

RESEARCH PAPER

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In vitro evaluation of mutant and wild strain of Trichoderma harzianum against soil borne plant pathogen

- A.A. WALUNJ*¹, P.B. ABHANG² AND PRIYA JOHN²
 - Department of Plant Pathology, College of Agriculture, Loni, AHMEDNAGAR (M.S.) INDIA
 - ²Department of Plant Pathology, N. M. College of Agriculture, Navsari Agricultural University, NAVSARI (GUJARAT) INDIA

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*Corresponding author:

Email: akshaya17289@gmail.com

ABSTRACT

Aqueous suspension of conidia of *Trichoderma harzianum* wild strain Th-W were placed on potato dextrose agar and expose to UV irradiation for 10, 20, 30, and 40 min at 20 cm distance from which four stable mutants of *T. harzianum i.e.*, Th-M-1, Th-M-2, Th-M-3 and Th-M-4 were obtained as it differed considerably from wild strain (Th-W) for their morphological characteristics. *In vitro* evaluation of mutant and wild strain of *T. harzianum* against three soil borne plant pathogens, *Fusarium oxysporum* f.sp. *lycopersici, Sclerotium rolfsii* and *Macrophomina phaseolina by* dual culture method, revealed that mutant strains overgrew all the pathogenic fungi more rapidaly than the wild strain.

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INTRODUCTION

Trichoderma harzianum has been promising biocontrol agent of many plant diseases caused by the soil-borne pathogens like fungi such as Macrophomina phaseolina, Rhizoctonia solani, Fusarium species and Alternaria species on various crops. In addition, species of Trichoderma also have growth promoting capabilities that may or may not be integral to biological control (Dubey et al., 2007). The of *Trichoderma* spp. mechanisms antagonize phytopathogenic fungi include competition, colonization, antibiosis and direct mycoparasitism (Howell, 2003). This antagonistic potential serves as the basis for effective biological control application of *Trichoderma* spp. as an alternative method to chemicals for the control of a wide spectrum of plant pathogens (Chet, 1987).

MATERIAL AND METHODS

Development of mutant Trichoderma harzianum:

The mutant of *Trichoderma harzianum* was developed through UV irradiation. Conidial suspension of *Trichoderma harzianum* (Th-W) was prepared by dislodging the conidia from the ager surface by pouring sterilized water. The conidial concentration was adjusted with the help of haemocytometer to 106 ml. One ml of conidial suspension was placed on PDA plates and exposed to 30 W UV-irradiation for 10, 20, 30, and 40 min at 20 cm distance from which four stable mutants of *Trichoderma harzianum i.e.*, Th-M-1, Th-M-2, Th-M-3 and Th-M-4 were obtained. All the isolates were grown on PDA in Petri dishes. Plates were inoculated at the centre with a 5 mm mycelial disc taken from stock slants. The plates inoculated were incubated at room temperature (27 ± 2°C). Each stable isolates after four repetitions were examined properly and

regularly. Colony diameter was measured after incubation period 12, 24 and 48 (hr). Baby (1998) developed induced mutation on *T. longibrachiatum* by using ultra-violet irradiation and obtained mutant strains tolerant to bavistin. Intana *et al.* (2003) developed mutant of *T. harzianum* by using ultra-violet irradiation. Hunjan *et al.* (2004) developed two mutant strains TV34-M4 and TV34-M5 from *T. viride* by using ultra-violet irradiation. Manav and Singh (2003) developed mutant *T. harzianum* by exposing to ultra-violet irradiation for 90 min.

Antagonistic capability of mutant and wild *T. harzianum* against soil borne plant pathogen:

Sterilized PDA *i.e.* 20 ml was poured aseptically in sterilized Petriplates of 90 mm diameter. Mycelial discs (5mm) of 5 days old actively growing culture of the bioagent and the test pathogens were cut separately with the help of sterilized cork borer and placed on solidified PDA approximately 4 cm away from each other. The experiment was repeated fourth times along with their controls, where test pathogen subjected alone for growth comparison. All the inoculated plates were incubated at room temperature (27±2°C). The radial growth of test pathogens in treated and control plates were recorded after 5 days of incubation and the per cent inhibition of mycelial growth of the pathogens was calculated by using formula of

Bliss (1934). i.e.:

$$I = \frac{(C - T)}{C} \times 100$$

where, I = Inhibition per cent, C = Colony diameter in control plate, T = Colony diameter in treated plate.

RESULTS AND DISCUSSION

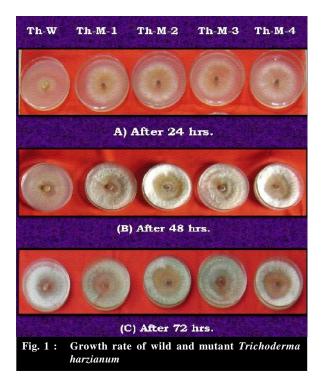
Radial growth rate of the wild and stable mutant of T. harzianum on PDA medium (Table 1) and depicted graphically in Fig. 1 showed that mutants of T. harzianum after 48 hr incubation showed higher radial growth as compared to 24 hr of incubation. Among the wild (Th-W) and mutant T. harzianum (Th-M-1, Th-M-2, Th-M-3, Th-M-4) mutant Th-M-2 recorded highly significant growth rate of 90 mm after 72 hr. These were followed by mutant isolates Th-M-3 (88.40 mm) and Th-M-4 (87.90 mm). However, after 24 hr, 48 hr and 72 hr mutant Th-M-2 showed increasing radial growth over other mutants and wild. Viswanth (2012) who recorded mutant T. harzianum (TM₁₇) showed highest growth rate and sporulation as compared to wild type. Henis et al. (1983) showed higher mycoparasitic ability of mutant against S. rolfsii. Mukherjee et al. (1997) showed higher mycoparasitic ability of mutant against S. rolfsii. Singh et al. (2002) found that T. harzianum inhibited the growth of F. udum.

Sr. No.	wth rate (mm) of wild and mutant Trichoderma harzianum Isolates of T. harzianum		*Average colony diameter (mm)			
		24 hr	48 hr	72 hr		
1.	Th-W	22.00	54.00	82.00		
2.	Th-M1	36.00	66.00	87.90		
3.	Th-M2	40.00	70.00	90.00		
4.	Th-M3	38.50	68.50	88.40		
5.	Th-M4	38.00	68.00	88.00		
S.E. ±		0.37				
C.D. $(P = 0.05)$		1.10				
C.V. (%)		2.57				

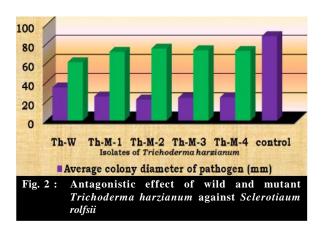
^{*}Average of four repetitions; Th-W: wild T. harzianum, Th-M-1, Th-M-2, Th-M-3, Th-M-4: Mutant T. harzianum

Sr. No.	Trichoderma harzianum isolates	% inhibition of mycelium growth (PIMG)			
		Fusarium oxysporum	Sclerotium rolfsii	Macrophomina phaseolina	
1.	Th-W	65.11	65.11	62.79	
2.	Th-M-1	73.25	73.25	70.93	
3.	Th-M-2	75.53	76.58	75.46	
4.	Th-M-3	73.83	74.34	72.09	
5.	Th-M-4	73.48	73.48	71.39	
5.	Control	-	-	-	
S.E.±		0.42	0.42	0.38	
C.D. (P=0	0.05)	1.25	1.27	1.15	
C.V. (%)		2.47	2.37	2.18	

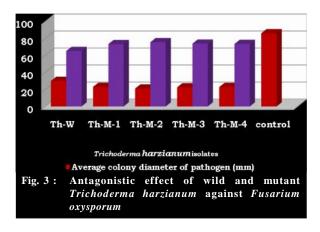
Th-W: wild T. harzianum, Th-M-1, Th-M-2, Th-M-3, Th-M-4: Mutant T. harzianum



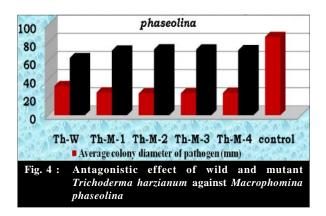
The antagonists tested against S. rolfsii were more or less inhibited the growth of the pathogen. Among the wild (Th-W) and mutant of T. harzianum (Th-M-1, Th-M-2, Th-M-3, Th-M-4) highest per cent mycelial inhibition was observed by mutant Th-M-2 (75.00 %) with 22.00 mm average colony diameter after five days of incubation which was followed by mutant Th-M-3 (73.29 %) and Th-M-4 (72.61 %) with average colony diameters 23.50 mm and 24.10 mm, respectively. The mutants had better per cent inhibition as compared to wild T. harzianum Th-W (60.79 %) with average colony diameter of 34.50 mm. which was showed in Table 2 and Fig. 2. Mech (2006) reported antagonistic effect of wild and mutant T. harzianum against the S. rolfsii showing maximum reduction of radial growth of pathogen by Th-M₂ as compared to Th-W, Th-M₁ and Th-M₂.



The antagonists tested against F. oxysporum were more or less inhibited the growth of the pathogen. Between wild T. harzianum (Th-W) and mutant T. harzianum (Th-M-1, Th-M-2, Th-M-3, Th-M-4), significantly highest mycelial inhibition was observed by isolate Th-M-2 (75.53 %) with 21.04 mm average colony diameter. which was followed by isolates Th-M-3 (73.83 %) and Th-M-4, (73.48 %) with average colony diameters 22.50 mm and 22.80 mm, respectively. While lowest per cent growth inhibition was found with isolates Th-W (65.11 %) with 30.00 mm average colony diameter of pathogen. which was showed in Table 2 and Fig. 3. Manay and Singh (2006) who tested mutant and parent strain of T. harzianum against three soil borne plant pathogens; F. oxysporum f. sp. ciceri, R. solani and M. phoseolina by dual culture method, revealed that mutant strain overgrew all the pathogenic fungi more rapidly than the parent strain. Balasubramanian et al. (2010) observed that the wild and mutant strains of T. harzianum showed differences in their spore colour, growth rate and pigmentation. The UV- mutant showed a puffy pale green pigmentation. Nagamani and Viswanth (2012) observed that the mutant T. harzianum (TM₁₇) showed highest growth rate and sporulation as compared to wild type. Johnson and Curl (1972) screened the selected mutant and parent strains of T. harzianum against three test pathogens, Fusarium oxysporum f. sp. ciceri, Macrophomina phaseolina and Rhizoctonia solani by dual culture method, revealed that mutant strain overgrew all the pathogenic fungi more rapidly than the parent strain.



Minimum mycelial growth and highest per cent growth inhibition were recorded in the case of T. harzianum isolate Th-M-2 (75.46 %) with 21.10 mm average colony diameter of the Macrophomina phaseolina. This was followed by isolates, Th-M-3 (72.09 %) with 24.00 mm average colony diameter of the pathogen and Th-M-4 (71.39 %) with 24.60 mm average colony diameter of pathogen. Lowest per cent growth inhibition was recorded with isolate Th-W (62.79 %) with 32.00 mm colony diameter which was showed in Table 2 and Fig. 4.



Patil and Kamble (2011) who reported that mutants had highest per cent reduction of mycelial growth. Similarly Ranganathswamy *et al.*, 2012; Chavan *et al.*, 2012; Lokesh *et al.*, 2012 and Sharma *et al.*, 2013 also worked in the related topic and the results coincides with the present findings.

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REFERENCES

Baby, U.I. (1998). Biocontrol potential of fungicide resistant mutants of *Trichoderma* spp. *Indian J. Microbiol.*, 38: 165-166.

Balasubramanian, N., Priya, V. Thamil, Gomathinayagam, S., Shanmugaiah, V., Jashnie, J. and Lalithakumari, D. (2010). Effect of chitin adapted and ultra violet induced mutant of *Trichoderma harzianum* enhancing biocontrol and chitinase activity. *Australian J. Basic & Appl. Sci.*, 4(10): 4701-4709.

Bliss, C.I. (1934). The method of probits. Sci., 79:38.

Chavan, A.A., Deshpande, G.D. and Zagade, S.N. (2012). Evaluation of *Trichoderma* species against *Pythium ultimum* pathogenic to tomato. *Internat. J. Pl. Protec.*, **5**(1): 147-149.

Chet I. (1987). Innovative Approaches to Plant Disease Control. New York: Wiley and Sons;. *Trichoderma* - Application, mode of action, and potential as a biocontrol agent of soilborne pathogenic fungi. 137–160pp.

Dubey, S.C., Suresh, M. and Birendra, S. (2007). Singh Evaluation of *Trichoderma* species against *Fusarium oxysporum* f.sp. *ciceris* for integrated management of chickpea wilts. *Biological Control*, **40**: 118–127.

Henis, Y., Adams, P.B., Lewis, J.A. and Papavizas, G.C. (1983). Penetration of sclerotia of *Sclerotium rolfsii* by *Trichoderma* spp. *Phytopathology*, **73**: 1043–1046.

Howell, C.R. (2003). Mechanisms employed by *Trichoderma* species in the biological control of plant diseases: the history and evolution of current concepts. *Plant Diseases*, 87: 4–10.

Hunjan, M.S., Astha, Singh Rama S. and Singh, Narinder (2004). Comparison of *Trichoderma viride* mutant and parent strains for their colony characters, tolerance to Bavistin and biocontrol efficacy against black scurf of potato. *J. Res.*, **41**(2): 231-238.

Intana, W., Chamswarng, C., Intanoo, W., Hongprayoon, C. and Sivasithamparam, K. (2003). Use of mutant strains for improved efficacy of *trichoderma harzianum* for controlling cucumber damping-off. *Thai J. Agric. Sci.*, 37: 429-439.

Johnson, Leander F., and Curl, Elroy A. (1972). Methods for Research on the Ecology of Soil-Borne Plant Pathogens. 426 So. Sixth St., Minneapolis, MN 55415: Burgess Publishing Company.

Lokesh, M.S., Patil, S.V., Gurumurthy, S.B., Palakshappa, M.G. and Anandaraj, M. (2012). Evaluation of combination of potassium phosphonate and *Trichoderma harzianum* on management of *Phytophthora* foot rot of black pepper (*Piper nigrum* L.) under arecanut cropping system. *Internat. J. Pl. Protec.*, 5(2): 356-360.

Manav, M. and Singh, R.S. (2003). Shelf life of different formulations of mutant and parent strain of *Trichoderma harzianum* at variable temperatures. *Plant Dis. Res.*, **18**(2):144-146.

Mech, S. (2006). UV-radiated mutant of *Trichoderma harzianum* Rifai and its carrier based formulation for management of white mold of French bean. M.Sc.(Ag.) Thesis, Assam Agricultural University, Jorhat, ASSAM (INDIA).

Mukherjee, P.K., Mukhopadhyay, A.N., Sarmah, D.K. and Shreshtha, S.M. (1995). Comparative antagonistic properties of *Gliocladium virens* and *Trichoderma harzianum* on *Sclerotium rolfsii* and *Rhizoctonia solani*—its relevance to understanding the mechanisms of biocontrol. *J. Phytopathol.*, 143 (5): 275-279.

Nagamani, P. and Viswanath, K. (2012). Variability studies of mutant and wild Trichoderma isolates against *Macrophomina phaseolina*. *J. Plant Dis. Sci.*, 7(1): 18-21.

Patil, V.B. and Kamble, S.S. (2011). The influence of ultraviolet light on antagonistic activity of *Trichoderma koningii* against *Macrophomina phaseolina* causing charcoal rot of sweet potato. *Internat. J. Acad. Res.*, **3** (1): 702–704.

Ranganathswamy, M., Patibanda, A.K., Chandrashekhar, G.S., Sandeep, D., Mallesh, S.B. and Halesh Kumar, H.B. (2012). Compatibility of *Trichoderma* isolates with selected fungicides *in vitro*. *Internat*. *J. Pl. Protec.*, **5**(1): 12-15.

Sharma, Prashant Kumar, Gothalwal, R. and Tiwari, R.K.S. (2013). Isolation of cold tolerant antifungal strains of *Trichoderma* sp. from Northern Hilly Zones of Chhattisgarh. *Internat. J. Plant Protec.*, **6**(2): 236-240.

Singh, S.K., Singh, R.H. and Dutta, S. (2002). Integrated management of pigeonpea wilt by biotic agents and biopesticides. *Ann. Plant Prot. Sci.*, **10** (2): 323–326.

