

RESEARCH PAPER

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# Evaluation of sanitization treatments for red amaranthus (*Amaranthus tricolor* L.)

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## SUMMARY :

Amaranthus is known as 'poor man's spinach' which is most commonly used leafy vegetable rich in nutrients. Increased need of sanitization in postharvest handling of leafy vegetables has been reported due to outbreak of disease causing pathogens and also due to huge postharvest loss. In the present study amaranthus (var. Arun) were sanitized with different sanitizing agents viz., ozonised water, sodium hypochlorite, brine and tap water (control). Effectiveness of these sanitizing agents for surface decontamination of harvested amaranthus was evaluated by analyzing microbial, physiological and visual parameters. The results revealed that amaranthus sanitized with 2 ppm ozonised water had highest microbial reduction (85.68%), lowest physiological loss in weight (25.63%), highest relative water content (63.11%) and highest score for visual parameters and overall acceptability which was followed by the treatment with 30 ppm sodium hypochlorite. Sanitization with 2 ppm ozonised water also extended the shelf-life of amaranthus upto 4 h of storage at room temperature.

**KEY WORDS** : Sanitization, Amaranthus, Ozonised water, Sodium hypochlorite, Shelf-life

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**A**maranth is one of the ancient leafy vegetables which could play an important role in rural economic and nutritional security because its cultivation allows good yields of high quality leaves to be integrated in daily poor diet. A loss of 5 to 10 per cent of fresh weight would make leafy vegetables to appear wilted and become unusable. In recent years there is an

increased need of sanitization in the chain of postharvest handling. Numerous cleaning compounds and processes have been developed to remove and destroy bacteria, viruses and parasites from vegetables. Washing with sanitizers is an important step in reducing the microbial population and quality deterioration (Kim, 2012). Use of sanitizers such as chlorine, peroxy acetic acid, hydrogen

peroxide, acidified sodium chloride or ozone can provide an additional reduction in the initial population of micro-organisms on the surface of fresh produce (Heard, 2000). Different protocols have been used for washing and disinfection of leafy vegetables and they comprise several washing times, kind of sanitizers and sanitizer concentrations and justifying its comparison (Oliveira *et al.*, 2012). Objectives of vegetable sanitization are reduction or elimination of micro-organisms from vegetables and guarantee it for consumption (Bachelli *et al.*, 2013). Hence the present study was conducted for evaluating the effectiveness of different sanitization treatments and shelf-life for red amaranthus (var. Arun).

## EXPERIMENTAL METHODS

The study was conducted at Department of Processing Technology, College of Agriculture, Vellayan, Kerala Agricultural University during the year 2013-2015. Freshly harvested Amaranthus (var. Arun) of uniform size without any visual defects were procured in the morning hours from VFPCCK (Vegetable and Fruit Promotion Council Keralam) progressive farmer's field, Vellayani cultivated as per Kerala Agricultural University package of practices. Harvested Amaranthus (stem with leaves) of 25-30 cm long (after removing root portion) were taken and fully immersed in following sanitizing agents for five minutes. For each treatment 200 g samples were taken with five replications.

- T<sub>1</sub> : Brine solution (2%)
- T<sub>2</sub> : Sodium hypochlorite (30 ppm)
- T<sub>3</sub> : Ozonised water (2 ppm)
- T<sub>4</sub> : Tap water

After the treatments excess water was drained and samples were air dried. Effectiveness of sanitizing agents for surface decontamination was evaluated microbiologically before and after the treatment. Sanitized amaranthus were stored at room temperature (30±2°C) and observed for visual and physiological parameters. The data were analyzed statistically by using ANOVA (Completely Randomized Design).

### Microbial count :

The quantitative assay of the micro flora in pre and post- treated samples was carried out by serial dilution spread plate technique. NA (nutrient agar) and RB (rose bengal agar) medium were used for the enumeration of bacterial and fungal population, respectively. Number of

micro-organisms (bacteria and fungi) per 10 g of pre and post-treated samples was calculated as per the following formula:

$$\text{No. of colony for ming uints} = \frac{\text{Total number of colony formed x dilution factor}}{\text{Aliquote plated}} \\ (\text{CFU per gram of the sample})$$

### Physiological parameters :

Physiological parameters *viz.*, physiological loss in weight and relative water content of sanitized amaranthus were recorded at hourly interval till the end of shelf-life. Amaranthus with cumulative physiological loss in weight of more than 25 per cent and relative water content less than 65 per cent with lower score for visual parameters and overall acceptability was considered as the end of shelf-life.

For determining physiological loss in weight, sample was weighed accurately at the time of storage (after treatment) and subsequently at an interval of one hour till the end of shelf-life. Cumulative physiological loss in weight was calculated using the formula :

$$\text{CPLW (\%)} = \frac{\text{Initial weight} - \text{Final weight}}{\text{Initial weight}} \times 100$$

### Relative water content (RWC) :

Percentage relative water content was estimated according to the method proposed by Smart and Bingham (1974). A composite sample of leaf discs is taken and the fresh weight is determined, followed by flotation in distilled water for up to 1 hour. The turgid weight is then recorded and the leaf tissue is subsequently oven dried to a constant weight at about 85°C. RWC is calculated in percentage by :

$$\text{RWC (\%)} = \frac{(\text{Fresh weight} - \text{Dry weight})}{(\text{Turgid weight} - \text{Dry weight})} \times 100$$

### Visual parameters :

Visual parameters like colour, texture, appearance, leaf wilting and defoliation/decay of sanitised amaranthus were taken initially and at an interval of one hour till the end of shelf-life by conducting a sensory evaluation performed by a 30 member semi-trained panel. A score card proposed by Swaminathan (1995) was used for assessing the sensory qualities with a 5 point hedonic scale with following scores.

Excellent- 5, Very good- 4, Good-3, Fair- 2, Poor - 1

The score given by 30 judges were statistically analyzed using the non-parametric ANOVA (Kruskall Wallis test) and mean ranks and critical values were calculated.

## EXPERIMENTAL FINDINGS AND ANALYSIS

The results obtained from the present investigation as well as relevant discussion have been summarized under following heads :

### Enumeration of total microbial count :

Before the sanitization treatments, bacterial population of amaranthus in each treatment was assessed and was in range of  $24.84$  to  $25.58 \times 10^5$  cfug<sup>-1</sup>. No significant difference was found between the treatments before the sanitization and no fungal population was found.

After the sanitization treatments, the treatment T<sub>3</sub> (2 ppm ozonised water) recorded the lowest bacterial population of  $3.24 \times 10^5$  cfug<sup>-1</sup> followed by T<sub>2</sub> (30 ppm sodium hypochlorite) with  $8.84 \times 10^5$  cfug<sup>-1</sup>. The highest bacterial population was recorded in T<sub>1</sub> (2 % brine) with  $15.51 \times 10^5$  cfug<sup>-1</sup> and it showed no significant difference with the treatment T<sub>4</sub> ( $14.17 \times 10^5$  cfug<sup>-1</sup>).

Surface sanitization of amaranthus (var. Arun) with 2 ppm ozonised water was found highly effective with 85.68 per cent reduction in microbial population followed by 30 ppm sodium hypochlorite with a reduction percentage of 64.26 (Table 1). Similar result was reported

by Olmez and Akbas (2009) that application of 2 ppm ozonated water treatment for two minutes was the optimum processing conditions for ozone disinfection of green leaf lettuce, in terms of reducing the microbial load during cold storage. The sanitizing efficiency of ozone may be due to the fact that ozone destroys micro-organism by disruption or disintegration of cell envelope and internal cellular proteins by progressive oxidation causing rapid cell death (Komanapalli and Lau, 1996 and Kim, 2012). Ibrahim *et al.* (2012) reported that in turnip greens microbial population was reduced to 1.43 log cfug<sup>-1</sup> when treated with 5 ppm ozone for five minutes. Chen *et al.* (2013) concluded that 3 ppm ozone is more effective in inactivating bacteria, moulds and yeasts on vegetables. Nath *et al.* (2014) observed that total bacterial count reduction was greater than 90 per cent in chinese cabbages treated with ozonated water (2.3 mg/l) for 60 minutes.

Surface sanitization with 30 ppm sodium hypochlorite reduced microbial population than brine and tap water. Similar result was reported by Varghese (2006) in fresh-cut vegetables like cowpea, okra, brinjal, ash gourd, pumpkin and elephant foot yam and Chandran (2013) in fresh-cut cabbage, beans, carrot and beetroot. Chlorine selectively destroys certain intracellular enzyme systems of micro-organisms causing rapid cell death that helped in reducing the microbial population (Karaca and Velioglu, 2007).

Surface sanitization of amaranthus with 2 per cent

**Table 1 : Effect of sanitizing treatments on the microbial population of amaranthus**

Treatments	Bacterial population $\times 10^5$ cfug <sup>-1</sup>			Fungal population
	Before treatment	After treatment	Reduction %	After treatment
T <sub>1</sub>	25.58	15.51 <sup>a</sup>	39.39 <sup>c</sup> (6.26 <sup>c</sup> )	TLTC
T <sub>2</sub>	24.84	8.84 <sup>b</sup>	64.26 <sup>b</sup> (8.02 <sup>b</sup> )	TLTC
T <sub>3</sub>	24.84	3.24 <sup>c</sup>	85.68 <sup>a</sup> (9.24 <sup>a</sup> )	TLTC
T <sub>4</sub>	24.84	14.17 <sup>a</sup>	42.91 <sup>c</sup> (6.54 <sup>c</sup> )	TLTC
C.D. (P=0.05)	NS	4.549	6.914 (0.444)	

Figure in parentheses is square root transformed values; TLTC- Too Less to count

NS= Non-significant

**Table 2 : Effect of sanitizing treatments on physiological loss in weight (%)**

Treatments	After storage		
	2 h	3 h	4 h
T <sub>1</sub>	12.55 <sup>a</sup>	28.95 <sup>a</sup>	31.04 <sup>a</sup>
T <sub>2</sub>	8.23 <sup>c</sup>	21.36 <sup>c</sup>	27.53 <sup>c</sup>
T <sub>3</sub>	6.88 <sup>d</sup>	11.17 <sup>d</sup>	25.63 <sup>d</sup>
T <sub>4</sub>	10.07 <sup>b</sup>	22.53 <sup>b</sup>	29.12 <sup>b</sup>
C.D. (P= 0.05)	1.285	0.253	0.021

brine had highest microbial population ( $15.51 \times 10^5$  cfug<sup>-1</sup>) with lowest reduction percentage of 39.39 per cent which was even lower than treatment with tap water (42.91%). This result is supported by the findings of Reddy (2010) in which brine (2%) dipping resulted in maximum decaying of rajagira leaves (*Amaranthus paniculatus*).

As salt acts as osmotic agent, draws water from the tissues leading to condition of moisture on inner surface of the film (Anonymous, 1999). Condensed moisture content inside the package adds to the acidity of the leafy vegetables which can end up as the cause of deterioration and growth of the microorganisms (King *et al.*, 1991; Ahvenainen, 1996 and Brackett, 1999).

**Physiological parameters :**

Physiological weight loss is a phenomenon of weight loss from produce during storage which in turn is related with shelf-life of vegetables whether whole or fresh-cut products. Amaranthus treated with 2 ppm ozonised water was found to have lowest physiological loss in weight of 6.88, 11.17 and 25.63 per cent at 2, 3 and 4 h of storage at room temperature (Table 2). This result was in agreement with the findings of Nadas *et al.* (2003) that fruits treated with ozonised water showed less weight loss than the non-treated samples after storage. Zhang *et al.* (2005) reported that fresh-cut celery treatment with ozonated water showed to be effective to reduce the population of micro-organisms and retard physiological

metabolism. Sothornvit and Kiatchanapaibul (2009) reported that ozone treatment for fresh or fresh-cut vegetables extended the shelf-life by inhibiting growth of micro-organisms.

Relative water content was also found to be highest in the 2 ppm ozonised water treated amaranthus during 2, 3, and 4 h of storage as 82.70, 75.92 and 63.11 per cent, respectively (Table 3). Similarly, Beltran *et al.* (2005) reported that fresh visual appearance of fresh-cut lettuce and fresh-cut potato was maintained by ozonated water during storage.

It could be noted that there is a relation between physiological loss in weight, relative water content and shelf-life or freshness of leafy vegetable. Increase in weight loss decreases the water content and in turn reduces the freshness or shelf-life of leafy vegetables.

Amaranthus treated with 2 per cent brine were unable to sustain edibility. Highest physiological loss in weight of 12.55, 28.95 and 31.04 per cent was recorded by 2 per cent brine after 2, 3 and 4 h of storage, respectively. Lowest relative water content of 74.34, 60.24 and 53.64 per cent was observed in 2 per cent brine treated amaranthus, respectively during 2, 3 and 4 h of storage. Amoah *et al.* (2007) found that higher concentration of brine treatment greatly reduced the quality of the lettuce leaves. Reddy (2010) reported that brine (2%) used as dipping solution reduced the moisture significantly and could not extend the shelf-life of *rajagira* leaves with or without stem.

**Table 3 : Effect of sanitizing treatments on relative water content (%)**

Treatments	Before treatment	After treatment	After storage		
			2 h	3 h	4 h
T <sub>1</sub>	83.12	82.03 <sup>b</sup>	74.34 <sup>d</sup>	60.24 <sup>d</sup>	53.64 <sup>d</sup>
T <sub>2</sub>	83.22	86.22 <sup>a</sup>	77.88 <sup>c</sup>	65.11 <sup>b</sup>	58.43 <sup>b</sup>
T <sub>3</sub>	82.69	87.41 <sup>a</sup>	82.70 <sup>a</sup>	75.92 <sup>a</sup>	63.11 <sup>a</sup>
T <sub>4</sub>	82.93	87.28 <sup>a</sup>	80.58 <sup>b</sup>	64.38 <sup>c</sup>	55.23 <sup>c</sup>
C.D. (P=0.05)	NS	2.930	1.831	0.219	0.028

NS= Non-significant

**Table 4 : Effect of sanitizing treatments on visual parameters after 2 h of storage**

Treatments	Colour		Texture		Leaf wilting		Defoliation		Overall acceptability	
	Mean rank	Mean score	Mean rank	Mean score	Mean rank	Mean score	Mean rank	Mean score	Mean rank	Mean score
T <sub>1</sub>	34.25	3.25	30.40	3.60	66.00	2.10	59.55	1.45	26.50	3.00
T <sub>2</sub>	52.40	3.55	43.65	4.00	31.35	1.15	40.50	1.00	38.30	3.35
T <sub>3</sub>	60.65	4.00	51.45	4.25	29.4	1.10	34.50	1.00	60.05	3.95
T <sub>4</sub>	39.75	3.35	36.50	3.80	35.25	1.25	52.50	1.15	37.15	3.30
C.V. (P= 0.05)						7.815				

**Visual parameters :**

Amaranthus is highly perishable in nature which leads to loss of freshness during storage. Amaranthus treated with 2 ppm ozonised water (T<sub>3</sub>) recorded the highest mean score for visual parameters viz., colour (4.00), texture (4.25) and overall acceptability (3.95) and lowest mean score for defoliation (1.00) and leaf wilting (1.1) (Table 4). It was followed by T<sub>2</sub> (30 ppm sodium hypochlorite) with a mean score for colour (3.55), texture (4.00), overall acceptability (3.35), defoliation (1.00) and leaf wilting (1.15). Minimum mean score was obtained for T<sub>1</sub> (2% brine) in colour (3.25), texture (3.6) and overall acceptability (3.00) and highest for leaf wilting (2.1) and defoliation (1.45) after 2 h of storage.

After 3 h of storage, amaranthus dipped in 2 ppm ozonised water (T<sub>3</sub>) recorded the highest mean score for colour (3.55), texture (3.3) and overall acceptability (3.5) and lowest mean score for leaf wilting (2.00) and defoliation (2.05) which was followed by 30 ppm sodium hypochlorite (T<sub>2</sub>) treated amaranthus with a mean score of 2.30, 2.80, 2.90, 2.25 and 2.55 for colour, texture, overall acceptability, leaf wilting and defoliation, respectively (Table 5). Zhang *et al.* (2005) revealed that the best preservation effect was found to be in treatment with ozonated water by which the microbial population was able to be lowered and nutritional and sensory quality of fresh-cut celery was maintained good for 9 days of storage at 4°C. Lowest mean score was observed for T<sub>1</sub>

(2% brine) for colour (1.8), texture (2.00) and overall acceptability (2.2) and highest for leaf wilting (3.6) and defoliation (3.05).

After 4 h of storage, amaranthus dipped in 2 ppm ozonised water (T<sub>3</sub>) showed highest mean score for colour (2.4), texture (2.75) and overall acceptability (2.00) and lowest for leaf wilting (2.3) and defoliation (2.65) which was followed by 30 ppm sodium hypochlorite (T<sub>2</sub>) (Table 6). Olmez and Akbas (2009) also reported similar results with application of 2 ppm ozonated water treatment for 2 minutes was the optimum processing conditions for ozone disinfection of green leaf lettuce, in terms of reducing the microbial load and maintaining the sensory quality during cold storage. Minimum mean score was obtained by amaranthus dipped in 2 per cent brine (T<sub>1</sub>) for colour (1.85), texture (1.55) and overall acceptability (1.6) and maximum for leaf wilting (3.7) and defoliation (3.45). This could be due to the dehydrating nature of salt which renders to loss of moisture during storage which results in loss of freshness and turgidity of produce (Anonymous, 1999).

Freshness of a produce is related to the shelf-life of a produce. Amaranthus treated with ozonised water (2 ppm) had the longest shelf-life upto 4 h when compared to other treatments. Beltran *et al.* (2005) reported that the use of ozonated water applied to fresh-cut vegetables for sanitation purposes reduced the microbial populations and extended the shelf-life of some of these products. Ozone treatment is effective in inhibiting the growth of micro-organisms

**Table 5 : Effect of sanitizing treatments on visual parameters after 3 h of storage**

Treatments	Colour		Texture		Leaf wilting		Defoliation		Overall acceptability	
	Mean rank	Mean score	Mean rank	Mean score	Mean rank	Mean score	Mean rank	Mean score	Mean rank	Mean score
T <sub>1</sub>	21.85	1.80	22.55	2.00	66.10	3.60	54.70	3.05	19.80	2.20
T <sub>2</sub>	35.60	2.30	47.55	2.80	28.37	2.25	36.67	2.55	43.60	2.90
T <sub>3</sub>	67.35	3.55	62.15	3.30	20.72	2.00	19.37	2.05	60.75	3.50
T <sub>4</sub>	37.20	2.35	29.75	2.25	46.80	2.80	51.25	2.90	37.85	2.75
C.V. (P=0.05)						7.815				

**Table 6 : Effect of sanitizing treatments on visual parameters after 4 h of storage**

Treatments	Colour		Texture		Leaf wilting		Defoliation		Overall acceptability	
	Mean rank	Mean score	Mean rank	Mean score	Mean rank	Mean score	Mean rank	Mean score	Mean rank	Mean score
T <sub>1</sub>	33.72	1.85	18.20	1.55	60.80	3.70	56.12	3.45	34.40	1.60
T <sub>2</sub>	40.42	2.05	48.25	2.55	28.90	2.60	36.47	2.85	41.82	1.85
T <sub>3</sub>	50.65	2.40	54.30	2.75	20.20	2.30	29.77	2.65	47.72	2.00
T <sub>4</sub>	37.20	1.95	41.25	2.35	52.10	3.40	39.62	2.95	38.05	1.7
C.V. (P=0.05)						7.815				

and extends the shelf-life of fresh or fresh-cut products by Sothornvit and Kiatchanapaibul (2009).

## Conclusion

Surface sanitization of red amaranthus (var.Arun) with different sanitizing agents had showed a significant reduction in microbial population. Amaranthus surface sanitized with 2 ppm ozone was found to be best the sanitizer with lowest microbial population and had highest quality parameters with prolonged shelf-life upto four hours during room temperature storage.

## LITERATURE CITED

- Ahvenainen, R. (1996).** New approaches in improving the shelf-life of minimally processed fruit and vegetables. *Trends Food Sci. & Technol.*, **7**: 179–187.
- Amoah, P., Drechsel, P., Abaidoo, R.C. and Klutse, A. (2007).** Effectiveness of common and improved sanitary washing methods in selected cities of West Africa for the reduction of coliform bacteria and helminth eggs on vegetables. *Trop. Medicine & Internat. Health*, **12** (2): 40–50.
- Anonymous (1999). Home scale processing and preservation of fruits and vegetables. CFTRI, Mysore, pp.1-13.
- Bachelli, M. L.B., Amaral, R.D.A. and Benedetti, B.C. (2013).** Alternative sanitization methods for minimally processed lettuce in comparison to sodium hypochlorite. *Braz. J. Microbiol.*, **44** (3): 673–678.
- Beltran, D., Selma, M. V., Marin, A. and Gil, M.I. (2005).** Ozonated water extends the shelf-life of fresh-cut lettuce. *J. Agric. & Food Chem.*, **53** (14): 5654–5663.
- Brackett, R.E. (1999).** Incidence contributing factors and control of bacterial pathogens in produce. *Postharvest Biol. Technol.*, **15**: 305-311.
- Chandran, T.T. (2013).** Protocol development for fresh cut vegetables. M.Sc. (Hort.) Thesis, Kerala Agriculture University, Thrissur, KERALA (INDIA).
- Chen, X., Binjun, L., Lingchuan, M. and Shuangxi, F. (2013).** Research progress in preservation of postharvest leafy vegetables. *Adv. Materials Res.*, **749**: 401- 407.
- Heard, G. (2000).** Microbial safety of ready-to-eat salads and minimally processed vegetables and fruits. *Food Sci. Technol. Today*, **14**: 15-21.
- Ibrahim, S.A., Mutamba, O.Z., Yang, H., Salameh, M.M., Gyawali, R. and Seo, C.W. (2012).** Use of ozone and chlorine dioxide to improve the microbiological quality of turnip greens. *Emir. J. Food Agric.*, **24** (3): 185-190.
- Karaca, H. and Velioglu, Y.S. (2007).** Ozone applications in fruit and vegetable processing. *Food Rev. Internat.*, **23**: 91–106.
- Kim, J.G. (2012).** Environmental friendly sanitation to improve quality and microbial safety of fresh-cut vegetables. In: *Biotechnology - Molecular Studies and Novel Applications for Improved Quality of Human Life*, Reda Sammour (Ed.), pp.173-196.
- King, J.A.D., Magnuson, J.A., Torok, T. and Goodman, N. (1991).** Microbial flora and storage quality of partially processed lettuce. *J. Food Sci.*, **56**: 459–461.
- Komanapalli, I.R. and Lau, B.H.S. (1996).** Ozone-induced damage of *Escherichia coli* K-12. *Appl. Microbiol. & Biotechnol.*, **46**: 610–614.
- Nadas, A., Olmo, M. and Garcia, J.M. (2003).** Growth of *Botrytis cinerea* and strawberry quality in ozone-enriched atmospheres. *J. Food Sci.*, **68** (5): 1798–1802.
- Nath, A., Mukhim, K., Swer, T., Dutta, D., Verma, N., Deka, B. C. and Gangwar, B. (2014).** A review on application of ozone in the food processing and packaging. *J. Food Product Develop. & Packag.*, **1**(2): 07-21.
- Oliveira, A.B.A., Ritter, A.C., Tondo, E.C. and Cardoso, M.I. (2012).** Comparison of different washing and disinfection protocols used by food services in Southern Brazil for Lettuce (*Lactuca sativa*). *Food & Nutr. Sci.*, **3**: 28- 33.
- Olmez, H. and Akbas, M.Y. (2009).** Optimization of ozone treatment of fresh-cut green leaf lettuce. *J. Food Engg.*, **90**: 487-494.
- Reddy, J.B. (2010).** Minimal processing of green leafy vegetables. M.Sc. (Home Sci.) Thesis, University of Agricultural Sciences, Dharwad KARNATAKA (INDIA).
- Smart, R.E. and Bingham, G.E. (1974).** Rapid estimates of relative water content. *Plant Physiol.*, **53**: 258-260.
- Sothornvit, R. and Kiatchanapaibul, P. (2009).** Quality and shelf-life of washed freshcut asparagus in modified atmosphere packaging. *Food Sci. Technol.*, **42**: 1484–1490.
- Swaminathan, M. (1995).** Food Science and Experimental Foods. Ganesh and Company, Madras, India. 293 p.
- Varghese, S. (2006).** Standardization of minimal processing techniques for selected vegetables. M.Sc. (Hort.) Thesis, Kerala Agriculture University, Thrissur (KERALA) INDIA.
- Zhang, L., Lu, Z., Yu, Z. and Gao, X. (2005).** Preservation of fresh-cut celery by treatment of ozonated water. *Food Control*, **16**: 279-283.

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