

Progressive changes in oxidative enzymes and some biochemical constituent of chickpea genotype under salinity stress

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SUMMARY

A pot culture experiment was conducted to study the progressive stress responses and mechanism of salinity stress tolerance in chickpea genotype. Two chickpea genotype (one tolerant and one susceptible) CSG-8962 and Vijay of different adaptation were taken for the study in control and saline stress (150mM NaCl) condition at interval of 7, 14 and 21 days. The sample was analyzed for the levels of peroxidase and polyphenol oxidase enzymes and some of the key biomolecules like reducing sugars, soluble protein, proline, polyphenol and free amino acids to find out the biochemical markers involved in identifying the salt tolerance in chickpea cultivar. The result revealed that the activity of oxidative enzyme peroxidase and polyphenol oxidase, proline, free amino acids and polyphenol were found to be increased comparatively higher in salt tolerance cultivar than the susceptible cultivar under the salinity stress situation. Where the soluble protein content in salt susceptible cultivar decreased with salinity stress.

Key words : Chickpea, Peroxidase, Polyphenol oxidase, Proline, Soluble protein, Polyphenol reducing sugar, Free amino acids and salinity stress

Chickpea (*Cicer arietinum* L.) is the world's second most important pulse crop next only pigeonpea and cultivated in more than 41 countries occupying approximately 15 percent of the total pulse area in the world (Datta, 2002). Among the various stress condition salinity causing losses quantitatively in chickpea is one of the major. Chickpea is cool-season legumes grown extensively through out world (12.03M ha), particularly in the India subcontinent, countries of North Africa, North America, West Asia and Mediterranean region (Anonymous, 1999). In India, it is grown on 8.40 Mha areas. Legumes have been found to be highly susceptible to words saline environment, which ultimately manifest in reduced growth and metabolism. Abiotic stress is known to disturb the intracellular water balance of the biological organisms. To counteract such condition, plant accumulates various low molecular weight compounds such as sugar, sugar alcohols amino acids and quaternary ammonia compounds. These metabolites, widely known as compatible solutes or osmolytes are neutral, non-toxic

and do not interfere with normal metabolic reaction even in high concentration (Pujni *et al.*, 2007). Salt tolerance mechanisms include array of characters, alterations and progressive adaptation in tolerance type and any flow that reflect on the metabolic failures resulting in sensitive behavior (Vasanth and Rajalakshmi, 2009). With these objective of studying progressive changes in chickpea and experiment was designed with tolerant and susceptible variety.

MATERIALS AND METHODS

The chickpea cultivar CSG-8962 and Vijay are available at all India Co-ordinated Pulses Improvement Project MPKV Rahuri, were used for the present investigation. The seeds were surface sterilized with aqueous solution of 0.1% mercuric chloride and then thoroughly washed with distilled water. The seeds were soaked in cold water and placed in germinating paper for even germination for 2 days. Uniformly germinated seed were sowed in pots filled with soil. Two replication were maintained for each cultivar and treatment. When the seedling reaches 3-4 leaf stage salinity stress of 150mM NaCl was imposed. The shoot of both cultivar were analyzed at 7, 14 and 21 days after imposition of salinity treatment. During the growing of plant the water level was maintained with distilled and saline water respectively. The fresh shoot samples were collected for analysis of biochemical analysis. The proline contains in the sample were determine by the Bates *et al.* (1975). The soluble protein was determination by the procedure of Lowry *et*

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al. (1951). The free amino acids were estimated by method of Rosen (1957). Reducing sugar was determined by Nelson Somagy (1944). Peroxidase and polyphenol oxidase enzyme activity was measured spectrophotometrically by monitoring the increase in absorbance at 420 nm and polyphenol by Swain and Hills, (1959).

RESULTS AND DISCUSSION

The data presented in figures on change in the oxidative enzymes and biochemical constituent under salinity stress in chickpea genotypes. The oxidative enzymes *viz.* peroxidase and polyphenol oxidase were increased under the influence of salinity. Peroxidase activity was increased from 7 to 21 days (Fig.1) and increased more in tolerant cultivar than susceptible. A significant increase in peroxidase activity influence of salinity was recorded for wheat and rice (Sgherri *et*

al., 2000 and Srivalli., 2003), Polyphenol oxidase activity increased under influence of salinity. The varietal behavior different under salinity. In CSG-8962, polyphenol oxidase activity increased gradually from 7 to 21 days (Fig. 2) while in Vijay an increase was slow. The result is similar with Hernandez *et al.* (2000).

Proline is a compatible osmolyte that accumulates in greater proportion under abiotic stress. Proline content increases in all days under salinity in both varieties (Fig. 3) but increase was higher in salt tolerant cultivar CSG 8962 as compared to Vijay. Same trend was observed in sugarcane cultivar under salinity stress (Vasantha and Rajalakshmi, 2009) in the sugarcane cultivar. Salt tolerant cultivar of barley accumulated over 80% proline than control and the levels increased only 40% in a sensitive cultivar (Kumar *et al.*, 1981). Damame *et al.* (2008) also proposed that the higher increase in proline content was stimulated by salt in the tolerant cultivar than susceptible in chickpea. Proline accumulation remained higher under stress condition and accumulation helps to maintain turgor and promotes continued growth under moisture stress condition (Pankaj kumar *et al.*, 2006). In the present study increased trend of proline content with the stress suggests its protective and stabilized role under stress.

The soluble protein content was significantly decreased in salinity condition and decrease was more in Vijay cultivar (Fig.4). On the other hand, soluble protein content in salt susceptible cultivar decreased in salinity condition, these results are in agreement with those of Garg, (2002) and Agarwal and Pandey (2003). The polyphenol content was increased in both cultivars in both control and saline condition but the increase was higher in tolerant cultivar in saline situation (Fig.5). Damame *et al.* (2008) reported induced salinity increases polyphenol content in chickpea seedling.

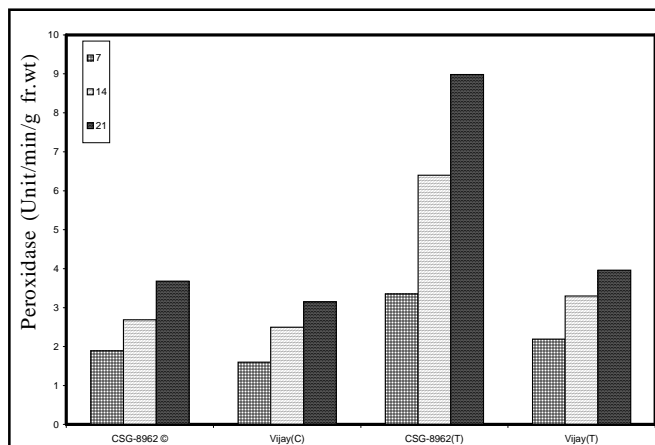


Fig.1 : Progressive change in peroxidase activity in chickpea as influenced by salinity at 7, 14 and 21 days, C and T indicate control and salinity treatment

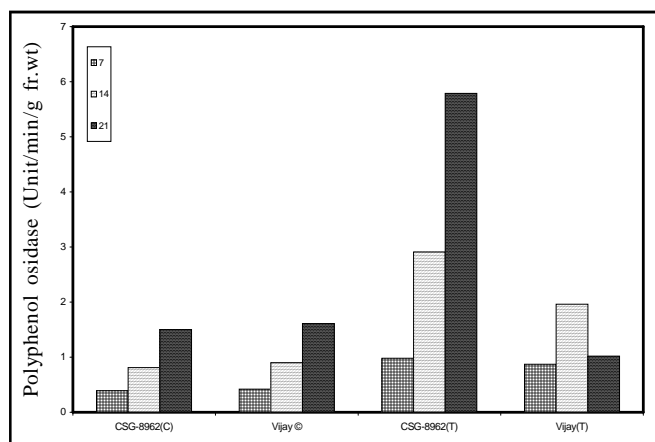


Fig.2 : Polyphenol oxidase activity in chickpea as influenced by salinity at 7, 14 and 21 days, C and T indicate control and salinity treatment

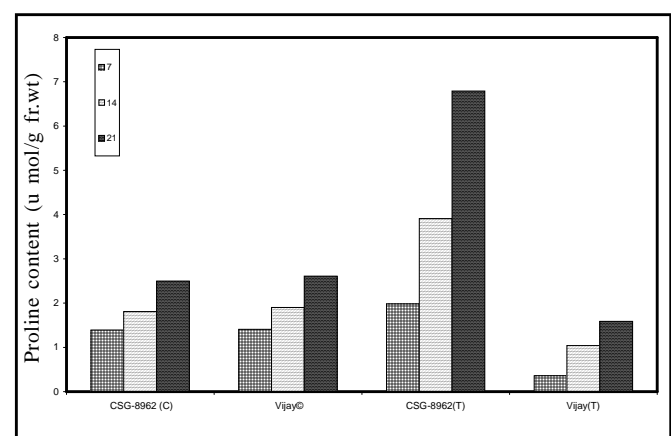


Fig.3 : Proline content in chickpea as influenced by salinity at 7, 14 and 21 days, C and T indicate control and salinity treatment

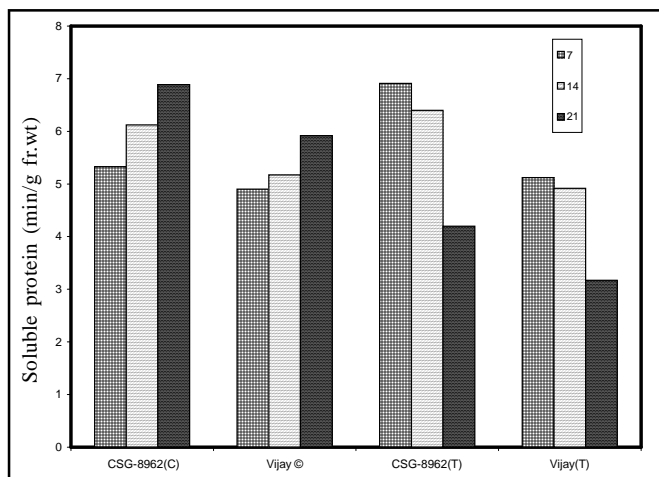


Fig.4 : Soluble protein in chickpea as influences by salinity at 7, 14 and 21 days, C and T indicate control and salinity treatment

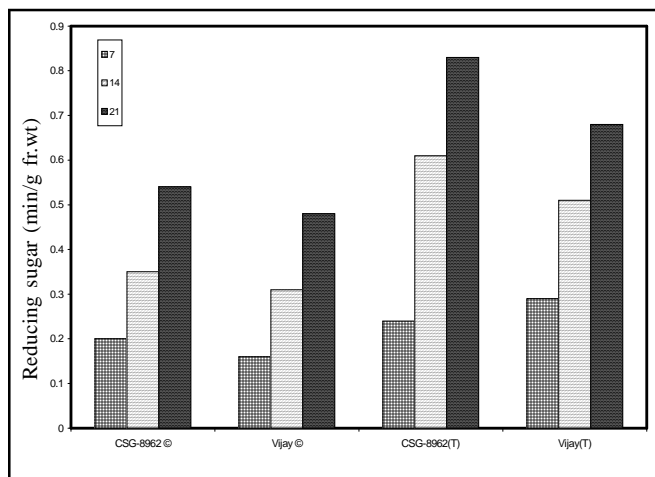


Fig.6 : Reducing sugar conten in chickpea as influences by salinity at 7, 14 and 21 days, C and T indicate control and salinity treatment

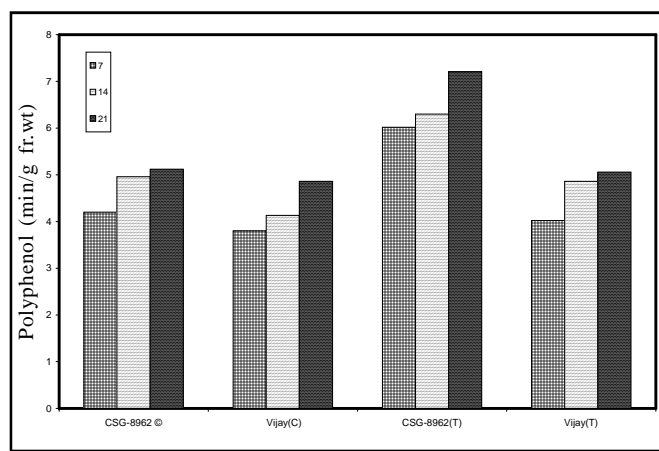


Fig.5 : Polyphenol activity in chickpea as influences by salinity at 7, 14 and 21 days, C and T indicate control and salinity treatment

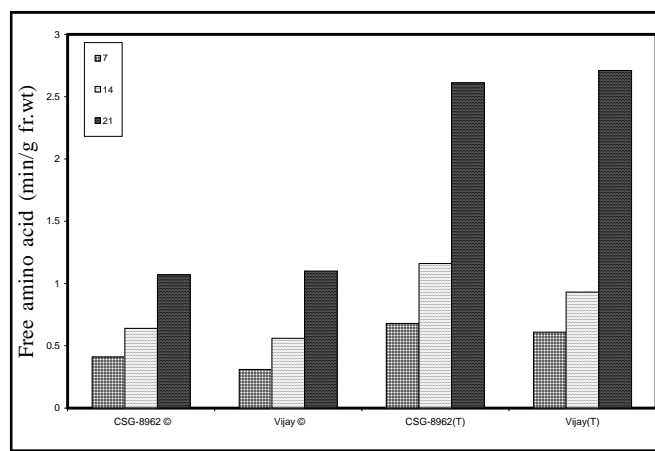


Fig.7 : Free amino acid activity in chickpea as influences by salinity at 7, 14 and 21 days, C and T indicate control and salinity treatment

The reducing sugar was higher in tolerant cultivar CSG-8962 in salinity situation, but the increase was observed in both cultivars in both situations(Fig.6). Cram, (1976) was view that of the major osmotic adjustment in glycophytes subject to salinity environment. Osmotic adjustment in plant exposed to salinity stress depends largely on soluble sugar. The accumulation of sucrose in plants has been widely reported as a response to salinity or drought (Ashraf, and McNeilly, 2004). Free amino acid content was also increase in both cultivar in both situation but it was higher in susceptible cultivar in both situation

but was higher in susceptible cultivar(Fig.7). Free amino acid play vital role in salinity stress and free amino acid are known to be increases significantly in 8 canola line with increase in salt concentration of the growth medium is reported by Qasim, (2000) is similar with present study.

It can be concluded from the present study that the oxidative enzymes peroxidase and polyphenol oxidase and biochemical constituent like proline, soluble protein, reducing sugar, polyphenol and free amino acid appears to be a predominant biochemical indicator in chickpea genotype to study the tolerance mechanisms.

REFERENCES

Agarwal, S. and Pandey, V. (2003). Water and UV-B dependent oxidative stress: Effect on anti-oxidative content in *Cassia angustifolia*. *Indian J. Plant Physiology (special issue)*, 298-302.

Anonymous (1999). *FAO Production Year Book*, FAO Rome
 Bates, L.S., Waldren, R.P. and Teare D. 1975. Rapid Determination of free proline from water stress studies. *Plant & Soil*, 39 : 205-207.

- Ashraf, M. and mcNeilly, T. ,(2004). Salinity tolerance in *Brassica* oilseeds. *Critical Review in Plant Sci.*, **23** : 157-174.
- Bates, L.S., Waldern, R.P. and Teak, T.D. (1973). Rapid determination of free praline for water stress studies. *Plant & Soil*, **39**: 205-207
- Cram, W.J.(1976). Negative feedback regulation of transport in cell. *The Maintenance of Plant Physiology, New Series*, **2** : 284-316
- Damame, S.V., Lokhande, P.K. and Dalvi, U.S. (2008). Biochemical response to salt in chickpea seedling. *Indian J. Environ. & Ecoplan.*, **15** : 413-414
- Datta, S.K. (2002). Recent development in transgenic for abiotic stress tolerance in rice JIRCAS work Report, 43.
- Garg, N. (2002). Salinity stress- induced changes in key metabolites in the nodules of *Glycine max* L. (Soybean) and *Cicer arietinum* L (Chikpea) and manoeuvradility of their response through plant growth regulation. *J.Plant Bio.*, **29**: 137-142
- Hernanez, J., Jimenez, A., Mullineaux, P. and Sevilla, F. (2000). Tolerance of pea plant (*Pisum sativum*) to long term salt stress is associated with induction of antioxidant defenses. *Plant Cell Environ.*, **23** : 853-862.
- Kumar, A.P., Arun Kumar, S.N., Masih and Shemshery, P. (1981). Tolerance of some barley varieties to salt stress at seedling stage. *Indian J. Plant Physiol.*, **24**: 304-311.
- Lowry, O.H. , Rosenbrough, N.J., Farr, A.L. and Randall, R.J. (1951) . *J. Biol. Chem.*, 193 265.
- Pankaj Kumar, P.S., Deshmukh, R.K., Sairam, S.R., Kushwaha and Tej Pal Singh (2006). Biochemical and phenological evaluation of chickpea genotypes differing in drought tolerance. *Indian J. Plant Physiol.*, **11** : 166-171.
- Pujni, D., Chudary A. and Rajam, M.V.(2007). Increase tolerance to salinity and drought in transgenic Indian rice by mannitol accumulation *J. Plant Biochem. & Biotechnol.*, **16**: 1-7.
- Qasim, M. (2000). Physiological and biochemical studies in a potential oilseed crop canola (*Brassica napus* L.) Under Salinity (NaCl) stress. Ph.D thesis. Department of Botany, University of Agriculture, Faisalabad, Pakistan.
- Sgherri, C., Maffei, M. and Izzo, F. (2000). Antioxidative enzymes in wheat subjected to increasing water deficit and rewatering. *J. Plant Physiol.*, **157**: 273-279.
- Somogyi, N. (1952). *J. Biol. Chem.*, **195** : 1.
- Srivalli, B., Chinnusamy, V. and Khanna-Chopra, R. (2003). Antioxidative defense in response to abiotic stress in plant. *J.Plant Biol.*, **30**: 121-139.
- Vasantha, S. and Rajalakshmi, R. (2009). Progressive change in biochemical characters of sugarcane genotypes under salinity stress. *Indian J.Plant Physiol.*, **14** : 34-38.

