of vesicles or spores varied among the cultivars. Gunasekaran *et al.* (1988) observed 89% and 77% reduction in mycorrhizal infection of cowpea roots due to

application of two systemic fungicides, benomyl and bavistin as soil drench. The plant parameters *viz.*, root length, shoot length, shoot fresh weight, shoot dry weight, nodule number, nodule fresh weight, per cent phosphorus and per cent nitrogen were negatively affected by bavistin spray except the shoot dry weight at 70 days and 90 days samplings (Table 3, 4, 5, 6, 7, 8 and 9). The reduction in

Cultivars	Per cent infection							
	50	days	70	days	90 days			
	Control	Sprayed	Control	Sprayed	Control	Sprayed		
877791 ICGV	46.6	41.6	55.0	46.6	67.0	56.3		
CS 52	43.3	40.0	55.0	47.5	64.0	55.0		
FDRS 4	48.3	43.3	60.0	48.3	71.6	58.6		
414332	45.0	41.6	58.6	50.0	70.3	60.0		
Gangapuri	40.0	36.6	51.6	41.5	66.3	54.6		
J 11	53.3	48.3	62.3	51.6	75.6	63.3		
F value due to								
treatments :	93	.70 <sup>*</sup>	$20.17^{*}$		$21.37^{*}$			
cultivars :	139	9.95*	207	'.59 <sup>*</sup>	281	.15*		

\* indicates significance of value at P=0.05

	No. of vesicles / spores ( $cm^{-1}$ root bit)						
Cultivars	50 0	days	70 0	lays	90 d	ays	
	Control	Sprayed	Control	Sprayed	Control	Sprayed	
877791 ICGV	32.0	28.0	41.0	33.0	56.0	36.0	
CS 52	31.0	26.0	44.0	30.0	63.0	42.0	
FDRS 4	42.0	31.0	52.0	38.0	71.0	44.0	
414332	39.0	28.0	51.0	37.0	68.0	50.0	
Gangapuri	27.0	22.0	36.0	28.0	54.0	34.0	
J 11	49.0	38.0	60.0	46.0	83.0	62.0	
F value due to							
treatments :	30.	26*	23.	44*	45.8	$30^{*}$	
cultivars :	14.	$72^{*}$	90.	$00^{*}$	286.	95 <sup>*</sup>	

\* indicates significance of value at P=0.05

			Root le	ength (cm)		
Cultivars	50	days	70 0	lays	90 days	
	Control	Sprayed	Control	Sprayed	Control	Sprayed
877791 ICGV	12.5	12.2	12.7	11.2	14.3	11.9
CS 52	13.1	11.5	13.2	12.3	14.1	11.9
FDRS 4	12.7	10.7	13.3	11.9	14.3	11.6
414332	12.5	11.6	12.5	10.9	15.5	11.6
Gangapuri	12.7	10.8	12.7	11.7	14.1	12.0
J 11	12.2	9.8	12.7	9.3	14.5	11.4
F value due to						
treatments :	1.6	55	1.9	99	0.4	15
cultivars :	19.	.39*	97.	43*	97.4	43 <sup>*</sup>

\* indicates significance of value at P=0.05

plant dry matter production due to bavistin foliar spray may be because of its deleterious effect on VAM fungi rather than its direct effect on plant growth as brought out by the reduced VAM development in the treated plants.

The reduction in plant growth and 'N' and 'P' contents was found considerable in bavistin treatment.

By and large, the systemic fungicides as a group appear to be more damaging to mycorrhizal symbiosis than nonsystemic ones (Jalali, 1990). And by virtue of their nature, systemic fungicides provide an undesirable environment inside the plant system to the invading organisms. Jalali and Domsch (1975) observed that seed as well as foliar applications with conventional and systemic fungal

Table 4 : Effect of ba	avistin spray on sho	oot length of six c	ultivars of ground	nut		
			Shoot l	ength (cm)		
Cultivars	50 0	days	70 0	days	90 d	ays
	Control	Sprayed	Control	Sprayed	Control	Sprayed
877791 ICGV	32.7	28.9	48.2	44.2	74.0	59.2
CS 52	33.6	29.9	56.2	41.9	73.9	58.0
FDRS 4	36.8	36.2	56.0	48.4	74.4	56.3
414332	26.5	25.1	41.4	39.0	59.9	41.3
Gangapuri	47.3	45.1	77.5	70.5	88.0	85.7
J 11	32.1	28.9	50.4	47.0	75.7	58.3
F value due to						
treatments :	116	6.71 <sup>*</sup>	28.	$48^{*}$	14.11*	
cultivars :	21	.69*	13.	14*	33.4	43 <sup>*</sup>

\* indicates significance of value at P=0.05

			Shoot fresh	n weight (g/pl)		
Cultivars	50 0	days	70 0	lays	90 d	ays
	Control	Sprayed	Control	Sprayed	Control	Sprayed
877791 ICGV	50.01	39.52	86.60	51.82	108.93	84.75
CS 52	41.22	33.53	75.88	71.33	106.01	70.85
FDRS 4	41.33	35.27	53.95	45.13	66.61	59.80
414332	39.87	36.38	68.33	61.05	82.72	80.00
Gangapuri	47.65	36.85	68.22	56.45	70.46	64.42
J 11	35.71	31.15	53.41	41.36	69.62	54.03
F value due to						
treatments :	6.	.79*	3.	73	5.3	34
cultivars :	33.	.29*	8.	75*	8.6	55 <sup>*</sup>

\* indicates significance of value at P=0.05

			Shoot dry	weight (g/pl)		
Cultivars	50 0	days	70 0	70 days		ays
	Control	Sprayed	Control	Sprayed	Control	Sprayed
877791 ICGV	10.66	7.63	20.64	10.92	25.99	17.88
CS 52	9.08	8.24	16.08	15.21	22.46	15.11
FDRS 4	9.11	7.49	12.48	10.57	15.42	14.0
414332	8.09	7.75	14.73	14.30	18.83	17.83
Gangapuri	9.66	9.12	14.36	12.12	14.83	13.82
J 11	7.13	5.97	12.10	8.42	15.79	11.03
F value due to						
treatments :	4.35		1.73		3.23	
cultivars :	9.	81*	5.	07*	5.8	34 <sup>*</sup>

\* indicates significance of value at P=0.05

Table 7 : Effect of 1	oavistin spray on no	dule number in si	x cultivars of grou	ındnut		
			Nodule nu	mber (Plant <sup>-1</sup> )		
Cultivars	50	days	70 0	days	90 d	ays
	Control	Sprayed	Control	Sprayed	Control	Sprayed
877791 ICGV	98.7	82.6	110.9	94.6	137.6	101.4
CS 52	91.9	58.0	95.6	70.6	131.2	76.6
FDRS 4	108.3	68.3	114.1	73.5	116.4	77.5
414332	94.2	68.5	117.9	70.5	175.4	102.0
Gangapuri	99.7	73.2	111.2	81.6	121.6	82.4
J 11	58.0	49.0	76.4	56.8	96.1	77.9
F value due to						
treatments :	5.	92	4.	51	4.	20
cultivars :	29.	.68*	36.	.08*	35.	$12^{*}$

\* indicates significance of value at P=0.05

Table 8 : Effect of ba	Table 8 : Effect of bavistin spray on nodule fresh weight in six cultivars of groundnut							
		Nodule fresh weight (mg/50 tap root nodules)						
Cultivars	50	days	70 -	days	90 d	lays		
	Control	Sprayed	Control	Sprayed	Control	Sprayed		
877791 ICGV	293.25	221.89	327.29	226.02	336.45	260.58		
CS 52	304.01	272.56	359.54	282.84	425.41	337.18		
FDRS 4	307.33	215.00	332.12	269.02	373.35	275.51		
414332	377.20	305.49	395.12	323.56	445.83	341.17		
Gangapuri	325.25	226.14	328.02	231.07	372.07	280.68		
J 11	330.09	307.29	410.12	356.82	356.82	379.64		
F value due to								
treatments :	4.	53	21	.50*	57.4	$40^{*}$		
cultivars :	25	.69*	101	.58*	312.	.52*		

\* indicates significance of value at P=0.05

Table 9 : Effect of bavistin spray on phosphorus and nitrogen contents of shoots in six cultivars of groundnut at 70 days after sowing							
Cultivars	% Pho	sphorus	% Ni	trogen			
Cultivals	Control	Sprayed	Control	Sprayed			
877791 ICGV	0.466	0.438	3.099	2.623			
CS 52	0.465	0.404	2.893	2.753			
FDRS 4	0.474	0.368	2.982	2.459			
414332	0.386	0.297	2.767	2.660			
Gangapuri	0.448	0.378	2.800	2.581			
J 11	0.448	0.398	2.977	2.660			
F value due to							
treatments :	35.	.640*	19.4	439 <sup>*</sup>			
cultivars :	cultivars : 7.689 <sup>*</sup> 0.536 <sup>*</sup>						

\* indicates significance of value at P=0.05

toxicants restricted the development of mycorrhizal endophytes on host roots in wheat. They also postulated that since foliarly applied pesticides may not be translocated intact to the roots, but the side effects on mycorrhizae may be brought about by changes in the spectrum of root exudates as a result of the stress exerted

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by the pesticides. A changed pattern of amino acid exudation was also observed when systemic fungicides namely triforine and tridemorph were applied to the wheat foliage (Jalali and Domsch, 1977). Oliveira *et al.* (1987) studied the effect of spraying benomyl, captafol, copper oxychloride on VAM colonization of Rangpur lime and observed that, except captafol the others reduced mycorrhizal infection and consequently growth stimulus of the inoculated plants. In conclusion, it is necessary to evaluate the impact of foliar sprays with plant protection chemicals on VAM development in crop plants which form mycorrhizal association in the field.

#### References

- Allen, F. (1991). *Ecophysiology of Mycorrhiza*, Oxford Press, London, 249pp.
- AOAC. (1978). Official and tentative methods of Analysis. Association of Official Agricultural Chemists. Washington, D.C.
- Bailey, J.E. and Safir. (1978). The effect of benomyl on soybean endomycorrhizae. *Phytopathol.*, **68** : 1810-1812.

- Giovanetti, M. and Mosse, B. (1980). An evaluation of technique for measuringvesicular-arbuscular mycorrhizal infection in the plants. *New Phytol.*, **95** : 69-82.
- Gunasekaran, P., Ubalthoose Raja, N. and Sundaresan, P. (1988). Toxic effects of some agricultural fungicides on the mycorrhizal association of cowpea. In: *Mycorrhiza Round Table*. Proc. of workshop (Varma, A.K., Oka, A.K., Mukerji, K.G, Tilak, K.V.B.R. and Janak Raj efs.), New Delhi : 550-559.
- Jackson, M.L. (1973). *Soil chemical analysis*. Prentice Hall of India Pvt., New Delhi, pp. 370-387.
- Jalali, B.L. (1990). Side effects of pesticides on mycorrhizal system – an over view. In: *Current Trends in Mycorrhizal Research*. Proc. of Nat. Conf. on Mycorrhiza (Jalali, B.L. and Chand, M. eds.), Hisar : 172-174.
- Jalali, B.L. and Domsch, K.H. (1975). Effect of systemic fungitoxicants on the development of endotropic mycorrhiza. In : *Endomycorrhizas* (Sanders, F.E., Mosse, B. and Tinker, P.B. eds.), Academic press, London: 619-626.

- Jalali, B.L. and Domsch, K.H. (1977). Effect of some fungitoxicants on the amino acid spectrum of root exudates in relation to endomycorrhizal development. *Phytopathol. Z.*, **90** : 22-27
- Oliveira, A.A.R., Jesus, I.S.De. and Campose, E.D. (1987). Effect of applying 3 fungicides on mycorrhization of Rangpur lime. *Fitopatologia Brasileira*, **12**(1): 57-60.
- Phillips, J.M. and Hayman, D.S. (1970). Improved procedures for clearing roots and staining parasitic vesicular arbuscular mycorrhizal fungi for rapid assessment of infection. *Trans. Br. Mycol. Soc.*, **55** : 158-161.
- Smith, S.E. and Read, D.J. (1997). *Mycorrhizal Symbiosis*. Academic press California; 298 pp
- Tilak, K.V.B.R. (1993). Associative effects of VA Mycorrhizae with nitrogen fixers. *Proc. Nat. Sc. Acad.*, B59 (3 and 4): 325-332.