

Management of anthracnose in soybean caused by *Colletotrichum truncatum*

■ S.L. KALE* AND B.G. BARHATE

Department of Plant Pathology and Agricultural Microbiology, Mahatma Phule Krishi Vidyapeeth, Rahuri, Ahmednagar (M.S.) INDIA

ARTICLE INFO

Received : 03.08.2016
Revised : 11.09.2016
Accepted : 25.09.2016

KEY WORDS :

Glycine max, Anthracnose,
Colletotrichum truncatum, Fungicides,
Bioagents

ABSTRACT

A study was conducted in the Department of Plant Pathology and Agricultural Microbiology, Mahatma Phule Krishi Vidyapeeth, Rahuri, Ahmednagar, Maharashtra during 2014 to 2015 to control *Colletotrichum truncatum* causing anthracnose or pod blight of soybean with fungicides and bioagents. All the fungicides and bioagents evaluated *in vitro* were found effective against *C. truncatum* and recorded significant inhibition of the test pathogen over untreated control. However, carbendazim was found most effective and recorded 0.66 mm mean colony diameter and significantly highest growth inhibition (99.26%) of the test pathogen. This was followed by mancozeb (98.88%), hexaconazole (84.44%), chlorothalonil (80.00%), propiconazole (78.15%) and difenconazole (32.22%). Out of the six bioagents evaluated *in vitro* *T. viride* and *T. harzianum* recorded significantly highest growth inhibition (78.88%) followed by *T. hamatum* (77.04%), yeast (40.37%), *P. fluorescens* (27.77%) and mehandi leaf extract (17.77%). *In vitro* physiological study of pathogen shows that *C. truncatum* grew well at 27°C temperature with 75 per cent relative humidity.

How to view point the article : Kale, S.L. and Barhate, B.G. (2016). Management of anthracnose in soybean caused by *Colletotrichum truncatum*. *Internat. J. Plant Protec.*, 9(2) : 583-588, DOI : 10.15740/HAS/IJPP/9.2/583-588.

*Corresponding author:

Email : savitakale417@gmail.com

INTRODUCTION

Soybean [*Glycine max* (L.) Merrill] is a species of legume native to East Asia. The plant is classed as an oilseed rather than a pulse. The genus *Glycine* is wild and divided into two subgenera, *Glycine* and *Soja*. Total yield of soybean in India is 12.07 q/ha. Production of soybean in India 14.68 million tonnes. Soybean growing major states in the country are Madhya Pradesh,

Maharashtra, Karnataka, Andhra Pradesh, Tamil Nadu, Rajasthan, Gujarat, Uttar Pradesh, Punjab and Haryana (Bhatnagar, 1997). About 80-85 per cent acreage of soybean in India is concentrated in Madhya Pradesh. More than 100 plant pathogens have been reported to affect soybean, but among them very few are economically important causing yield losses to the tons of 12-20 per cent (Mittal *et al.*, 1993). Among the most important diseases reported to cause economic losses to

the soybean anthracnose incited by *Colletotrichum truncatum* (Schw) Andrus and Moore causing yield losses of 16-100 per cent (Sinclair, 1992 and Anonymous, 1999). Among the major fungal diseases of soybean anthracnose (pod blight) caused by *Colletotrichum truncatum* (Schw) Andrus and Moore, has been reported as the major constraint in the successful cultivation of soybean (Khan and Sinclair, 1992 and Mittal *et al.*, 1993).

Objectives :

- To isolate and identify the pathogen causing anthracnose of soybean.
- To study the symptomatology.
- To prove the pathogenicity.
- *In vitro* chemical control of *Colletotrichum truncatum* by poison food technique.
- *In vitro* biological control of *Colletotrichum truncatum* by dual culture technique.
- To study effects of temperature and humidity on growth and sporulation of pathogen.

MATERIAL AND METHODS

In vitro evaluation of fungicides :

A total of six fungicide *viz.*, carbendazim (0.1%), chlorothalonil (0.2%), difenconazole (0.1%) hexaconazole (0.1%), propiconazole (0.1%) and mancozeb (0.1%) were evaluated *in vitro* applying "poison food technique". The requisite quantity of each fungicide was calculated and thoroughly mixed with autoclaved and cooled (40-45°C) PDA in conical flasks to obtain desired concentrations. An appropriate quantity of each fungicide was incorporated separately in the molten and cooled PDA medium in conical flask (250 ml capacity) to get desired concentrations of each fungicide. The fungicide amended PDA was then poured (15 – 20 ml/plates) in sterilized Petri plates (90 mm dia.) under aseptic conditions. Three plates / treatments were maintained and each treatment was replicated for three times. On solidification of PDA in Petri plates, all treatment plates were inoculated / seeded aseptically by placing in the centre with 5.0 mm uniform mycelial disc obtained from 7 days old culture of *C. truncatum* multiplied on agar plates. Petri plates containing plain PDA without any fungicide were inoculated with 5.0 mm disc of the test pathogen and maintained as suitable untreated control. All the treatment (inoculated) and control Petri plates were then incubated at 24 +2°C in

BOD incubator till the control plates were fully covered with mycelial growth of the test pathogen.

In vitro evaluation of bioagents :

Five bioagents *Trichoderma viride*, *T. harzianum*, *T. hamatum* and *P. fluorescens* and yeast were also evaluated *in-vitro* applying dual culture technique and one botanical mehandi leaf extract evaluated *in-vitro* applying poison food technique.

Healthy and disease free fresh leaf sample of mehandi plant species were brought to the laboratory and washed with sterile distilled water and then chopped into small bits with sterilized sharp knife. Leaf sample was then separately grind and homogenized in mechanical grinder with equal quantity of sterile distilled water (1:1, w:v). The homogenate obtained was then strained through double layered muslin cloth and filtrate collected was then filtered through Whatman number 1 filter paper using volumetric flasks (50 ml capacity). The clear leaf extracts obtained formed the stock solution of 100 per cent. An appropriate quantity of mehandi extract was incorporated separately in the molten and cooled PDA medium in conical flask (250 ml capacity) to get desired concentrations (10%) of extract and autoclaved at 15 Lbs pressure for 15 min. The leaf extracts added PDA was then poured (15 – 20 ml/plates) in sterilized Petri plates (90 mm dia.) under aseptic conditions. Disc (5 mm) of *Colletotrichum truncatum* was placed at the center of one corner of different Petri plates containing solidified PDA medium and disc of *Trichoderma viride*, *Trichoderma harzianum*, *Trichoderma hamatum* was placed at other corner of different Petri plates. The disc of each bio-agent kept with *Colletotrichum truncatum* disc in three Petri plates. A loopful of 24 hour old culture of *Pseudomonas fluorescens* and *Yeast* was inoculated at 2 cm just opposite to the pathogen on each plate (Dennis and Webster, 1971). Observations on radial mycelial growth of *C. truncatum* were recorded in each treatment and per cent growth inhibition of the test pathogen over control was worked out (Vincent, 1927) as follows.

$$\text{Per cent inhibition } I = \frac{C - T}{C} \times 100$$

where,

C = Growth of test fungus (mm) in control plate,

T = Growth of test fungus (mm) in treatment plates

In vitro evaluation of effect of temperature and humidity on growth and sporulation of pathogen :**Mycelial growth :**

Experiment of effect of temperature and humidity on growth and sporulation of *C. truncatum* were conducted *in vitro* on potato dextrose agar medium. The Petri plate were inoculated with fresh culture of effective *C. truncatum* isolates and incubated at 5, 20, 27, 34, 40, 45°C temperature with humidity 95, 85, 75, 65, 55, 50 per cent, respectively. Each treatment replicated for four times. Growth of each isolates in each Petri plate was recorded at every alternate day upto 7th day after inoculation.

Sporulation :

The cavity slides containing 0.1 ml of conidial suspension (10⁶ conidia/ml) were placed in Petri plates containing moist filter papers at the bottom were incubated for 24 hrs. at the different temperature and humidity as mentioned earlier. Each treatment was replicated for four times and in each replication 100 conidia were counted for the germination.

All Observations on radial mycelial growth and

sporulation of *C. truncatum* recorded in each treatment and replication and mean colony diameter is calculated.

RESULTS AND DISCUSSION

The findings of the present study as well as relevant discussion have been presented under the following heads:

In vitro evaluation of fungicides :

Results (Table 1) indicated that, all the fungicide tested significantly inhibited the mycelial growth of *C. truncatum* over untreated control Among the six fungicides, tested Carbendazim recorded least mean colony diameter (0.66 mm) and highest inhibition (99.26 %) of mycelial growth of the test pathogen over untreated control followed by mancozeb (98.44%), hexaconazole (84.44%), chlorothalonil (80.00%), propiconazole (78.15 %) and difenconazole (32.22%) which recorded mean colony diameter of test pathogen 1.00, 14.00, 18.00, 19.66 and 61.00 mm, respectively. Fungicide Difenconazole was found comparatively least effective than other fungicides. Carbendazim was reported

Table 1 : In vitro evaluation of fungicides on radial growth of *C. truncatum*

Treat. No.	Treatments	Concentration (%) used	Mean colony diameter (mm)*	Inhibition(%)
T ₁	Carbendazim	0.1%	0.66	99.26
T ₂	Chlorothalonil	0.2%	18.00	80.00
T ₃	Difenconazole	0.1%	61.00	32.22
T ₄	Hexaconazole	0.1%	14.00	84.44
T ₅	Propiconazole	0.1%	19.66	78.15
T ₆	Mancozeb	0.1%	1.00	98.88
T ₇	Control	-	90.00	0.00
	S.E.±	1.17		
	C.D. (P=0.05)	3.63		

* Mean of three replications

Table 2 : In vitro evaluation of effect of bio-agents on radial growth of *C. truncatum*

Sr. No.	Biological agents	Mean colony diameter (mm)*	Inhibition(%)
T ₁	<i>Trichoderma viride</i>	19.00	78.88
T ₂	<i>Trichoderma harzianum</i>	19.00	78.88
T ₃	<i>Trichoderma hamatum</i>	20.66	77.04
T ₄	<i>Pseudomonas fluorescens</i>	65.00	27.77
T ₅	Yeast (<i>S. cerevisiae</i>)	53.66	40.37
T ₆	Mehandileaf extract (10%)	74.00	17.77
T ₇	Control	90.00	00.00
	S.E.±	1.28	
	C.D. (P=0.05)	3.94	

* Mean of three replications

inhibitory to the *C. lindemuthianum* causing anthracnose of French bean (Chakraborty and Shyam, 1988), *C. truncatum* causing anthracnose of soybean (Ghawde *et al.*, 1996), *C. graminicola* (Singh and Dwivedi, 2002), *C. capsici* causing blight of bitter melon (Dubey and Ekka, 2003), *C. gloeosporioides* causing anthracnose of mango (Kumar *et al.*, 2003).

In vitro evaluation of bioagents :

Results (Table 2) revealed that among the six bioagents, *T. viride* and *T. harzianum* was found most effective and recorded 19.00 mm mean colony diameter and 78.88 per cent inhibition of the test pathogen followed by *T. hamatum* (77.045%), yeast (40.37%), *P. fluorescens* (27.77%) and *Mehandi* leaf extract (17.77%) with mean colony diameter 20.66, 53.66, 65, 74 mm, respectively. The bioagents, *T. viride* and *T. Harzianum* were reported as effective antagonists against *Colletotrichum* species by several workers (Barros *et al.*, 1995; Jayalakshmi *et al.*, 1998; Ingle *et al.*, 2002; Devis *et al.*, 2003; Raheja and Thakur, 2002;



Fig. 1 : *In vitro* evaluation of fungicides against *C. truncatum*



Fig. 2 : *In vitro* evaluation of bioagents against *C. truncatum*

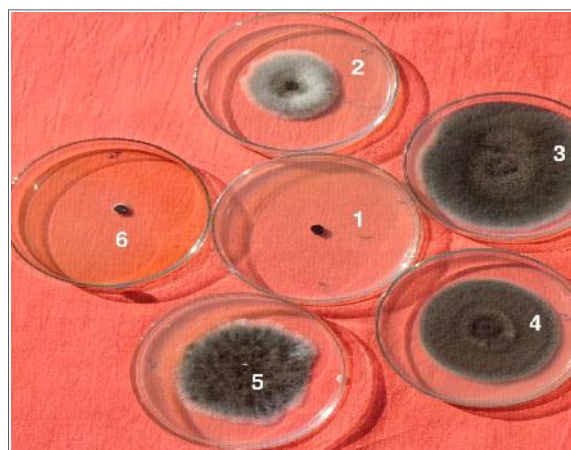


Fig. 3 : Effect of temperature and humidity on *C. truncatum*

Rao and Narayana, 2005 and Kaur *et al.*, 2006).

***In vitro* evaluation of effect of temperature and humidity on *C. truncatum* :**

The results presented in Table 3 reveals that the

Table 3 : <i>In vitro</i> evaluation of effect of temperature and humidity on radial growth and sporulation of <i>C. truncatum</i>				
Sr. No.	Temperature (°C)	Humidity (%)	Mean colony diameter (mm)*	Sporulation
1.	5	95	00.00	-
2.	15	85	45.25	+
3.	27	75	84.00	+++
4.	34	65	74.25	++
5.	40	55	63.75	+
6.	45	50	00.00	-
	S.E.±	0.39		
	C.D. (P=0.05)	1.12		

* Mean of four replications

growth of pathogen was maximum at 27°C temperature with humidity 75 per cent. Below 5°C and above 40°C were inhibitory to the test pathogen. There was no growth observed at 5°C and 45°C temperature. As regards to sporulation the good sporulation was observed at 27°C temperature with humidity 75 per cent and moderate sporulation at 34°C with humidity 65 per cent. Scanty sporulation was recorded at 15°C and 40°C temperature with humidity 85 per cent and 55 per cent, respectively. There was no sporulation at 5°C and 45°C temperature. Results are more or less similar with several workers Doornik (1982); Leng *et al.* (1984); Muniz *et al.* (1998) and Prabhakar *et al.* (2003).

Conclusion :

The fungal pathogen *Colletotrichum truncatum* caused anthracnose of soybean. From this experiment it was concluded that among the six fungicides tested *in vitro*. Carbendazim (0.1%) found most effective in inhibiting growth of test pathogen, followed by mancozeb (0.1%), hexaconazole (0.1%), chlorothalonil (0.2%) propiconazole (0.1%) and difenconazole (0.1%) in order of merit *in vitro*. In biological control the *Trichoderma viride* and *Trichoderma harzianum* found most effective in inhibiting growth of test pathogen followed by *Trichoderma hamatum*, yeast, *Pseudomonas fluorescens* and mehandi leaf extract in order of merit *in vitro*. Test pathogen grew well with maximum sporulation at optimum temperature of 27°C with 75 per cent relative humidity.

Acknowledgement :

The authors are grateful to the Associate Dean of Post Graduate Institute, Mahatma Phule Agricultural University, Rahuri and Head Department of Plant Pathology and Agricultural Microbiology Post Graduate Institute, Mahatma Phule Agricultural University, Rahuri for providing necessary facilities for conducting the research.

REFERENCES

- Barros, S.T., Oliveira, N.T. and Bastos, S.T.G. (1995). *Trichoderma* spp. in the biological control of *Colletotrichum lindemuthianum* causal agent of bean (*Phaseolus vulgaris* L.) anthracnose. *Bull. Mycologia*, **10** (1 / 2) : 5-11.
- Bhatnagar, P.S. (1997). An overview of soybean in India strategies for augmenting productivity and production with special reference to combating soybean rust, In: *Global focus on soybean and crop outlook for India*. Soybean Kharif, 1997-98. SOPA, Indore.
- Chakarabarty, P.K. and Shyam, K.R. (1988). Evaluation of systemic fungitoxicants against *C. lindemuthianum*, the incitant of French bean anthracnose. *Indian Phytopath.*, **6** (1) 67-70.
- Dennis, C. and Webster, J. (1971). Antagonistic properties of species-group of *Trichoderma* and hyphal interactions. *Trans. British Mycol. Soc.*, **57** : 363-369.
- Devis, D., Beena, S., Salley, K. and Mathew (2003). *In-vitro* evaluation of chemicals, plant extracts and microbial antagonists against *Cercospora cocciniae*, the incitant of leaf spot disease of Ivy gourd. *J. Mycol. Pl. Pathol.*, **33** (3) : 474.
- Doornik, A.W. (1982). Studies on anemone leaf curl diseases caused by *C.gloeosporioides*. Annual rept. Lab. for flower bud research, Lsse.Netherlands.pp.192. (Abstr.RPP. 1984, **64** (5): 172-173.
- Dubey, S.C. and Ekka, S. (2003). Integrated chemical management of *Colletotrichum* blight of bitter gourd. *Indian Phytopath.*, **56** (3) : 348.
- Ghawde, R.S., Gaikwad, S.J. and Borkar, S.L. (1996). Evaluation of fungicides and screening of varieties against pod blight of soybean caused by *C.truncatum* (Schw). *Andrus & Moore. J. Soils Crops*, **6** (1) : 97-99.
- Ingle, Y.V., Ingle, R.W. and Jamdade, S.R. (2002). *In-vitro* studies on leaf spot of turmeric caused by *C. capsici* (Syd.). *Pl. Dis. Res.*, **17** (1) : 217.
- Jeyalakshmi, C., Durairaj, P., Seetharaman, K. and Sivaprakasam, K. (1998). Biocontrol of fruit rot and die-back of chilli using antagonistic microorganisms. *Indian Phytopath.*, **51** (2) : 180- 183.
- Kaur, M., Sharma, O.P. and Sharma, P.N. (2006). *In vitro* effect of *Trichoderma* species on *C. capsici* causing fruit rot of chilli. *Indian Phytopath.*, **59**(2) : 243-245.
- Khan, M. and Sinclair, J.B. (1992). Pathogenicity of Sclerotia and non-sclerotiaforming isolates of *C. truncatum* on soybean plants and roots. *Phytopathology*, **82** (3) : 314-319. <http://dx.doi.org/10.1094/Phyto-82-314>.
- Kumar, P.M.K., Nargund, V.B., Khan, A.N.A. and Venkataravanappa, V. (2003). *In vitro* evaluation of fungicides and botanicals against *C. gloeosporioides* and *Alternaria alternata* causing post harvest diseases in mango. *Indian Phytopath.*, **56** (3) : 343.
- Leng, H.G., Liq, X.C. and Sen, I.C. (1984). On the sporulation of secondary conidia *C. gloeosporioides*. *Acta Phytopathologica*, **14** (2) : 95100 (Abstr.RPP.1985(3): pp-91).

Mittal, R.K., Prakash, V. and Koranne, K.D. (1993). Package of practices for the cultivation of pulses in the hills of the Uttar Pradesh. *Indian Fmg.*, **42** (10) : 3-5.

Muniz, M.F.S., Lemos, E.E.P., Rodrigues, C.J., Bessa, A.M.S., Topper, C.P., Caligari, P.D.S., Kullaya, A.K., Shomari, S.H., Kasuga, L.J., Masawe, P.A.L. and Mpunami, A.A. (1998). Characterization of *C.gloeosporioides* (Penz.) Sac. Isolates and resistance of cashew to pathogen. Proc. Int. Cashew and coconut conference, trees for life –the key development, Dares Salaam, Tanzania. 17-21 Feb. 1997: 249-253.

Prabhakar, K., Muthulakshmi, P., Raguchandar, T. and Parthiban, V.K. (2003). Influence of temperature and relative humidity on anthracnose pathogen growth and disease development in mango under *in vitro*. *Madras Agric. J.*, **90** (7-9) : 495-501.

Raheja, S. and Thakore, B.B.L. (2002). Effect of the physical factors, plant extracts and bioagents on *Colletotrichum gloeosporioides*, causing anthracnose in Yam. *J. Mycol. Pl. Pathol.*, **32** (2) : 282.

Rao, C.H. and Narayana, Y.D. (2005). *In vitro* evaluation of fungicides, plant extracts and biocontrol agents against *C.*

dematum (Pers. Ex. Fr.) Grove the causal organism of chickpea (*Cicerarietenum* L.) blight. Innational symposium on crop disease management in dry land Agril. and 57th Annual meeting IPS, Jan. 12-14, 2005, Marathwada Agricultural University, Parbhani (M.S.) INDIA.

Sinclair, J.B. (1992). Discoloration of soybean seeds an indicator of quality. *Pl. Dis.*, **76** (11) : 1087-1091. <http://dx.doi.org/10.1094/PD-76-1087>

Singh, D.P. and Dwivedi, R.R. (2002). Effect of fungicides and antibiotics on spores germination of *C. graminicola*. *Indian Phytopath.*, **55** (3) : 384.

Vincent, J.M. (1927). Distortion of fungal hyphae in presence of certain inhibitors. *Nature*. **159**, 850. <http://dx.doi.org/10.1038/159850b0>.

■ WEBLIOGRAPHY

Anonymous (1999). Development of selection and clonal propagation techniques for multiplication of elite yield and anthracnose tolerant cashew (*Anacardium occidentale* L.). Summary reports of European Commission supported STD-3 projects (1992-1995), Published by CTA. [online <http://interships.cta.int/pubs/std/vol2/pdf/221/pdf>] 111-116.

9th
Year
★★★★★ of Excellence ★★★★★