INTERNATIONAL JOURNAL OF PLANT PROTECTION VOLUME 8 | ISSUE 2 | OCTOBER, 2015 | 241-244



RESEARCH PAPER

DOI: 10.15740/HAS/IJPP/8.2/241-244

In vitro efficacy of different botanicals, bioagents, chemicals against *Xanthomonas axonopodis* pv.*citri* by turbidiometrical method

P.B. ABHANG*, M.V. TOTAWAR AND P.R. BRAHMANE

Department of Plant Pathology, Dr. Panjabrao Deshmukh Krishi Vidyapeeth, AKOLA (M.S.) INDIA

ARITCLE INFO

Received	:	20.02.2015
Revised	:	24.07.2015
Accepted	:	09.08.2015

KEY WORDS:

Xac, Citrus canker, Botanicals, bioagents, Turbidiometrical method, Spectrophotometer

ABSTRACT

Citrus canker disease of acid lime caused by *Xanthomonas axonopodis* pv. *citri* is an important disease in many parts of MH region .The bacterium infects the twigs, petioles, fruit stalks and fruits. Action of botanicals,bioagents,chemicals against *Xac* was studied by turbidiometrically at 24hrs.intervals upto 96hrs by spectrophotometer at 620nm.At 96 hours of incubation least bacterial growth(0.232OD) was exhibited in copper-oxychloride + streptomycin sulphate (0.2 % + 200ppm) followed with copper-oxychloride + streptomycin sulphate (0.2 % + 200ppm) followed with copper-oxychloride + streptomycin sulphate (0.2 % + 200ppm) followed with copper-oxychloride + streptomycin sulphate (0.2 % + 200ppm) followed with copper-oxychloride + streptomycin sulphate (0.2 % + 200ppm) followed with copper-oxychloride + streptomycin sulphate (0.2 % + 100ppm) (0.303, 0.306, respectively. In botanicals and bioagents neem seed kernel extract (5%) was effective in reducing the growth of bacteria with 0.446 OD at 96 h followed by *Pseudomonas flourescence* 1x10⁸ cell and *Bacillus subtilis* 1x10⁸ cell with 0.506 and 0.486 OD, respectively.

***Corresponding author:** Email: preranaabhang09@gmail.com **How to view point the article :** Abhang, P.B., Totawar, M.V. **and** Brahmane, P.R. (2015). *In vitro* efficacy of different botanicals, bioagents, chemicals against *Xanthomonas axonopodis* pv.*citri* by turbidiometrical method. *Internat. J. Plant Protec.*, **8**(2) : 241-244.

INTRODUCTION

Citrus is an important value as fruit crop. Present day citrus is delectable, juicy, and seedless is of great nutritional significance as well (Khan *et al.*, 1992). Additionally, it possesses enormous therapeutic qualities (Chaudhry *et al.*, 1992). Citrus is a member of Rutaceae family and grown in varying densities in countries with tropical or subtropical climates. Citrus plant is attacked by a number of diseases like citrus canker, gummosis, citrus decline, citrus tristeza virus, greening, etc. Citrus canker is caused by "Xanthomonas campestris pv. citri" that is probably the worst enemy to citrus plants (Sahi et al., 2007). Citrus canker disease is of regular occurrence on several citrus cultivars in varying degrees of incidence depending on the climatic conditions. The bacterium, Xanthomonas causes different symptoms ranging from pustules to necrotic lesions consisting of erumpent corky tissue surrounded by water soaked tissues and yellow halo on leaves, stems and fruits (Zekri *et al.*, 2005; Graham *et al.*, 2004; Das, 2003). *Xanthomonas axonopodis* pv. *citri* is a rod shaped, gram negative bacterium with single polar flagellum.

MATERIAL AND METHODS

The citrus canker diseased sample were collected from Akola district of Maharashtra state. The isolate of pathogen were obtained from infected leaves, twigs of acid lime showing typical symptoms of citrus canker. These isolate were isolated by tissue isolation method. A bacterial suspension of each specimen was then cultured on NA medium. Following incubation, colonies similar to *Xanthomonas* were maintained on NA medium at room temperature by adopting subsequent sub culturing at periodical, regular intervals. Three days old cultures were used for further studies.

Identification of the pathogen :

The identification of the pathogen involved in causing

of citrus canker in acid lime was determined by conducting studies on its morphological, biochemical, cultural and physiological features of the pathogen as per standard microbiological procedures.

Efficacy of botanicals, bioagents, chemicals against *Xac* was studied by turbidiometrically at 24hrs. intervals upto 96hrs by spectrophotometer at 620nm.

RESULTS AND DISCUSSION

Efficacy of chemicals, botanicals and bioagents was studied turbidiometrically. The results are given in Table 1 and Fig 1. At 96 hours of incubation least bacterial growth (0.232OD) was exhibited in copper oxychloride + streptomycin sulphate (0.2 % + 200ppm) followed with copper oxychloride +streptomycin sulphate (0.2 % + 100ppm) 0.266 OD statistically superior over all treatments. Similar findings are observed after 24h growth in T₆ and T₅ (0.303, 0.306OD), respectively. In botanicals and bioagent neem seed kernel extract (5%) was effective in reducing the growth of bacteria with

Table 1 : Efficacy of chemicals, botanicals and bioagents against Xanthomonas axonopodis pv. citri by Turbidometric method at 620nm					
Sr. No.	Treatments	Conc.	24 hrs	96Hrs	
T_1	Streptomycin sulphate	100 ppm	0.383	0.293	
T ₂	Streptomycin sulphate	200 ppm	0.373	0.286	
T ₃	Copper oxychloride	0.2%	0.364	0.273	
T_4	Copper oxychloride	0.3%	0.353	0.270	
T ₅	COC+Streptomycin sulphate	0.2%+100 ppm	0.306	0.266	
T ₆	COC+Streptomycin sulphate	0.2%+200 ppm	0.303	0.232	
T ₇	Kasugamycin	100 ppm	0.381	0.286	
T ₈	Kasugamycin	200 ppm	0.366	0.274	
T9	Agrimycin	100 ppm	0.373	0.293	
T ₁₀	Agrimycin	200 ppm	0.376	0.283	
T ₁₁	COC + Agrimycin	0.2%+100 ppm	0.351	0.292	
T ₁₂	COC + Agrimycin	0.2%+200 ppm	0.343	0.283	
T ₁₃	COC+ Kasugamycin	0.2%+100 ppm	0.323	0.276	
T ₁₄	COC+Kasugamycin	0.2%+200 ppm	0.344	0.276	
T ₁₅	Neem extract	1%	0.561	0.393	
T ₁₆	Neem extract	2.5%	0.576	0.403	
T ₁₇	Neem extract	5%	0.596	0.446	
T ₁₈	Pseudomonas fluorescens	1x10 ⁸ cfu /ml	0.615	0.506	
T ₁₉	Bacillus subtilis	$1 \mathrm{x} 10^8 \mathrm{cfu} /\mathrm{ml}$	0.636	0.486	
T ₂₀	Control		0.756	0.951	
	Blank		0.260	0.260	
	'F' test		Sig.	Sig.	
	S.E. (m) <u>+</u>		0.002	0.001	
	C.D. (P = 0.01)		0.007	0.003	



0.446 OD at 96 h followed by *Pseudomonas flourescence* 1x10⁸ cell and *Bacillus subtilis* 1x10⁸ cell with 0.506 and 0.486 OD, respectively. The present findings are in agreement with Sharma *et al.* (1980). *In vitro* evaluation of eight chemicals against *Xanthomonas vesicatoria* by paper disc and turbidometric method and described that, combination of streptocycline and copper sulphate was most effective in inhibiting the growth of pathogenic organism in turbidometric method. Das (2005) who assessed different of chemicals, antagonist and botanicals *in vitro* against *Xanthomonas axonopodis* pv. *citri* by measuring the growth turbidometrically at 620nm and reported maximum growth of inhibition after 96 hrs of incubation in COC (0.3%) + streptomycin sulphate (100ppm) *i.e.* 0.230 OD.

Conclusion :

Lower OD were found in chemical treatment *i.e.* copper oxychloride 0.2% + streptomycin sulphate 200ppm followed by concentration COC (0.2%)+Streptomycin sulphate (100ppm) by turbidometrical method *i.e.* 0.232 OD and 0.266 OD, respectively.

Acknowledgement :

The authors would like to thank Department of Plant Pathology, Post Graduate Institute, Dr. Panjabrao Deshmukh Krishi Vidyapeeth, Akola (M.S.) for providing necessary facilities for reaserch to the first author as a part of his M.Sc. (Plant Pathology) dissertation.

REFERENCES

Chaudhry, N.A., Khan, A.R. and Hameedullah (1992). Introduction of acclimatized exotic citrus.p: 15. Citrus fruit varieties at Horticultural Research Station, Sahiwal. Proc. 1st Int. Sem. Citriculture in Pakistan, Dec. 2–5. University of Agriculture Faisalabad–Pakistan.

Das, A.K. (2003). Citrus canker-A review. *J. Appl. Hort.*, **5**(1): 52-60.

Das, S. (2005). Variability among the isolates of *Xanthomonas axonopodis* pv. *citri*. M.Sc. Thesis, Dr. Panjabrao Deshmukh Krishi Vidyapeeth, Akola, M.S. (INDIA).

Deshmukh, V.V. (1994). Chemical control on bacterial leaf spot of greengram caused by *Xanthomonas campestries* pv.*vignardiatae*. M.Sc. Thesis, Dr. Panjabrao Deshmukh Krishi Vidyapeeth, Akola, M.S. (INDIA).

Graham, J.H, Gottwald, T.R., Cubero, J. and Achor, D.S. (2004). *Xanthomonas* axonopodis pv. *citri*: factors affecting successful eradication of citrus canker. *Mol. Plant Pathol.*, 5 (1):1-15.

Khan, M.M, Khan, M.A., Inam-ul Haq, M., Ahmad, R. and Aziz, I. (1992). Incidence of citrus canker caused by *X. campestris* pv. *citri* orchard in Faisalabad District. In: Proceed. 1st Inter. sem. citriculture in Pakistan. Dec. 2-5. University of Agriculture Faisalabad. pp. 311-314

Sahi, S.T., Ghazanfar, M.U., Afzal, M., Rasheed, A. and Habib, A. (2007). Incidence of citrus canker disease caused by *Xanthomonas campestris* pv. *citri* (Hasse) dows on kinnow (*Citrus reticulata*) and its chemotherapy. *Pak. J. Bot.*, **39**(4): 1319-1327.

Sharma, G.K., Czoppelt, Ch. and Rembold, H. (1980). Further evidence of growth dtsruption by neem seed fracrions. *Z. Angew. Ent.*, 90: 439-444.

Zekri, M., Chamberlain, H., Timmer, P., Roberts, P., and Muchove, R. (2005). Field identification of citrus canker symptoms and decontamination procedures. Uni. Florida. IFAS extension.

