

RESEARCH ARTICLE

Studies on pre- and post emergence damping off on chilli caused by *Pythium ultimum*

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ABSTRACT

Chilli crop is attacked by more than dozen diseases of fungal, bacterial and viral nature leading to great loss to cultivators. Among those diseases, damping off of chilli incited by *Pythium* spp. is responsible for 90 per cent mortality either as pre or post-emergence damping off in nurseries and field conditions. Results indicated that the pre-emergence damping off was responded with significantly lowest seed rot found in PATH-34 (42.4 %) and this variety was at par with PATH-32 (60.3 %), PATH-24 (60.3%), PATH-9 (61.1%), PATH-7 (60.02%), PATH-6 (65.00 %) and PATH-26 (65.0%). Significantly highest seed mortality was noted in 58 entries where number of rotten seeds ranged from 69.6 to 96.7 per cent. Significantly higher germination in sick soil was PATH-34 (28.63 %), PATH-9 (16.41%), PATH-24 (15.61%), PATH-32 (15.61 %), PATH-6 (11.71 %), PATH-7 (11.71%) and PATH-26 (11.71%). While 58 entries have significantly less germination in sick soil. However, the post-emergence damping off the genotypes PATH-6 (22.86%), PATH-9 (22.86%) and PATH-34 (22.86%) were resistant (R) reaction with seedling mortality and the moderately resistant (MR) reaction of seedling mortality were PATH-24 (40.16%), PATH-32 (40.06%), PATH-30 (36.17%) and PATH-07 (31.46%). The 12 germplasm lines expressed moderately susceptible (MS) reaction having post-emergence mortality from 40.2 to 56.8 (%). The 18 germplasm lines expressed susceptible reaction (S) having 56.9 to 73.4 (%) post emergence mortality. The highly susceptible reaction (HS) was expressed by 28 entries having post-emergence mortality from 73.5 to 90 (%).

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INTRODUCTION

Chilli (*Capsicum annuum* L.) also known as red pepper is the member of family Solanaceae. Two species of chilli are under cultivation. The *Capsicum annuum* L. is small in size, more pungent types. Whereas, the *Capsicum frutescense* L. is somewhat larger, mild to moderately pungent types and referred as 'Dhobli Mirchi' and is used mostly as green vegetable. The pungency is due to the active principle capsaicin contained in the skin and septa of the fruit. Chillies are valued principally for their high pungency and for their colour. Chilli forms an indispensable culinary spice in several parts of the

world. It is also used in beverages and in the preparation of medicines. India is the largest producer, consumer and exporter of chillies in the world. The important states growing chilli are Andhra Pradesh, Maharashtra, Orissa, West Bengal, Karnataka, Rajasthan and Tamil Nadu. As per the latest statistics, India produced 1.24 million tonnes of dry chilli from an area of 0.077 million hectares (Anonymous, 2009).

Chilli crop is attacked by more than dozen diseases of fungal, bacterial and viral nature leading to great loss to cultivators. Among these diseases, damping off of chilli incited by *Pythium* spp. is responsible for 90 per cent mortality either as pre or post-emergence damping off in nurseries and fields

(Sowmini, 1961). *Pythium* damping off is very common problem in fields and greenhouse, where the organism kills newly emerged seedlings (Jarvis, 1992). Many *Pythium* species, along with their close relatives, *Phytophthora* spp. are plant pathogens of economic importance in agriculture. *Pythium* spp. tend to be very generalistic and unspecific in their host-range. They infect a large range of hosts (Owen-Going, 2002), while *Phytophthora* spp. are generally more host-specific. In chilli crop, two species of *Pythium* have been reported. *i.e. Pythium ultimum* and *Pythium aphanidermatum*. *P. ultimum* occurs relatively in higher frequency in dry areas.

MATERIALS AND METHODS

Pre-emergence damping off in sick soil :

This experiment was planned in split plot design with two main treatments *i.e.* I_0 = Uninoculated control and I_1 = Inoculation through sick soil and Sub treatments = 65 germplasm lines *viz.*, PATH 1 to PATH 65 with four replications. For preparation of inoculum in the form of sick soil, the normal surface soil was collected randomly from 15 spots, it was thoroughly mixed, sieved through screen and the sorghum flour was added @ 25 g/kg soil, so as to boost the organic carbon level of soil, for providing nutrition to pathogen. Then the soil was sterilized at 15-pound pressure per inch² for 1 hr consequently for two times an interval of 24 hrs. Then the plastic containers with drainage hole of 1 cm diameter at the bottom were filled with sterilized amended soil. Inoculum disc (5 mm) of the reisolate (RI₁) was slotted with cork borer and was transferred to sterile soil in plastic container. For incubation, these plastic containers were transferred to plastic trays containing sterile water at the bottom (1 cm height) and trays were covered with plastic sheet so to create the humidity for 4 days. After 4 days of incubation the seeds of each germplasm lines were sown @ 4 seeds/container. Four replications of the inoculated containers were maintained for each germplasm line. In the same way for uninoculated absolute control the sterilized soil was used. The observations on the pre-emergence damping off seed rot (%) and germination (%) were noted 14 days after sowing. The data on pre-emergence damping off and germination (%) were subjected to statistical analysis.

Post-emergence damping off in sick soil :

Post emergence damping off is also important component of the disease and this experiment was planned in split plot design with three main treatments *i.e.* T_0 = Absolute control, T_1 = Abiotic stress (stagnation + amendment of organic carbon) and T_2 : Inoculation through sick soil and Sub treatments = 65 germplasm lines *viz.*, PATH 1 to PATH 65 with four replications. The details of main treatments are as below:

T_0 : Absolute control :

Sterile soil was used and there was no amendment of

sorghum flour in the soil as well as there was no inoculation of pathogen. Therefore, this treatment was called as absolute control.

T_1 : Relative control :

It was maintained with stagnation by 1 cm of sterile water at the bottom of the trays and soil amendment with sorghum flour @ 25 g/kg soil and followed by sterilization of amended soil for 1 hour at 15 pound pressure psi consequently for two times at 24 hr interval.

T_2 : Inoculation through sick soil :

For filling the plastic containers of 125 ml of capacity with drainage hole of 1 cm diameter, the sterilized sorghum flour amended soil was used. Then these plastic containers were inoculated with *Pythium ultimum* (5 mm inoculum disc) and these were incubated in trays with sterile water of 1 cm height at the bottom. These were covered with plastic sheet. After 4 days of incubation, 12 days old seedlings of germplasm lines were transplanted in sick soil @ 4 seedlings/container/replication. In the same way, in T_0 and T_1 treatments seedlings were transplanted @ 4 seedlings / container / replication. The observations on seedling mortality were recorded after 3 to 5 days of transplanting. The data were subjected to statistical analysis.

RESULTS AND DISCUSSION

The results of the present study as well as relevant discussions have been presented under following sub heads:

Pre-emergence damping off in sick soil :

The resistant variety is the most cost effective and cheap input in vegetable production like chilli, so as to identify the source of resistance to pre-emergence damping off. For I_0 control; soil was sterilized and watered with sterile water. For the preparation of sick soil (I_1) the soil was added with 25 g of sorghum flour per kg of soil and then it was sterilized and transferred to plastic container of 130 ml capacity with drainage hole of 1 cm diameter. Each plastic container was inoculated with 5 mm inoculum disc of pathogen. Then these plastic containers were transferred to stagnant tray having 1 cm column of sterile water at the bottom. These trays were covered with polythene sheet so as to maintain maximum humidity. After inoculation the containers were incubated for 4 days. These containers were then sown with 4 seeds per container. The observations on seed rot and seed germination were recorded replication wise and were subjected to statistical analysis.

Results (Table 1 and 2) indicated that the seven entries responded significantly and lowest seed rot was found in PATH-34 (42.4 %) and this variety was at par with PATH-32 (60.3 %), PATH-24 (60.3%), PATH-9 (61.1%), PATH-7 (60.02%), PATH-6 (65.00 %). Significantly highest seed mortality was

Table 1: Screening of germplasm lines against pre- emergence damping off

GPL	Seed rot (%)			GPL	Seed rot (%)			GPL	Seed rot (%)			Main treat. (Inoculation)	
	I ₀	I ₁	Mean		I ₀	I ₁	Mean		I ₀	I ₁	Mean	S.E.±	C.D. 0.05
Path-1	46.29	79.85	63.07	Path-23	50.19	89.98	70.09	Path-45	34.07	89.98	62.03		
Path-2	59.59	69.72	64.66	Path-24	50.19	60.32	55.26	Path-46	26.76	89.98	58.37		
Path-3	31.46	89.98	60.72	Path-25	67.63	79.85	73.74	Path-47	59.59	89.98	74.79		
Path-4	36.16	79.85	58.01	Path-26	40.05	65.02	52.54	Path-48	40.05	89.98	65.02		
Path-5	42.39	79.85	61.12	Path-27	23.66	89.98	56.82	Path-49	44.75	89.98	67.37		
Path-6	46.29	65.02	55.66	Path-28	23.93	89.98	56.96	Path-50	11.71	89.98	50.85		
Path-7	37.69	65.02	51.36	Path-29	75.15	89.98	82.57	Path-51	26.76	89.98	58.37		
Path-8	40.86	89.98	65.42	Path-30	44.75	75.15	59.95	Path-52	37.69	89.98	63.84		
Path-9	54.89	61.12	58.01	Path-31	59.59	89.98	74.79	Path-53	22.86	89.98	56.42		
Path-10	49.45	89.98	69.72	Path-32	50.19	60.32	55.26	Path-54	15.34	89.98	52.66		
Path-11	31.46	89.98	60.72	Path-33	50.19	79.85	65.02	Path-55	50.99	89.98	70.49		
Path-12	54.89	79.85	67.37	Path-34	50.19	42.39	46.29	Path-56	23.93	89.98	56.96		
Path-13	52.52	89.98	71.25	Path-35	23.93	89.98	56.96	Path-57	79.85	89.98	84.92		
Path-14	54.89	89.98	72.44	Path-36	28.63	89.98	59.31	Path-58	69.72	89.98	79.85		
Path-15	35.35	89.98	62.67	Path-37	69.72	89.98	79.85	Path-59	60.32	89.98	75.15		
Path-16	22.86	79.85	51.36	Path-38	54.89	89.98	72.44	Path-60	69.72	89.98	79.85		
Path-17	37.69	89.98	63.84	Path-39	50.19	89.98	70.09	Path-61	40.05	89.98	65.02		
Path-18	41.59	89.98	65.79	Path-40	31.46	89.98	60.72	Path-62	31.46	89.98	60.72		
Path-19	56.42	89.94	73.18	Path-41	15.34	89.98	52.66	Path-63	28.63	89.98	59.31		
Path-20	59.59	75.15	67.37	Path-42	31.46	89.98	60.72	Path-64	27.56	89.98	58.77		
Path-21	41.59	89.98	65.79	Path-43	27.56	89.98	58.77	Path-65	27.56	89.98	58.77		
Path-22	23.66	79.85	51.76	Path-44	44.75	89.98	67.37	-----	-----	-----	-----		
Mean	42.32	84.72	63.52	Mean	42.32	84.72	63.52	Mean	42.32	84.72	63.52	1.58	7.14*
S.E. ±			6.9				6.9				6.9		
C.D.(0.05)			19.19*				19.19*				19.19*		
Interaction: I x GPL (Inoculation x Germ plasm lines)													
			S.E.±				9.75						
			C.D. (0.05)				27.13						

* GPL : Germplasm line, I₀ : Control, I₂ : Inoculated**Table 2: Categorization of germplasm lines on the basis of seed rot percentage in sick soil**

Category	Seed rot (%) (Arc sin)	Number of entries	Name of the entries
I	42.4-69.5	7	Path-34 (42.4), Path-09 (61.1), Path-24 (60.3), Path-32 (60.3), Path-06 (65.0), Path-07 (65.0), Path-26 (65.0)
II	69.6-96.7	58	Path-20 (75.2), Path-30 (75.1), Path-02 (69.7), Path-01 (80.0), Path-04 (80.0), Path-05 (79.9), Path-12 (79.9), Path-16 (80.0), Path-22 (79.6), Path-25 (79.9), Path-33 (79.9), Path-03 (90.0), Path-08 (90.0), Path-10 (90.0), Path-11 (90.0), Path-13 (90.0), Path-14 (90.0), Path-15 (90.0), Path-17 (90.0), Path-18 (90.0), Path-19 (89.9), Path-21 (90.0), Path-23 (90.0), Path-27 (90.0), Path-28 (90.0), Path-29 (90.0), Path-31 (90.0), Path-35 (90.0), Path-36 (90.0), Path-37 (90.0), Path-38 (90.0), Path-39 (90.0), Path-40 (90.0), Path-41 (90.0), Path-42 (90.0), Path-43 (90.0), Path-44 (90.0), Path-45 (90.0), Path-46 (90.0), Path-47 (90.0), Path-48 (90.0), Path-49 (90.0), Path-50 (90.0), Path-51 (90.0), Path-52 (90.0), Path-53 (90.0), Path-54 (90.0), Path-55 (90.0), Path-56 (90.0), Path-57 (90.0), Path-58 (90.0), Path-59 (90.0), Path-60 (90.0), Path-61 (90.0), Path-62 (90.0), Path-63 (90.0), Path-64 (90.0), Path-65 (90.0)
SE ±	9.75		For Interaction: I x V
C.D. 0.05	27.13		For Interaction: I x V

noted in 58 entries where number of rotten seeds ranged from 69.6 to 96.7 per cent. It can be concluded that (Table 3 and 4) the significantly higher germination in sick soil was found in PATH-34 (28.63 %), PATH-9 (16.41%), PATH-24 (15.61%), PATH-32 (15.61 %), PATH-6 (11.71 %), PATH-7 (11.71%) and PATH-26 (11.71%). While 58 entries had significantly less germination in sick soil. In present investigation, I x GPL interaction (Inoculation x Germplasm line) was also significant. Similar trend was also observed in respect of germination. Dahiphale (2006) and Chavan (2007) screened popular varieties of tomato. Rasal (2008) also screened popular varieties of brinjal against *P. ultimum* and found that Sel-5 variety was resistant. Zagade (2007) also screened popular varieties of chilli and noted that varieties DCL-1 and Maina

were resistant. The resistance to *Pythium arrhenomes* has also been reported by Subramainiam (1936) in sugarcane. Resistance has also been recorded in case of sorghum against *P. arrhenomos* (Wanger, 1936). The monogenic resistance against *Pythium* has been recorded in case of soybean and pea (Brown and Kennedy, 1965).

Post-emergence damping off :

Post emergence damping off is an important component of the damping off disease. In order to know the host resistance to post-emergence damping off, the main treatment of absolute control (T_0) was maintained by transplanting the seedlings in sterilized soil. The plastic container in absolute control (T_0) was watered with sterile water. In T_1 treatment, sterilized soil

Table 3: Screening of germ plasm lines against pre- emergence damping off

GPL	Germination (%)			GPL	Germination (%)			GPL	Germination (%)			Maint treat. (Inoculation)	
	I_0	I_1	Mean		I_0	I_1	Mean		I_0	I_1	Mean	S.E. \pm	C.D. 0.05
Path-1	23.94*	4.19	14.07	Path-23	19.24	0.57	9.91	Path-45	42.66*	0.57	21.62		
Path-2	11.44	7.82	9.63	Path-24	19.24	15.61	17.43	Path-46	35.35*	0.57	17.96		
Path-3	31.46*	0.57	16.02	Path-25	22.92	4.19	13.56	Path-47	11.44	0.57	6.01		
Path-4	27.56*	4.19	15.88	Path-26	22.86	11.71	17.29	Path-48	22.86*	0.57	11.72		
Path-5	28.63*	4.19	16.41	Path-27	40.86*	0.57	20.72	Path-49	18.96	0.57	9.77		
Path-6	23.93	11.71	17.82	Path-28	46.29*	0.57	23.43	Path-50	65.02*	0.57	32.80		
Path-7	32.53*	11.71	22.12	Path-29	8.09	0.57	4.33	Path-51	35.35	0.57	17.96		
Path-8	23.66*	0.57	12.12	Path-30	18.96	8.09	13.53	Path-52	32.53*	0.57	16.55		
Path-9	15.34	16.41	15.88	Path-31	11.44	0.57	6.01	Path-53	40.05*	0.57	20.31		
Path-10	15.06	0.57	7.82	Path-32	19.24	15.61	17.43	Path-54	54.89*	0.57	27.73		
Path-11	31.46*	0.57	16.02	Path-33	19.24	4.19	11.72	Path-55	20.04	0.57	10.31		
Path-12	15.34	4.19	9.77	Path-34	19.24	28.63*	23.94	Path-56	46.29*	0.57	23.43		
Path-13	25.01*	0.57	12.79	Path-35	46.29*	0.57	23.43	Path-57	4.19	0.57	2.38		
Path-14	15.34	0.57	7.96	Path-36	42.39*	0.57	21.48	Path-58	7.82	0.57	4.20		
Path-15	26.76*	0.57	13.67	Path-37	7.82	0.57	4.20	Path-59	15.61	0.57	8.09		
Path-16	40.05*	4.19	22.12	Path-38	15.34	0.57	7.96	Path-60	7.82	0.57	4.20		
Path-17	32.53*	0.57	16.55	Path-39	19.24	0.57	9.91	Path-61	22.86*	0.57	11.72		
Path-18	27.83*	0.57	14.20	Path-40	31.46*	0.57	16.02	Path-62	31.46*	0.57	16.02		
Path-19	20.31*	0.57	10.44	Path-41	54.89*	0.57	27.73	Path-63	42.39*	0.57	21.48		
Path-20	11.44	8.09	9.77	Path-42	31.46*	0.57	16.02	Path-64	36.16*	0.57	18.37		
Path-21	27.83*	0.57	14.20	Path-43	36.16*	0.57	18.37	Path-65	36.16*	0.57	18.37		
Path-22	40.86*	4.19	22.53	Path-44	18.96	0.57	9.77	-----	-----	-----	-----		
Mean	26.92	3.011	14.97	Mean	26.92	3.011	14.97	Mean	26.92	3.011	14.97	1.48	6.69*
SE \pm			5.0				5.0				5.0		
C.D.(0.05)			13.9				13.9				13.9		
Interaction: I x GPL (Inoculation x Germ plasm lines)													
S.E. \pm 7.08													
C.D. (0.05) 19.7*													

* GPL: Germplasm line, I_0 : Control, I_1 : Inoculated

was used and favourable abiotic factors were provided, which included stagnation with sterile water and amendment of sorghum flour to raise organic carbon level. The sorghum flour was added @ of 25 g/kg of soil. After amendment of sorghum flour, the soil was sterilized at 15 lb pressure per inch² for 1 hr. So as to have perfect penetration of heat in soil during sterilization, the soil was sterilized in cottony or gunny bag. After sterilization, the soil was transferred to plastic container for T₁ treatment. In case of T₀ treatment the soil was sterilized at 15 lb pressure inch² for 1 hour but it was without sorghum flour. In T₂ treatment, all favourable abiotic factors added. All the plastic containers having drainage hole of 1 cm diameter at bottom were transferred to trays having 1 cm column of sterile water. Then these plastic containers for T₂ main treatment were inoculated with 5 mm inoculum disc of *Pythium ultimum* Trow. After inoculation, the trays were covered with polythene sheet so as to provide maximum humidity for the development of the inoculum in soil. After 5

days of incubation under humid condition, 8 days old seedlings of each germplasm line grown on sterile soil were transplanted to plastic containers. Observation on seedlings mortality was noted on 4th day of transplanting. Observation of post-emergence damping off expressed under T₀, T₁ and T₂ treatments in different germplasm line were recorded.

Results indicated that (Table 5 and 6) the mortality ranged from 8 per cent to 32.8 per cent. Significantly lowest mortality was expressed in PATH-6 (8 %) and significantly highest mortality was noted in PATH-48 (32 %). The interaction inoculation x Germplasm line (I x GPL) was also significant. The interaction inoculation x Germplasm line (I x GPL) revealed that three germplasm lines responded with significantly highest resistance to post-emergence mortality. These were PATH-6, PATH-9 and PATH-34 with only 22.9 per cent post emergence mortality. In screening, 4 entries responded with moderately resistant (MR) reaction with seedling mortality from 23.6 to 40.1 (%). The 12 germplasm lines expressed

Table 4 : Classification of germplasm lines on the basis of germination (%) in sick soil

Category	(%) Germination (Arc sin)	Resistance category	Number of entries	Name of the entry with germination (%) in bracket
I	28.63-8.93	R	7	Path-34 (28.63), Path-09 (16.41), Path-24 (15.61), Path-32 (15.61), Path-06 (11.71), Path-07 (11.71), Path-26 (11.71)
II	< 8.94	S	58	Path-20 (8.09), Path-30 (8.09), Path-02 (7.82), Path-01 (4.19), Path-04 (4.19), Path-05 (4.19), Path-12 (4.19), Path-16 (4.19), Path-22 (4.19), Path-25 (4.19), Path-33 (4.19), Path-03 (0.57), Path-08 (0.57), Path-10 (0.57), Path-11 (0.57), Path-13 (0.57), Path-14 (0.57), Path-15 (0.57), Path-17 (0.57), Path-18 (0.57), Path-19 (0.57), Path-21 (0.57), Path-23 (0.57), Path-27 (0.57), Path-28 (0.57), Path-29 (0.57), Path-31 (0.57), Path-35 (0.57), Path-36 (0.57), Path-37 (0.57), Path-38 (0.57), Path-39 (0.57), Path-40 (0.57), Path-41 (0.57), Path-42 (0.57), Path-43 (0.57), Path-44 (0.57), Path-45 (0.57), Path-46 (0.57), Path-47 (0.57), Path-48 (0.57), Path-49 (0.57), Path-50 (0.57), Path-51 (0.57), Path-52 (0.57), Path-53 (0.57), Path-54 (0.57), Path-55 (0.57), Path-56 (0.57), Path-57 (0.57), Path-58 (0.57), Path-59 (0.57), Path-60 (0.57), Path-61 (0.57), Path-62 (0.57), Path-63 (0.57), Path-64 (0.57), Path-65 (0.57)
S.E. ±	7.08			For Interaction: I x GPL
C.D. 0.05	19.68			For Interaction: I x GPL

Statistical significance

Germination (%)							
0.57	4.19	7.82	8.09	11.71	15.61	16.41	28.63
S				R			

Table 15: Seedling of *Ipomoea pes-caprae* (L.) DC. and *Ipomoea pes-caprae* (L.) DC. under different treatments.

C.T.	Seedling mortality %			C.T.			Seedling mortality %			C.T.			Seedling mortality %			Variance				
	T ₁	T ₂	T ₃	T ₁	T ₂	T ₃	T ₁	T ₂	T ₃	T ₁	T ₂	T ₃	T ₁	T ₂	T ₃	T ₁	T ₂	T ₃		
Rep 1	0.6	0.6	50.2	1.2	0.6	79.9	28.2	Rep 4	0.6	0.6	5.9	18.7	0.6	0.6	5.9	18.7	0.6	0.6	5.9	18.7
Rep 2	0.6	0.6	79.5	0.6	0.6	70.1	13.7	Rep 5	0.6	0.6	90.0	30.7	0.6	0.6	90.0	30.7	0.6	0.6	90.0	30.7
Rep 3	0.6	1.2	60.3	0.6	1.2	79.9	28.2	Rep 6	0.6	0.6	69.7	23.6	0.6	0.6	69.7	23.6	0.6	0.6	69.7	23.6
Rep 4	0.6	0.6	59.6	1.2	0.6	7.8	3.2	Rep 7	0.6	0.6	50.0	32.8	0.6	0.6	50.0	32.8	0.6	0.6	50.0	32.8
Rep 5	0.6	0.6	50.0	0.6	1.2	90.0	31.6	Rep 8	0.6	0.6	59.6	21.5	0.6	0.6	59.6	21.5	0.6	0.6	59.6	21.5
Rep 6	0.6	0.6	22.9	0.6	0.6	69.7	23.6	Rep 9	0.6	0.6	79.5	16.9	0.6	0.6	79.5	16.9	0.6	0.6	79.5	16.9
Rep 7	1.2	0.6	31.5	1.2	0.6	90.0	31.6	Rep 10	0.6	0.6	65.0	23.3	0.6	0.6	65.0	23.3	0.6	0.6	65.0	23.3
Rep 8	0.6	0.6	50.2	0.6	1.2	36.2	13.7	Rep 11	0.6	0.6	69.7	21.8	0.6	0.6	69.7	21.8	0.6	0.6	69.7	21.8
Rep 9	1.2	1.2	22.9	0.6	0.6	79.9	21.0	Rep 12	0.6	0.6	71.3	21.1	0.6	0.6	71.3	21.1	0.6	0.6	71.3	21.1
Rep 10	1.2	0.6	90.0	0.6	0.6	70.1	13.7	Rep 13	0.6	0.6	65.0	22.1	0.6	0.6	65.0	22.1	0.6	0.6	65.0	22.1
Rep 11	0.6	0.6	50.0	0.6	0.6	90.0	30.7	Rep 14	0.6	0.6	69.7	21.8	0.6	0.6	69.7	21.8	0.6	0.6	69.7	21.8
Rep 12	0.6	0.6	59.6	0.6	1.2	22.9	9.2	Rep 15	0.6	0.6	65.0	23.3	0.6	0.6	65.0	23.3	0.6	0.6	65.0	23.3
Rep 13	1.2	0.6	69.7	0.6	0.6	79.9	21.0	Rep 16	0.6	0.6	79.9	28.2	0.6	0.6	79.9	28.2	0.6	0.6	79.9	28.2
Rep 14	0.6	0.6	75.2	0.6	0.6	90.0	30.7	Rep 17	0.6	0.6	90.0	31.6	0.6	0.6	90.0	31.6	0.6	0.6	90.0	31.6
Rep 15	0.6	0.6	57.9	0.6	1.2	69.7	21.8	Rep 18	0.6	0.6	65.0	22.1	0.6	0.6	65.0	22.1	0.6	0.6	65.0	22.1
Rep 16	1.2	1.2	50.0	0.6	0.6	59.6	21.5	Rep 19	0.6	0.6	59.6	20.3	0.6	0.6	59.6	20.3	0.6	0.6	59.6	20.3
Rep 17	1.2	0.6	57.9	0.6	0.6	90.0	30.7	Rep 20	0.6	0.6	61	21.6	0.6	0.6	61	21.6	0.6	0.6	61	21.6
Rep 18	0.6	0.6	75.2	0.6	0.6	50.0	30.7	Rep 21	0.6	0.6	69.7	23.6	0.6	0.6	69.7	23.6	0.6	0.6	69.7	23.6
Rep 19	1.8	0.6	32.8	0.6	1.2	79.9	28.2	Rep 22	0.6	0.6	69.7	23.6	0.6	0.6	69.7	23.6	0.6	0.6	69.7	23.6
Rep 20	0.6	0.6	50.2	0.6	1.2	79.9	29.7	Mean	1.6	1.5	66.7	23.2	1.6	1.5	66.7	23.2	1.6	1.5	66.7	23.2
Rep 21	0.6	1.2	50.2	0.6	0.6	79.9	21.0	S.D.	3.75	3.75	5.98*	5.98*	3.75	3.75	5.98*	5.98*	3.75	3.75	5.98*	5.98*
Rep 22	1.2	0.6	60.3	0.6	0.6	76.3	15.8	C.D. (0.05)	9.98*	9.98*	16.60	16.60	9.98*	9.98*	16.60	16.60	9.98*	9.98*	16.60	16.60

Category	Seedling mortality (%)	No. of entry	Category	Seedling mortality (%)	No. of entry
1	19.23.5	3	R	32.85.6	3
2	23.6.10.	1	MS	32.85.6	1
3	10.2.56.8	12	MS	32.85.6	12
4	56.9.19.1	18	S	32.85.6	18
5	19.5.50.0	28	MS	32.85.6	28

moderately susceptible (MS) reaction having post-emergence mortality from 40.2 to 56.8 (%). The 18 germplasm lines expressed susceptible reaction (S) having 56.9 to 73.4 (%) post emergence mortality. The highly susceptible reaction (HS) was expressed by 28 entries having post-emergence mortality from 73.5 to 90 (%). Screening of soybean varieties against *Pythium ultimum* was undertaken by Zhang *et al.* (1998) and Brown and Kennedy (1965). Dahiphale (2006) and Chavan (2007) also noted Parbhani Yashashri a popular variety of tomato which has shown resistance to *P. ultimum*. Rasal (2008) has also observed Sel-5 variety of brinjal resistant to *P. ultimum*.

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