

RESEARCH PAPER

In vitro antiproliferative and hepatoprotective activity studies of *Momordica cymbalaria*

S.J. PRASHANTH, D. SURESH*, V.H. POTTY AND P. SADANANDA MAIYA

Dr. P. Sadananda Maiya Centre for Food Science and Research, BENGALURE (KARNATAKA) INDIA

ABSTRACT

Natural products are well known to exhibit the antiproliferative and hepatoprotective activities. The present study aimed at assessing the potency of hydroalcoholic extracts of fruits, aerial parts and roots of *Momordica cymbalaria*. Antiproliferative activity of the extracts assessed using *in vitro* cell lines such as MCF-7, HepG2, HeLa, PC3, A549 and Vero cell lines. The ability of extracts to exert toxic insult on cancer cells has been the basis of anticancer activity. The extracts were evaluated for hepatoprotective activity by employing primary rat hepatocytes. In the case of HeLa cells, MCR was found to be most potentially toxic with average CTC_{50} of 67 $\mu\text{g/ml}$. MCR possesses more toxicity to PC3 cells than the others. Potential toxicity towards Vero cell lines was exhibited by MCR with average CTC_{50} value of 63 $\mu\text{g/ml}$. The results clearly demonstrate that the extract MCR exert potential anticancer activity. *In vitro* hepatoprotective activity of the plant extracts was studied by employing primary rat hepatocytes. Our results indicate that the drug Silymarin was found to exhibit 96 per cent protection against Paracetamol induced toxicity in Hep G2 cells at the tested concentration of 250 $\mu\text{g/ml}$. Among the parts of *Momordica cymbalaria*, MCA and MCR found to have slightly lesser activity compared to Silymarin. In paracetamol induced toxicity of primary rat hepatocytes, the drug Silymarin was found to exhibit 85.28 per cent protection against Paracetamol induced toxicity in Primary rat hepatocytes at the tested concentration of 250 $\mu\text{g/ml}$. MCA was found to exhibit comparatively similar protective power than silymarin.

Key Words : *Momordica cymbalaria*, Antiproliferative, Hepatoprotective, Hydroalcoholic extract

View point paper : Prashanth, S.J., Suresh, D., Potty, V.H. and Maiya, P. Sadananda (2014). *In vitro* antiproliferative and hepatoprotective activity studies of *Momordica cymbalaria*. *Asian Sci.*, 9 (1&2): 41-46.

Momordica cymbalaria is one of the species of cucurbitaceae family. The synonyms are *Momordica tuberosa* Roxb. or *Luffa tuberosa* Roxb. The plant is a perennial climber available only during the monsoon season and is found in the south Indian states of Andhra Pradesh, Karnataka, Madhya Pradesh, Maharashtra, and Tamil Nadu. The plant is a climbing annual or perennial herb with slender, scandent, branched, striate stem. The plant is traditionally used for the treatment of diabetes mellitus and also as an antiovarian agent. Bharathi *et al.* (2011) have reported the study on somatic chromosome number and detailed Karyotype analysis of six Indian *Momordica* species which includes *M. cymbalaria* also. The Somatic chromosome number of *M. cymbalaria* was reported to be $2n=18$ against its previous reports of $2n=16, 22$. The karyotype analysis reveals that it was possible to distinguish chromosomes of *M. cymbalaria* from other *Momordica* species and also between monoecious and dioecious taxa of the genus. A study on antiulcer activity of aqueous extracts of fruits of *Momordica cymbalaria* revealed that pretreatment

*** Author for correspondence**

D. Suresh, Department of Studies and Research in Chemistry, Tumkur University, TUMKUR (KARNATAKA) INDIA (Email: pbd_suresh@gmail.com)

with aqueous extract of *M. cymbalaria* fruits showed significant decrease in the total acidity and ulcer index and also showed improvements in all histopathological parameters in the *M. cymbalaria* treated group. It was also learnt that there is significant decrease in gastric lesion and NP-SH and gastric wall mucus concentration in the *M. cymbalaria* treated group in ulcer induced rats (Bharathi *et al.*, 2010). Studies on chemical composition and utilization of the wild edible vegetable Athalakkai (*Momordica tuberosa*) was shown that it has higher amounts of minerals such as calcium, potassium and sodium. Also it is rich in vitamin C than the bitter gourd (*Momordica charantia*) and also it is rich in high crude fibre (6.42g/100g) (Parvathi and Kumar, 2002). Its versatile utility as a nutritious vegetable, folk medicine and functional food ingredient provoked us to compile a comprehensive review of this multipurpose fruit on the distribution, nutritional attributes and phytochemicals composition and its medicinal properties. *M. cymbalaria* fruits were considered as tonic, stomachic, stimulant, laxative and alterative. The fruit is useful in treating gout, rheumatism and sub-acute cases of the spleen and liver diseases. It has also been shown to have hypoglycemic properties (antidiabetic) in animal as well as human studies. The fruit juice and leaf tea of *M. cymbalaria* is employed for diabetes, malaria, colic, sores and wounds, infections, worms and parasites, as an emmenagogue, and for measles, hepatitis and fevers. Fruit pulp, leaf juice and seeds possess anthelmintic activity. Root is astringent, abortifacient, aphrodisiac and also used to treat constipation, indigestion, diabetes, diarrhea and rheumatism. For the last few decades, the medicinal value of plants has been recognized. The extracts from many plants contain not only minerals and primary metabolites but also a diverse array of secondary metabolites, mostly with antioxidant properties. Plants belonging to *Momordica* species have been used as therapeutic agents for the treatment of diabetes mellitus. The other species of this genus, *Momordica charantia* and *Momordica foetida* have been reported to have hypoglycemic effects (Fernandes *et al.*, 2007 and Osinubi *et al.*, 2008). Pramod *et al.* (2008) have reported the antioxidant and hepatoprotective activity of ethanolic extract of tubers of *Momordica tuberosa* against CCl_4 induced liver injury in rats. CCl_4 is metabolized by CYP 450 enzyme system to trichloromethyl radical ($\text{CCl}_2\cdot$). This in turn reacts with molecular oxygen and gets converted to trichloromethyl peroxy radical. This radical forms covalent bonds with sulfhydryl group of several membrane molecules like reduced glutathione leading to this depletion and causes lipid peroxidation. The lipid peroxidation initiates a cascade of reactions leading to tissue necrosis. The antioxidant property of ethanolic extract of tubers of *Momordica tuberosa* prevented the formation of trichloromethyl peroxy radical there by reducing tissue damage. Pramod *et al.* (2011) have reported the hepatoprotective activity of 70 per cent ethanol

extract of tubers of *Momordica tuberosa* against thiocetamide (100 mg/kg of bodyweight) induced hepatic damage in albino rats.

The current study evaluates the hepatoprotective and antiproliferative activities of fruit, aerial parts and roots of *Momordica cymbalaria*.

RESEARCH METHODOLOGY

Chemicals :

3-(4, 5-dimethyl thiazol-2-yl)-5-diphenyltetrazolium bromide (MTT), Fetal Bovine Serum (FBS), Phosphate Buffered Saline (PBS), Dulbecco's Modified Eagle's Medium (DMEM) and Trypsin were obtained from Sigma Aldrich Co, St Louis, USA. EDTA, Glucose and antibiotics from Hi-Media Laboratories Ltd., Mumbai. Dimethyl Sulfoxide (DMSO) and Propanol from E. Merck Ltd., Mumbai, India.

3-(4, 5-dimethyl thiazol-2-yl)-5-diphenyltetrazolium bromide (MTT), Trypan blue, Fetal Bovine serum (FBS), Phosphate Buffered Saline (PBS), Dulbecco's Modified Eagle's Medium (DMEM) and Trypsin were obtained from Sigma Aldrich Co, St Louis, USA. EDTA, Glucose, Trichloroacetic acid (TCA), Acetic acid, Tris base and antibiotics from Hi-Media Laboratories Ltd., Mumbai. Dimethyl Sulfoxide (DMSO), Paracetamol and Propanol from E. Merck Ltd., Mumbai, India.

Cell lines and culture medium :

MCF-7, HepG2, HeLa, PC3, A549 and Vero cell lines. was cultured in DMEM supplemented with 10 per cent inactivated Fetal Bovine Serum (FBS), penicillin (100 IU/ml), streptomycin (100 µg/ml) and amphotericin B (5 µg/ml) in an humidified atmosphere of 5 per cent CO_2 at 37°C until confluent. The cells were dissociated with TPVG solution (0.2% trypsin, 0.02% EDTA, 0.05% glucose in PBS). The stock cultures were grown in 25 cm² culture flasks and all experiments were carried out in 96 microtitre plates (Tarsons India Pvt. Ltd., Kolkata, India).

HepG2 cell culture was procured from National Centre for Cell Sciences (NCCS), Pune, India. Stock cells of HEPG2 were cultured in DMEM supplemented with 10 per cent inactivated Fetal Bovine Serum (FBS), penicillin (100 IU/ml), streptomycin (100 µg/ml) and amphotericin B (5 µg/ml) in an humidified atmosphere of 5 per cent CO_2 at 37°C until confluent. The cells were dissociated with TPVG solution (0.2% trypsin, 0.02% EDTA, 0.05% glucose in PBS). The stock cultures were grown in 25 cm² culture flasks and all experiments were carried out in 96 microtitre plates (Tarsons India Pvt. Ltd., Kolkata, India).

Preparation of test solutions :

For cytotoxicity studies, each weighed test drugs were

separately dissolved in distilled DMSO and volume was made up with DMEM supplemented with 2 per cent inactivated FBS to obtain a stock solution of 1 mg/ml concentration and sterilized by filtration. Serial two fold dilutions were prepared from this for carrying out cytotoxic studies.

For Hepatoprotective studies, each weighed test drugs were separately dissolved in distilled DMSO and volume was made up with DMEM supplemented with 2 per cent inactivated FBS to obtain a stock solution of 1 mg/ml concentration and sterilized by filtration. Serial two fold dilutions were prepared from this for carrying out cytotoxic studies.

Determination of cell viability by MTT assay :

The ability of the cells to survive a toxic insult has been the basis of most cytotoxicity assays. This assay is based on the assumption that dead cells or their products do not reduce tetrazolium. The assay depends both on the number of cells present and on the mitochondrial activity per cell. The principle involved is the cleavage of tetrazolium salt 3-(4, 5 dimethyl thiazole-2-yl)-2, 5-diphenyl tetrazolium bromide (MTT) into a blue coloured product (formazan) by mitochondrial enzyme succinate dehydrogenase. The number of cells was found to be proportional to the extent of formazan production by the cells used (Francis and Rita, 1986).

The monolayer cell culture was trypsinized and the cell count was adjusted to 1.0×10^5 cells/ml using DMEM containing 10 per cent FBS. To each well of the 96 microtitre plate, 0.1 ml of the diluted cell suspension (approximately 10,000 cells) was added. After 24 h, when a partial monolayer was formed, the supernatant was flicked off, wash the monolayer once with medium and 100 μ l of different test concentrations of extracts were added on to the partial monolayer in microtitreplates. The plates were then incubated at 37°C for 3 days in 5 per cent CO₂ atmosphere and microscopic examination was carried out and observations were noted every 24 h interval. After 72 h, the drug solutions in the wells were discarded and 50 μ l of MTT in PBS was added to each well. The plates were gently shaken and incubated for 3 h at 37°C in 5 per cent CO₂ atmosphere. The supernatant was removed and 100 μ l of propanol was added and the plates were gently shaken to solubilize the formed formazan. The absorbance was measured using a microplate reader at a wavelength of 540 nm. The percentage growth inhibition was calculated using the following formula and concentration of test drug needed to inhibit cell growth by 50 per cent (CTC₅₀) values is generated from the dose-response curves for each cell line.

$$\% \text{ growth inhibition} = 100 - \left[\frac{\text{Mean OD of individual test group}}{\text{OD of control group}} \right] \times 100$$

Determination hepatoprotective activity in HepG2 cell line :

The monolayer cell culture was trypsinized and the cell count was adjusted to 1.0×10^5 cells/ml using DMEM medium containing 10 per cent FBS. To each well of the 96 well microtitre plate, 0.1 ml of the diluted cell suspension (approximately 10,000 cells) was added. After 24 h, when a partial monolayer was formed, the supernatant was flicked off, washed the monolayer once with medium. 50 μ l of DMEM with 2 x 100mm Paracetamol and 2 x 50 μ l of different non-toxic test concentrations of test drugs were added. The plates were then incubated at 37°C for 24 h in 5 per cent CO₂ atmosphere. After 24 h, the cell supernatants were discarded and 50 μ l of MTT in PBS was added to each well. The plates were gently shaken and incubated for 3 h at 37°C in 5 per cent CO₂ atmosphere. The supernatant was removed and 100 μ l of propanol was added and the plates were gently shaken to solubilize the formed formazan. The absorbance was measured using a microplate reader at a wavelength of 540 nm. The percentage cell viability was determined, based on which the percentage protection offered by test and standard drugs was calculated over the Paracetamol control.

Isolation of rat hepatocytes :

The primary rat hepatocyte culture was prepared as described earlier (Liu *et al.*, 2001). Briefly, abdomen of an adult rat, weighing 200-220 g will be opened with a midline incision. Portal cannula will be placed and the liver will be perfused with HEPES solution followed by collagenase type-iv solution. The liver capsule will be disrupted and the resulting cell suspension will be filtered and washed, cell viability will be measured by trypan blue exclusion technique. The cells will be suspended in Ham's F₁₂ medium with 20 per cent FBS and incubated at 37°C for further use. The handling and treatment of animals was as per Control and Supervision of Experiments on Animals (CPCSEA), Ministry of Environment and Forests, Government of India rules.

Hepatoprotective study in isolated rat hepatocytes :

Isolated primary rat hepatocytes were suspended fresh Ham's F₁₂ medium containing 20 per cent FBS and incubated at 37°C for 16h. Medium supernatant was removed carefully and primary hepatocytes were exposed to fresh medium containing toxicant along with/without test product at different doses, along with negative and standard controls. After 24h incubation period, cell supernatant will be aspirated and cell viability was determined by MTT assay.

RESULTS AND REMONSTRATION

In vitro anticancer activity of the extracts was performed by assessing cytotoxicity studies in 6 cell lines among which

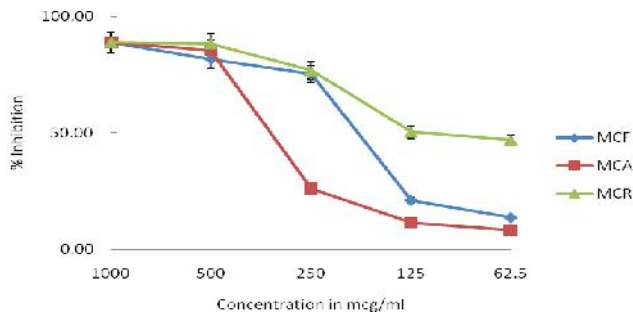
5 were cancerous cell lines and one was normal cell line. The cell lines used were MCF-7, HepG2, HeLa, PC3, A549 and Vero cell lines.

Among the extracts of MCA, MCF and MCR, MCF was found to be comparatively more toxic to HEP G2 cells. Its average CTC₅₀ was found to be 196 µg/ml compared to the extract MCR which is least toxic (>1000 µg/ml) (Table 1, Fig. 1). Among AGA, AGR and AGF, AGR was found to be potentially toxic at significantly low concentration of 95 µg/ml. However, the other two extracts are comparatively

Table 1 : Cytotoxicity of *Momordica cymbalaria* fruit, aerial parts and root extracts against HEP G2 cell line by MTT assay

Sr. No.	Test sample	CTC ₅₀ (µg/ml) ± SD
1.	MCF	196.67±15.28
2.	MCA	346.67±15.28
3.	MCR	>1000

Values are mean ± standard error of three replicates



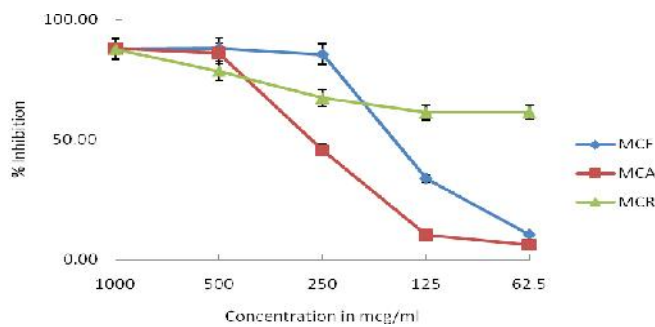
Values are mean ± standard error of three replicates

Fig. 1: Per cent inhibition of HEP G2 cell line by *Momordica cymbalaria* fruit, aerial parts and root extracts

Table 2 : Cytotoxicity of *Momordica cymbalaria* fruit, aerial parts and root extracts against MCF 7 cell line by MTT assay

Sr. No.	Test sample	CTC ₅₀ (µg/ml) ± SD
1.	MCF	156.67 ± 15.28
2.	MCA	283.33 ± 15.28
3.	MCR	>1000

Values are mean ± standard error of three replicates



Values are mean ± standard error of three replicates

Fig. 2: Per cent inhibition of MCF 7 cell line by *Momordica cymbalaria* fruit, aerial parts and root extracts

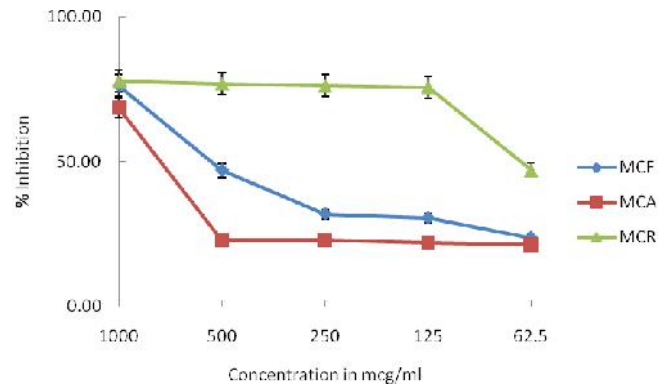
not very toxic (Table 2, Fig. 2).

MCF and MCA were found to be moderately toxic at 156 and 283 µg/ml, respectively (Table 3, Fig. 3). GXA possesses the CTC₅₀ of 810 µg/ml. However, other extracts were found to be not toxic up to the concentration of 1000 µg/ml to MCF 7 cells (Table 4, Fig. 4).

Table 3 : Cytotoxicity of *Momordica cymbalaria* fruit, aerial parts and root extracts against HeLa cell line by MTT assay

Sr. No.	Test sample	AVG CTC ₅₀ (µg/ml)±SD
1.	MCF	556.67±20.82
2.	MCA	800.00±17.32
3.	MCR	67.50±4.33

Values are mean ± standard error of three replicates



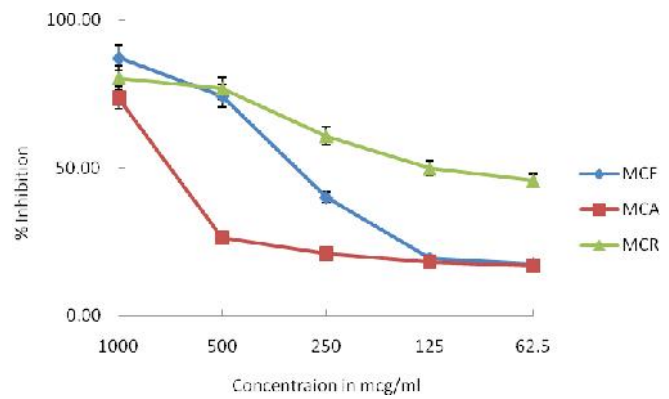
Values are mean ± standard error of three replicates

Fig. 3: Per cent inhibition of HeLa cell line by *Momordica cymbalaria* fruit, aerial parts and root extracts

Table 4 : Cytotoxicity of *Momordica cymbalaria* fruit, aerial parts and root extracts against PC 3 cell line by MTT assay

Sr. No.	Test sample	AVG CTC ₅₀ (µg/ml)±SD
1.	MCF	406.66±11.54
2.	MCA	750±10.00
3.	MCR	123.33±15.27

Values are mean ± standard error of three replicates



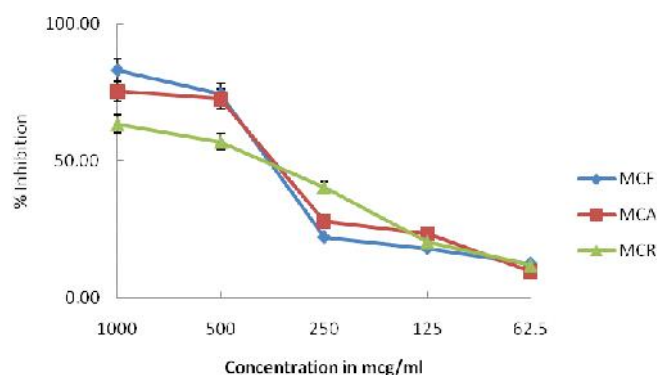
Values are mean ± standard error of three replicates

Fig. 4: Per cent inhibition of PC 3 cell line by *Momordica cymbalaria* fruit, aerial parts and root extracts

In the case of toxicity to HeLa cells, MCR was found to be most potentially toxic with average CTC_{50} of $67\mu\text{g/ml}$ (Table 5, Fig. 5). GXR was found to be toxic with average CTC_{50} of $303\mu\text{g/ml}$. This was followed by GXF and GXA with average CTC_{50} values of 303 and $456\mu\text{g/ml}$ (Table 6, Fig. 6). MCF, AGR, AGA and MCA were found to be

Sr. No.	Test sample	AVG CTC_{50} ($\mu\text{g/ml}$) \pm SD
1.	MCF	386.67 ± 5.77
2.	MCA	373.33 ± 5.77
3.	MCR	403.00 ± 7.64

Values are mean \pm standard error of three replicates

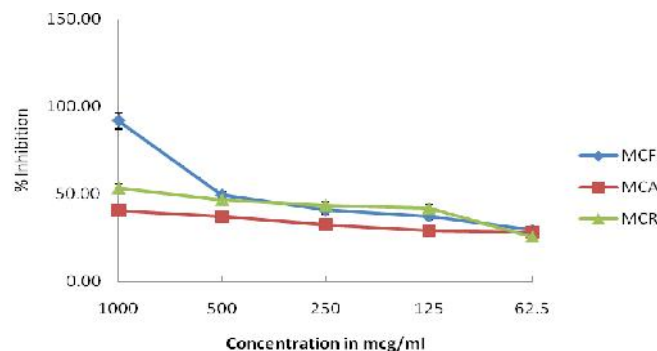


Values are mean \pm standard error of three replicates

Fig. 5 : Per cent inhibition of A 549 cell line by *Momordica cymbalaria* fruit, aerial parts and root extracts

Sr. No.	Test sample	AVG CTC_{50} ($\mu\text{g/ml}$) \pm SD
1.	MCF	513.33 ± 5.77
2.	MCA	>1000
3.	MCR	63.67 ± 11.93

Values are mean \pm standard error of three replicates



Values are mean \pm standard error of three replicates

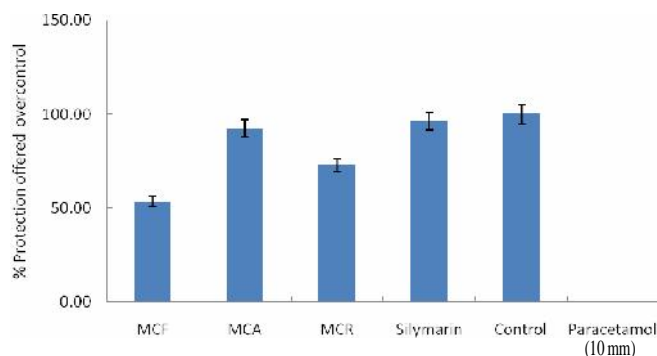
Fig. 6 : Per cent inhibition of Vero cell line by *Momordica cymbalaria* fruit, aerial parts and root extracts

possessing average CTC_{50} values of 556, 570, 660 and $800\mu\text{g/ml}$ respectively. AGF was found to be not toxic up to the concentration range of $1000\mu\text{g/ml}$ to HeLa cells (Table 7, Fig. 7).

Table 7 : Hepatoprotective activities of *Momordica cymbalaria* fruit, aerial parts and root extracts in HepG2 cell line

Sr. No.	Plant parts	Test Concn. in $\mu\text{g/ml}$	% Protection offered over control
1.	MCF	200	53.6 ± 0.11
2.	MCA	200	82.4 ± 0.04
3.	MCR	200	72.8 ± 0.04
4.	Silymarin	250	96.2 ± 0.02
5.	Cell control	-	100.0
6.	Paracetamol (10mm)	-	0

Values are mean \pm standard error of three replicates



Values are mean \pm standard error of three replicates

Fig. 7 : Percentage protection offered by *Momordica cymbalaria* fruit, aerial parts and roots extracts ($200\mu\text{g/ml}$) and standard drug Silymarin ($250\mu\text{g/ml}$) in HepG2 cell line

Among MCA, MCF and MCR extracts, MCR possesses more toxicity to PC_3 cells than the others (Table 8, Fig. 8).

Table 8 : Hepatoprotective activities of *Momordica cymbalaria* fruit, aerial parts and root extracts in primary rat hepatocytes

Sr. No.	Test drug	Test concn. in $\mu\text{g/ml}$	% Protection offered over control
1.	MCF	200	42.61 ± 0.04
2.	MCA	200	78.89 ± 0.02
3.	MCR	200	57.66 ± 0.03
4.	Silymarin	250	85.28 ± 0.03
5.	Cell control	-	100.0
6.	Paracetamol (10mm)	-	0

Values are mean \pm standard error of three replicates

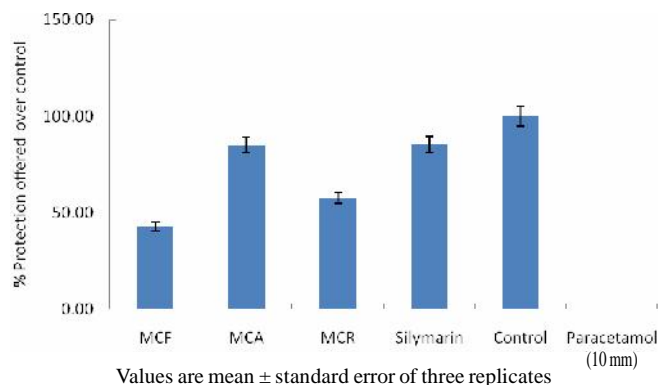


Fig. 8 : Percentage protection offered by *Momordica cymbalaria* fruit, aerial part and root extracts (200 μ g/ml) and standard drug Silymarin (250 μ g/ml) in primary hepatocytes

With regard to A549 cell lines, the extracts MCA, MCR and MCF found to possess moderate toxicity and all the three extracts shown to exhibit almost same toxicity (Table 5, Fig. 5).

Potential toxicity towards Vero cell lines was exhibited by MCR (Table 6, Fig. 6) and GXF with average CTC_{50} values of 63 and 93 μ g/ml, respectively (Table 7, Fig. 7). MCA and AGF were not toxic in the tested concentrations up to 1000 μ g/ml. Whereas GXR, AGR, MCF, AGA and GXA were having the CTC_{50} values of 341, 403, 513, 763 and 813 μ g/ml, respectively (Table 8, Fig. 8).

Hepatoprotective activity :

The drug Silymarin was found to exhibit 96.2 per cent protection against Paracetamol induced toxicity in Hep G2 cells at the tested concentration of 250mg/ml. It was found that MCA and MCR were found to have comparatively protective like Silymarin. These extract exhibited 82.4 and 72.8 per cent protection against Paracetamol induced toxicity in Hep G2 Cells at the concentration of 200 μ g/ml, respectively. However, other extract fruit (56.6% protection) did not exhibit much activity at the tested concentration of 200 μ g/ml (Table 7, Fig. 7).

The drug Silymarin was found to exhibit 85.28 per cent protection against paracetamol induced toxicity in Primary rat hepatocytes at the tested concentration of 250 μ g/ml. It was found that MCA and MCR were found to have comparatively protective like Silymarin. These extract exhibited 78.89 and 57.66 per cent protection against Paracetamol induced toxicity in primary rat hepatocytes at the concentration of 200 μ g/ml, respectively. However, other extract (fruit) did not exhibit much activity at the tested concentration of 200 μ g/ml (Table 8, Fig. 8).

Conclusion :

In the case of HeLa cells, MCR was found to be most

potentially toxic with average CTC_{50} of 67 μ g/ml. MCR possesses more toxicity to PC3 cells than the others. Potential toxicity towards Vero cell lines was exhibited by MCR with average CTC_{50} value of 63 μ g/ml.

Our results indicate that the drug Silymarin was found to exhibit 96 per cent protection against Paracetamol induced toxicity in Hep G2 cells at the tested concentration of 250 μ g/ml. Among the parts of *Momordica cymbalaria*, MCA and MCR found to have slightly lesser activity compared to Silymarin. In paracetamol induced toxicity of primary rat hepatocytes, the drug Silymarin was found to exhibit 85.28 per cent protection against Paracetamol induced toxicity in Primary rat hepatocytes at the tested concentration of 250 μ g/ml. MCA was found to exhibit comparatively similar protective power than silymarin. The results clearly demonstrate that the extract MCR exert potential anticancer and hepatoprotective activities.

REFERENCES

- Bharathi, D.P., Jegadeesan, M. and Kavimani, S.** (2010). Antiulcer activity of aqueous extract of fruits of *Momordica cymbalaria* Hook f. in Wistar rats. *Pharmacognosy Res.*, **2**(1): 58–61.
- Bharathi, L.K., Munshi, A.D., Chandrashekhara, S., Behera, T.K., Das, A.B., John, K.J. and Vishal Nath** (2011). Cytotaxonomical analysis of *Momordica* L. (Cucurbitaceae) species of Indian occurrence. *J. Geneti.*, **90**(1): 21–30.
- Fernandes, P.N., Lagishetty, C.V., Panda, V.S. and Naik, S.R.** (2007). An experimental evaluation of the antidiabetic and antilipidemic properties of a standardized *Momordica charantia* fruit extracts. *BMC Compl. Altern. Med.*, **7**: 29.
- Francis, D. and Rita, L.** (1986). Rapid colourimetric assay for cell growth and survival modifications to the tetrazolium dye procedure giving improved sensitivity and reliability. *J. Immunological Methods*, **89**: 271–277.
- Osinubi, A.A., Enye, L.A., Adesiyun, A.E. and Ajayi, G.O.** (2008). Comparative effects of three herbs and standard hypoglycaemic agents on blood glucose in normoglycaemic, hyperglycemic and alloxan-induced diabetic male rats. *Afr. J. Endocrinol. & Metab.*, **7**: 1.
- Pramod, K., Deval, R.G., Lakshmayya and Ramachandra, S.S.** (2008). Antioxidant and hepatoprotective activity of tubers of *Momordica tuberosa* Cogn. against CCl₄ induced liver injury in rats. *Indian J. Exp. Biol.*, **46**: 510–513.
- Pramod, K., Deval, R.G., Lakshmayya and Ramachandra, S.S.** (2011). Nephroprotective and nitric oxide scavenging activity of tubers of *Momordica tuberosa* in rats. *Avicenna J. Med. Biotech.*, **3**(2): 87–93.
- Parvathi, S. and Kumar, V.J.** (2002). Studies on chemical composition and utilization of wild edible vegetable Athalakkai (*Momordica tuberosa*). *Plant Foods Hum Nutr.*, **57**(3-4): 215–222.

Received : 14.07.2014; Revised : 06.11.2014; Accepted : 21.11.2014